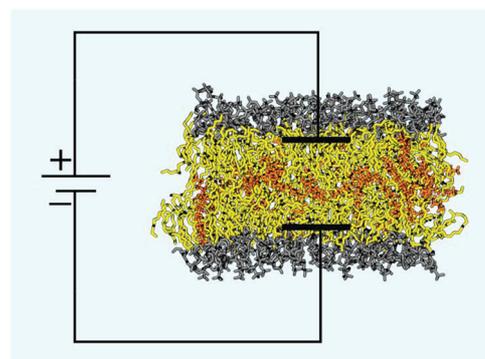


Accurate Estimation of Membrane Capacitance from Atomistic Molecular Dynamics Simulations of Zwitterionic Lipid Bilayers

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ABSTRACT: Lipid membranes are indispensable to life, and they regulate countless cellular processes. To investigate the properties of membranes under controlled conditions, numerous reconstitution methods have been developed over the last few decades. Several of these methods result in the formation of lipid bilayers containing residual hydrophobic molecules between the two monolayers. These contaminants might alter membrane properties, including bilayer thickness, that is usually inferred from measurements of membrane capacitance assuming a simple slab model. However, recent measurements on solvent-free bilayers raised significant questions on the reliability of this approach. To reconcile the observed discrepancies, we developed a protocol to predict membrane capacitance from the dielectric profile of lipid bilayers computed from molecular dynamics simulations. Our methodology shows excellent agreement against available data on solvent-free noncharged bilayers, and it confirms that the uniform slab model is a reliable approximation from which to infer membrane capacitance. We find that the effective electrical thickness contributing to membrane capacitance is different from the hydrophobic thickness inferred from X-ray scattering form factors. We apply our model to estimate the concentration of residual solvent in reconstituted systems, and we propose that our protocol could be used to infer membrane properties in the presence of hydrophobic solvents.



■ INTRODUCTION

Cellular membranes play a pivotal role in the modulation of numerous biological processes.^{1–4} Therefore, a thorough quantitative understanding of their physical and chemical properties is crucial to understanding and predicting how membranes might selectively modulate cellular mechanisms.

To simplify the extreme chemical complexity of cellular membranes^{5,6} and to systematically and quantitatively investigate the influence of membrane properties on biologically relevant processes, various approaches to reconstituting model cellular membranes *in vitro* have been proposed in the last few decades.^{7–14} The ultimate goal of these approaches is to reconstitute systems that mimic biological behavior, for example, by membrane embedding functionally active proteins while being able to concomitantly measure membrane properties.

From a practical point of view, however, the precise measurement of membrane properties is not always a straightforward task. Membrane thickness, for example, has been shown to play a major role in regulating membrane protein localization¹⁵ or lateral diffusion,¹⁶ yet the unambiguous determination of membrane thickness in reconstituted systems is still an open challenge. This is particularly

problematic since several reconstitution approaches are not entirely solvent-free but rather contain residual oil in the bilayer,^{17–19} potentially altering the membrane thickness to nonphysiological levels.

The gold standard for the determination of the lipid bilayers' thickness is based on the estimation of the electron density profiles from X-ray form factors.²⁰ While extremely accurate, this methodology is incompatible with the concomitant investigation of protein activity, and it can thus be performed exclusively for protein-free lipid bilayers. As a practical alternative, the determination of membrane thickness based on capacitance measurements has been used for a number of reliable membrane reconstitution methods, such as black lipid membranes (BLMs),¹⁴ droplet interface bilayers (DIBs),¹¹ large-area model biomembranes (LAMBs),⁷ and recently giant unilamellar vesicles (GUVs).²¹ This approach takes advantage

of the peculiar organization of a lipid bilayer, where a thin layer of low dielectric constant ($\epsilon_h \approx 2$), the hydrophobic membrane core, separates two aqueous regions of high dielectric constant ($\epsilon_{\text{wat}} \approx 80$). As a consequence, if we assume that the relative dielectric of the entire system can be described with a uniform slab model (i.e., with constant (high) permittivity outside the bilayer and constant (low) permittivity inside the bilayer), then the bilayer capacitance is directly proportional to its relative dielectric constant and inversely proportional to its hydrophobic core thickness, thus allowing an estimation of the thickness from capacitance measurements.²²

While this approach has potential, it relies on several untested assumptions: first, that a slab model faithfully describes the dielectric profile of a lipid bilayer; second, that the contribution of the phospholipid head groups ($\epsilon_p \approx 10\text{--}30$)^{23,24} to the total capacitance is negligible; third, that the thickness of the bilayer hydrophobic core is precisely known; and fourth, that the relative dielectric of the membrane interior can be considered to be identical to that of alkanes, with values of around 2.0–2.1.^{25,26}

Recently, Garten et al.²⁷ managed to measure for the first time the capacitance of solvent-free lipid bilayers using GUVs. Using a fitting procedure based on a uniform slab model, they found that their capacitance measurements led to a value of 2.06 for the dielectric constant of the bilayer interior ϵ_h and observed a good correlation between membrane capacitance and bilayer thickness measured using X-ray form factors. In order to achieve a good correlation, however, they had to subtract from the experimentally derived headgroup-to-headgroup thickness a fixed value of 0.78 nm for the interfacial thickness (interpreted as the thickness of the polar head groups), which is different from the experimental value of 0.55 nm.²⁸

In an independent study, Beltramo et al.²⁹ recently noted that in order to convert capacitance measurements to membrane thickness values comparable to those obtained using small-angle X-ray scattering (SAXS), a value of 2.5 for the hydrophobic core dielectric constant provided the best fit.

These discrepancies raise important questions about our theoretical understanding of the electrostatic behavior inside lipid bilayers and our ability to predictively infer membrane thicknesses from experimental measurements of membrane capacitance. This is particularly relevant in the case of oil-enriched lipid bilayers, such as BLMs,¹⁴ DIBs,¹¹ or LAMBs,⁷ that are extremely convenient to use in performing experiments in the presence of proteins but for which X-ray measurements cannot be directly performed.

In this article, we provide a protocol to directly determine membrane capacitance from molecular dynamics (MD) simulations of uncharged lipid bilayers, and our results show very good agreement with capacitance measurements on solvent-free lipid bilayers. We show that the “textbook” slab model is a good approximation and that the “electrical” thickness of membranes differs from experimentally determined values of hydrophobic thickness using X-ray spectroscopy. We conclude by showing how the approach we present here can be used to infer the concentration of hydrophobic solvents within a bilayer, which cannot be measured otherwise.

METHODS

Molecular Dynamics Simulations. All MD simulations were performed under periodic boundary conditions using the GROMACS software.^{30,31} Suitable starting configurations for

the various solvated lipid bilayers employed in our classical MD studies (Tables 1 and 2) were generated using the CHARMM-GUI membrane builder.³² The all-atom CHARMM36m force field³³ was used to describe lipids, and the TIP3P³⁴ model was employed for water molecules because this specific water model was used in the development of the CHARMM36 force field for lipids.³³ Long-range electrostatic interactions were taken into account by means of the particle mesh Ewald (PME)³⁵ algorithm with a Fourier grid space of 0.12 nm and a real space cutoff of 1.2 nm. The van der Waals interactions were truncated using the same cutoff value, and a standard smoothing function for the tail region (1.0–1.2 nm) was employed. The bonds involving hydrogen atoms were constrained using the LINCS³⁶ and SETTLE³⁷ algorithms, thus allowing the use of an integration time step of 2 fs. The temperature (298.15 K) and pressure (1 atm) were controlled by coupling the systems to a Nosé–Hoover chain thermostat³⁸ with a time constant of 1 ps⁻¹ and a Parrinello–Rahman barostat³⁹ with a coupling constant of 5 ps⁻¹, respectively. After an equilibration period of about 50 ns, all of the systems (containing about 128 lipids and about 6000 water molecules) were run for at least 400 ns to collect statistics. We simulated bilayer systems for which capacitance values were measured by Garten et al.²⁷ (shown in Table 2), and the ion concentration was assumed to be zero.

To perform simulations containing hexadecane and squalene, topologies for the two molecules were obtained using available resources online. For hexadecane, the parameters were obtained using the “ligand reader and modeler” tool in CHARMM-GUI. For squalene, topologies compatible with the CHARMM 27 force field were obtained from data available at the online repository.⁴⁰ The parameters for a new atom-type (CEL0) were integrated into the CHARMM36 force field, and the writemxtp function of TopoTools for VMD was used to convert CHARMM topologies to a format compatible with GROMACS.⁴¹

To build these systems, boxes consisting of about 7000 water molecules, 128 DOPC molecules, and a variable number of squalene and hexadecane molecules were initially prepared (Table 1). The two DOPC monolayers were displaced along the z axis in order to allow the insertion of the oil molecules and avoid steric clashes due to overlapping molecules. The oil molecules were subsequently placed randomly between the two monolayers. The systems were equilibrated for 375 ps while slowly removing constraints using the *Membrane Builder* six-step process.³² After equilibration for about 50 ns, production simulations were run for 400 ns using the same parameters described above for systems not containing oil.

Production runs were divided in two parts of 200 ns each and analyzed separately. The values reported are averages and standard deviations from the two portions.

Table 1. Fraction of Oil Molecules Used in Simulations with Squalene and Hexadecane in DOPC Bilayer Systems

oil fraction	no. of oil molecules
0.065	9 (squalene)
0.10	15 (squalene)
0.19	30 (squalene/hexadecane)
0.32	60 (squalene/hexadecane)
0.43	98 (hexadecane)
0.5	128 (hexadecane)

Membrane Capacitance. Membrane-specific capacitance values were computed assuming the slab model. The justification for the use of this model is presented in the [Results and Discussion](#). According to this “textbook” model, lipid bilayers can be approximated as uniform parallel plate capacitors embedded in water ([Figure 1](#)) because they consist

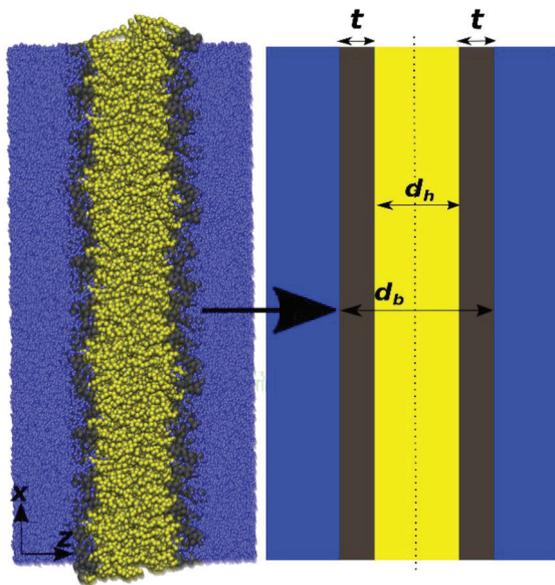


Figure 1. Membrane simplified as a uniform slab model along the line of symmetry. (Left) Atomistic model. (Right) Simplified model. The definitions of the various thicknesses used in the article are shown here: hydrophobic atoms, yellow; polar headgroups, gray; and water molecules, blue.

of low-dielectric hydrocarbon chains connected to a very high dielectric continuum through the small region where the polar headgroups are located and whose contribution to the capacitance is negligible.⁴² As a result, the membrane effective specific capacitance can be calculated from

$$\frac{1}{C} = \frac{1}{C_h} + \frac{1}{C_p} \approx \frac{1}{C_h} \quad (1)$$

$$C \approx C_h = \frac{\epsilon_0 \epsilon_h}{d_b - 2t} = \frac{\epsilon_0 \epsilon_h}{d_h} \quad (2)$$

where C is the effective specific capacitance of the membrane, C_h and C_p are the specific capacitances of the hydrophobic and polar regions, respectively, d_b is the bilayer thickness, d_h is the thickness of the hydrophobic core, t is the interfacial thickness, ϵ_0 is the vacuum permittivity, and ϵ_h is the dielectric constant of the hydrophobic region.

The parameters of this model are therefore the dielectric permittivity of the hydrophobic region and the widths of the hydrophobic and polar regions. A general and detailed theory regarding this model is available for inorganic substrates next to solvents, and this topic has been thoroughly discussed in the literature.^{43–45}

Physical Membrane Thickness from MD Simulations.

From the MD simulations, we defined two different physical bilayer thicknesses. The first one (d_{p-p}) was defined as the ensemble average of the distance between the phosphorus atoms belonging to both upper and lower lipid leaflets. The second one was defined from the water density profile along

the direction perpendicular to the membrane plane (z axis): from the first derivative of this profile, we obtained the bilayer thickness (d_b) as the separation between its extrema, as shown in [Figure 2](#). Similarly, the interfacial thickness t was also extracted as the distance between the neighboring extrema of the first and second derivatives ([Figure 2](#)).

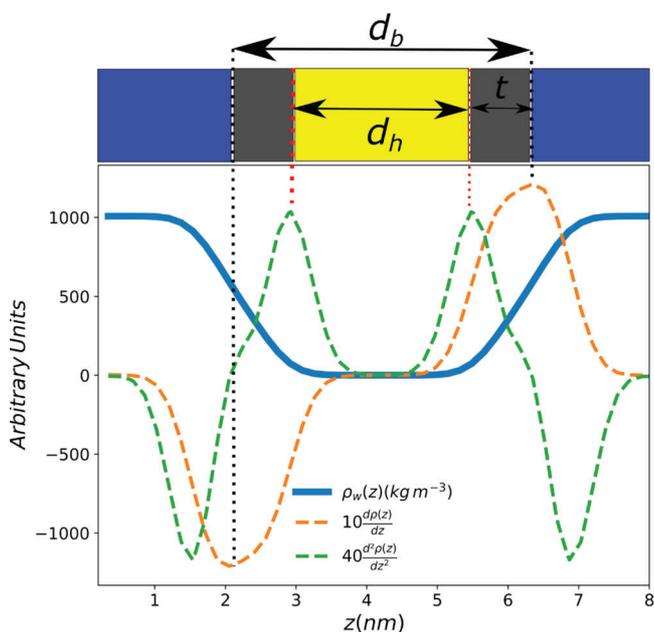


Figure 2. Definitions of various thicknesses used from the relevant profile (water density (ρ_w) shown here). The extrema from the first and second derivatives of the density are used to define the thicknesses, bilayer thickness (d_b), hydrophobic thickness (d_h), and interface thickness (t). To facilitate visualization, in the figure the quantities are scaled by an arbitrary prefactor.

GROMACS tools were used to obtain water density profiles, with a binning size for all systems of about 0.4 Å. In instances where the first derivative was showing two close maxima/minima, the average value between the two peaks was chosen for the calculation of d_b . The distance between the phosphorus atoms, $d_b(P-P)$, was calculated using the FATSLIM program.⁴⁶

Electrical Membrane Thickness from MD Simulations.

An analogous strategy to that described above was followed to compute the interfacial and bilayer thicknesses using information from the dielectric permittivity profile of the solvated bilayers; in particular, in this case, the topology of function $(\epsilon_{zz} - 1)/4\pi\epsilon_{zz}$ was used instead of that of the water density profile to define d_b , d_h , and t from the atomistic simulations.

The dielectric permittivity profiles of the lipid bilayers were constructed using the methodology proposed by Stern and Feller,⁴⁷ which we summarize briefly here. The calculation of the corresponding profiles is based on the evaluation of the components of the dielectric constant parallel and perpendicular to the bilayer surface according to the following expressions

$$\epsilon_{\parallel} = 1 + \frac{4\pi}{2k_B T} \langle P_{\parallel}(z) M_{\parallel} \rangle \quad (3)$$

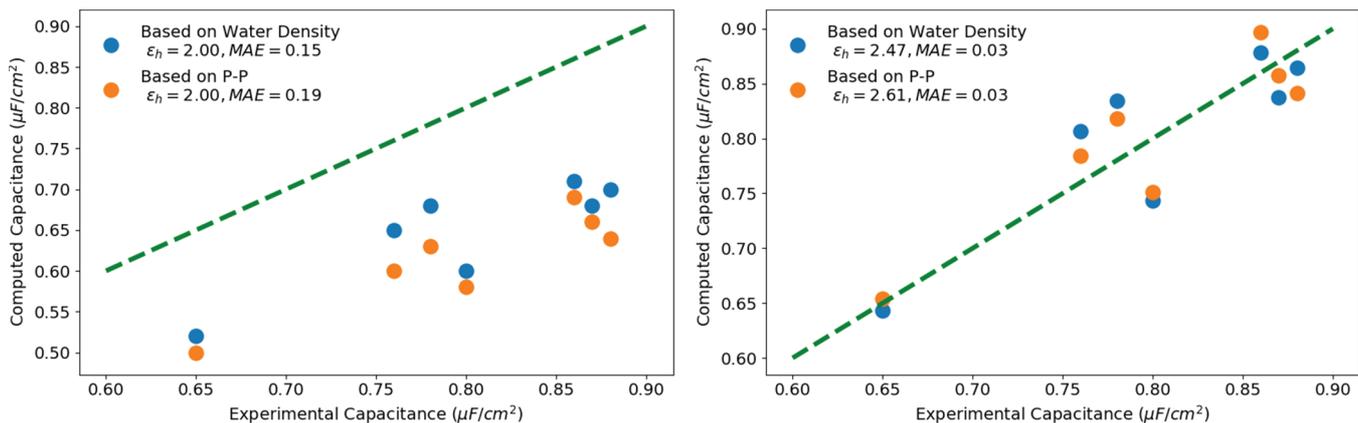


Figure 3. Comparison between experimental measurements of membrane capacitance from Garten et al.²⁷ and predicted values using two definitions of bilayer thickness from atomistic MD trajectories using the slab model, namely, the water density profile and P–P distance. (Left) Capacitance derived using $\epsilon_h = 2.0$. (Right) Capacitance computed using best-fit relative dielectric constants. The dotted lines in green represent the $y = x$ line.

$$\frac{\epsilon_{zz} - 1}{4\pi\epsilon_{zz}} = \frac{1}{k_B T} \frac{(\langle P_z(z)M_z \rangle - \langle P_z(z) \rangle \langle M_z \rangle)}{1 + \frac{4\pi(\langle M_z^2 \rangle - \langle M_z \rangle^2)}{Vk_B T}} \quad (4)$$

where V is the volume, $\langle \rangle$ indicates the ensemble average, k_B is the Boltzmann constant, T is the temperature, z is the direction perpendicular to the bilayer plane, and $\mathbf{P}(z)$ and \mathbf{M} are the polarization density and total dipole moment vectors, respectively, which are defined as

$$\mathbf{P}(z) = \frac{1}{A\Delta z} \sum_i \mu_i \delta(z - z_i) \quad (5)$$

$$\mathbf{M} = \sum_i \mu_i \quad (6)$$

where A is the cross-sectional area of the simulation box, Δz is the bin width, and μ_i is the dipole moment vector associated with a bond i between two atoms. Equations 3 and 4 are the result of combining two different descriptions of the polarization and dipole moment vectors. Thus, combining their microscopic statistical mechanical description with the macroscopic continuum electrostatics one (where the dielectric tensor is defined as a proportionality constant) allows us to express the dielectric permittivity tensor in terms of the fluctuations in both vectors.^{45,47–51}

Spatial polarization density profiles were evaluated by binning along the z direction, after computing the local dipole moments defined at the covalent bond midpoints according to the protocol described elsewhere.⁴⁷ All of the aforementioned quantities were computed by averaging over the MD trajectories. To this end, we used an in-house parallel implementation coded in Python 3 that includes calls to the MDTraj library⁵² to iteratively read the binary xtc files generated by GROMACS. The implementation is freely available and can be downloaded from GitHub: <https://github.com/vannilab/MemCap>.

Notably, ϵ_{zz} presents big fluctuations and a low signal-to-noise ratio; however, the functional form in eq 4, because of its nature, is smoother, so after appropriate denoising (see below), the topological features of this function can be unambiguously identified. For that reason, $(\epsilon_{zz} - 1)/4\pi\epsilon_{zz}$, termed the dielectric function in the rest of the article, was used in this study instead of ϵ_{zz} itself.

Noise Removal from the Dielectric Function. As mentioned above, our procedure requires the accurate determination of the topological properties of the first and second derivatives of the dielectric function to determine reliable membrane thicknesses. Consequently, prior to any analysis, the noise that still exists in this relatively rough function must be removed. To this end, we considered the noisy data to be the independent variable and built a generalized linear model in which cubic B-splines were used as regressors. Finding a unique solution (coefficients of the B-spline basis set) to this inverse problem requires solving the corresponding least-squares problem by imposing some kind of regularization. We used ridge regression,⁵³ which imposes a penalty on the size of the B-spline coefficients. This involves minimizing the following penalized residual sum of squares

$$\min \|X\omega - y\|_2^2 + \lambda \|\omega\|_2^2$$

where X is the design matrix, y is the independent variable (noisy data), ω are the B-spline coefficients, and λ is the regularization parameter required to avoid overfitting the data. We determined the value for λ via generalized cross validation.⁵⁴

Relative Dielectric Determination from Linear Regression Fitting. The linear regression protocol used to determine the optimal value for the dielectric constant of the hydrophobic region, ϵ_h , was based on minimizing the mean absolute error (MAE) between the computed and experimental membrane capacitances. Although other loss functions could have been used with a minimal influence on the optimal value of ϵ_h , we chose MAE as the loss and metric because it provides a direct interpretation of the error. When no fitting was performed, a value of $\epsilon_h = 2.0$ was employed.

RESULTS AND DISCUSSION

Estimation of Bilayer Capacitance from Physical Properties. The origin of the discrepancies found between the bilayer thicknesses and dielectric constants that had to be assumed in previous experimental studies^{27,29} to fit capacitance measurements is unknown and calls for further investigation. Although the conclusions from these experiments seem to indicate that the slab model captures with sufficient accuracy the physics underlying the process under study, the above-mentioned inconsistency questions some of the decisions that

must be made for the application of this model to this kind of system, such as the manner in which membrane thickness is defined or the protocols used to calculate the dielectric permittivity and its different contributions. In this work, we explored the use of atomistic simulations as an alternative methodology to correlate bilayer thickness with membrane capacitance and to provide a molecular explanation for the validity of the slab model.

To this end, we performed MD simulations of various solvated lipid bilayers for which membrane capacitances have been experimentally measured in solvent-free bilayers,²⁷ and from the corresponding trajectories, we evaluated the hydrophobic thickness (d_h) by averaging over all of the snapshots extracted from the dynamics by using different strategies for d_h determination.

Two distinct approaches were initially considered. In the first one, we first estimated d_b as the ensemble average of the upper-to-lower distance between the phosphorus atoms belonging to the upper and lower lipid leaflets. In the second one, d_b was determined from the water density profile as the distance between the extrema of its first derivative. In both cases, we relied on the topological properties of the first derivatives of the water density profile obtained from the MD simulations to compute the interfacial thickness. (See the [Methods](#) section for further details.)

Remarkably, membrane capacitance values obtained from these measurements using $\epsilon_h = 2.0$ for the relative dielectric constant, a value that is generally accepted for nonpolar hydrocarbons,⁵⁵ resulted in very poor agreement with experimental measurements of membrane capacitance ([Figure 3a](#)). Next, rather than assuming a fixed value for ϵ_h , we sought to estimate it by fitting the best value from the data using a linear regression model ([Methods](#)). This led to relatively high dielectric constants: $\epsilon_h \approx 2.61$ (MAE = 0.03) for the former approach and $\epsilon_h \approx 2.47$ (MAE = 0.03) for the latter one. Taken together, these results indicate that, assuming the validity of the slab model in this scenario, both protocols lead to inadequate d_h definitions.

Estimation of Bilayer Capacitance from Electrical Properties. To overcome these limitations, we sought to estimate d_h directly from the electrical properties of the lipid bilayer by calculating the bilayer dielectric profile from the fluctuations found in the polarization and dipole moment vectors along the MD trajectories.^{43,45,47} This approach is limited to uncharged molecules,⁴⁷ but it is theoretically rigorous and can be applied to the investigation of the role of organic solvents, such as hexadecane and squalene, typically used in reconstitution studies.⁵⁶ In addition, with the sole exception of one composition (DOPC/DOPS), all available capacitance measurements on solvent-free membranes have been performed on lipid bilayers constituted by noncharged lipids.²⁷

The bilayer dielectric profile is shown in [Figure 4](#) for a representative lipid composition (DOPC) together with its first and second derivatives.

Using the value estimated for d_h from this profile, we computed, in a parameter-free approach, the capacitance values directly from [eq 2](#) using a fixed value of $\epsilon_h = 2.0$ for all of the systems investigated. In [Figure 5](#), we report the values of capacitances obtained using this approach. As clearly shown there, they show good agreement with the corresponding experimental values. Furthermore, the best fit for ϵ_h using a

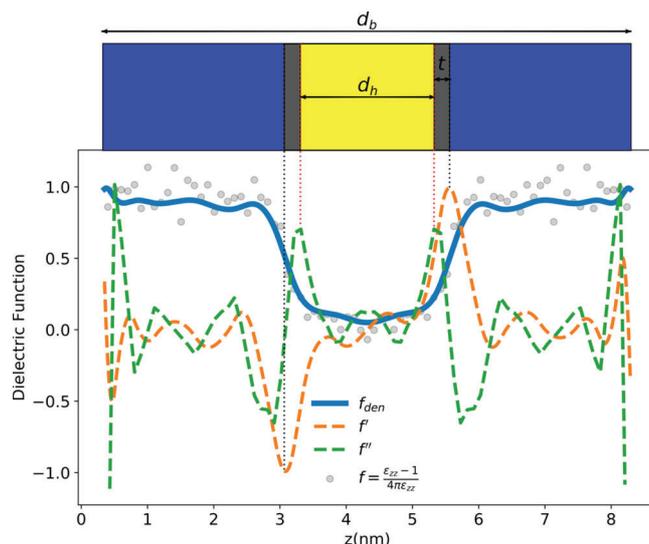


Figure 4. Dielectric function of a DOPC bilayer before (f) and after (f_{den}) denoising in the direction perpendicular (z) to the membrane surface shown together with its first and second derivatives. The extrema from the first (f') and second derivatives (f'') of the dielectric function are used to define the thicknesses, bilayer thickness (d_b), hydrophobic thickness (d_h), and interface thickness (t). To facilitate visualization, in the figure, the quantities are scaled by an arbitrary prefactor.

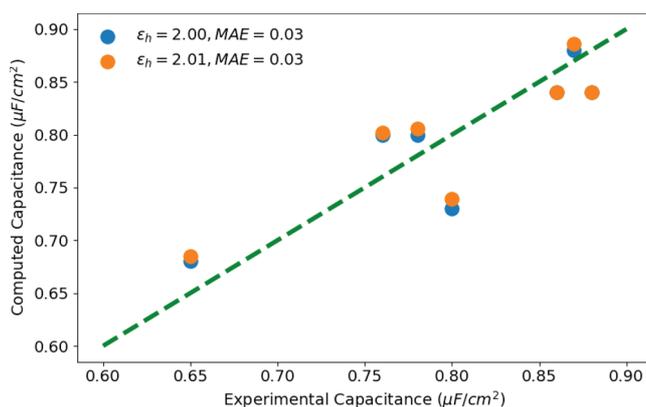


Figure 5. Capacitance values computed using the hydrophobic bilayer thickness from the dielectric profile in the z direction vs experimental values from Garten et al.²⁷ (The error bars are about the same size as the symbols.) The dotted line in green represents the $y = x$ line.

linear regression model led to a value of $\epsilon_h = 2.01$ that is consistent with experimentally derived values for alkanes.²⁵

Notably, the computed bilayer thicknesses d_b show a poor match with the experimental data obtained from X-ray experiments ([Table 2](#)). This suggests that the “membrane capacitor thickness” of the bilayer is different from simply the physical thickness of the hydrocarbon chains, possibly as a result of molecular interactions arising in the lipid bilayer at the interface between the hydrophobic and hydrophilic regions.

Finally, although thickness and capacitance values computed via the various approaches described above correlate to varying degrees with the experimental values, it appears that the simple slab model is able to describe the underlying physics quite well. The molecular origin of such behavior is apparent from [Figure 4](#), where the dielectric profile in the direction perpendicular to the membrane surface resembles that of a low dielectric slab

Table 2. Capacitance Values, Hydrophobic Thicknesses Obtained from Experiments (exp), and the Dielectric Profiles (ϵ_{zz}) Are Presented^a

lipid systems	capacitance ($\mu\text{F}/\text{cm}^2$)			d_h (nm)		
	exp	from ϵ_{zz}	Δ	exp	from ϵ_{zz}	Δ
DOPC	0.87 ²⁷	0.88 \pm 0.01	0.01	2.11 ⁵⁷	2.01 \pm 0.01	-0.10
SOPC	0.78 ²⁷	0.80 \pm 0.01	0.02	2.36 ⁵⁷	2.21 \pm 0.02	-0.15
DPhPC	0.86 ²⁷	0.84 \pm 0.11	-0.02	2.08 ⁵⁸	2.12 \pm 0.28	0.04
DOPC/CHOL	0.80 ²⁷	0.73 \pm 0.01	-0.07	2.34 ⁵⁷	2.41 \pm 0.01	0.07
SOPC/CHOL	0.65 ²⁷	0.68 \pm 0.01	0.03	2.80 ⁵⁷	2.60 \pm 0.03	-0.20
DPhPC/CHOL	0.76 ²⁷	0.80 \pm 0.11	0.04	n.a.	2.22 \pm 0.30	-
DOPC/DPPC	0.88 ²⁷	0.84 \pm 0.05	-0.04	n.a.	2.12 \pm 0.14	-

^aFor the sake of comparison, all values are rounded to the second decimal even in the presence of larger errors. Δ is the difference between computed and experimental values.

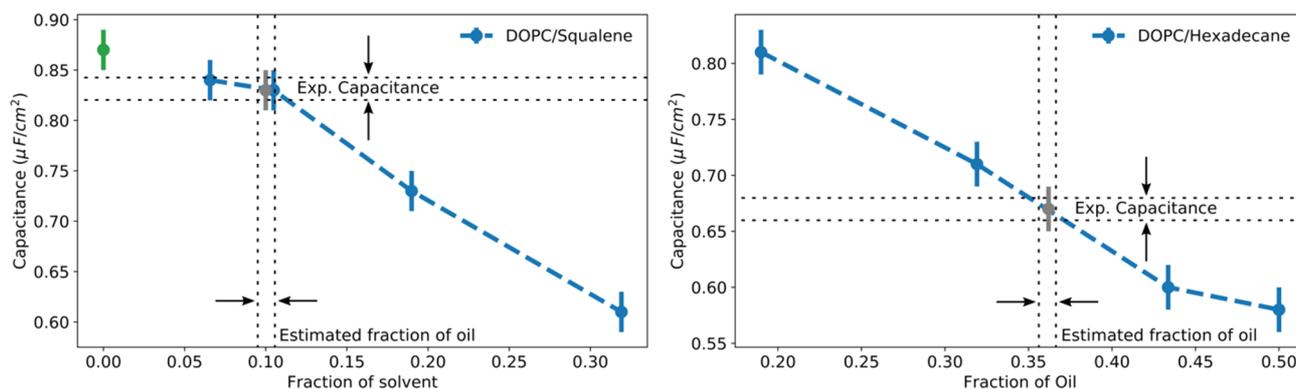


Figure 6. Capacitance measured for DOPC with oil solvents: squalene and hexadecane. The experimental values²⁹ (indicated in gray) can be used to infer the unknown oil concentration. The experimental capacitance of pure DOPC bilayers is 0.87 $\mu\text{F}/\text{cm}^2$ (marked in green).

submerged in water, with the dielectric function switching its value rather abruptly as we move from bulk water to the lipid bilayer. Remarkably, the switching between low and high values is significantly more gradual in the water density profile (Figure 2), further supporting why estimations of bilayer thickness from physical properties are unsatisfactory for the prediction of capacitance values.

Capacitance Measurements in Solvent-Rich Lipid Bilayers. In several reconstituted systems, membranes might contain residual synthetic or natural hydrophobic solvents (e.g., oils) as a result of the bilayer formation process. Measuring the oil concentration in this kind of systems is extremely challenging; therefore, an accurate theoretical determination of residual solvent based on the procedure outlined in this article could be of interest because no alternative approaches are currently available.

We thus decided to use our protocol to predict the solvent concentration from available membrane capacitance measurements. To do so, we computed the membrane capacitance from MD simulations of solvent-rich lipid bilayers at multiple solvent concentrations and subsequently used these data as calibration curves to identify the correct solvent concentration by comparing the predicted data with those from experiments (Figure 6). In particular, we focused on DOPC bilayers including various concentrations of squalene and hexadecane within the bilayer because these solvents are widely used in the formation of lipid membranes.^{29,59}

Our theoretical results are summarized in Table 3 and Figure 6 and show that, as expected, the membrane thickens when increasing the oil concentration. As a consequence of membrane thickening, capacitance values decrease in the

Table 3. Capacitances and Thicknesses Computed from MD Simulations of DOPC Bilayers in the Presence of Different Amounts of Squalene and Hexadecane^a

oil fraction	C ($\mu\text{F}/\text{cm}^2$)	$d_h(\epsilon_{zz})$ (nm)	$d_b(\text{P-P})$ (nm)
Hexadecane			
0.00	0.88 \pm 0.01	2.01 \pm 0.01	3.89 \pm 0.01
0.19	0.81 \pm 0.01	2.18 \pm 0.01	4.13 \pm 0.01
0.32	0.72 \pm 0.12	2.51 \pm 0.43	4.38 \pm 0.01
0.43	0.60 \pm 0.01	2.93 \pm 0.01	4.77 \pm 0.02
0.50	0.58 \pm 0.01	3.05 \pm 0.01	5.11 \pm 0.01
Squalene			
0.00	0.88 \pm 0.01	2.01 \pm 0.01	3.89 \pm 0.01
0.065	0.84 \pm 0.01	2.10 \pm 0.01	4.00 \pm 0.01
0.1	0.84 \pm 0.06	2.11 \pm 0.15	4.07 \pm 0.01
0.19	0.73 \pm 0.08	2.44 \pm 0.28	4.26 \pm 0.01
0.32	0.61 \pm 0.04	2.89 \pm 0.17	4.78 \pm 0.03

^aThe bilayer thickness from P-P distances is shown as well. Capacitance values were computed using the hydrophobic thickness estimated from the dielectric profile. For the sake of comparison, all values are rounded to the second decimal point even in the presence of larger errors.

presence of solvents. This behavior is markedly different for different solvents, as was expected from the widely distinct measurements reported in the literature between squalene and hexadecane.²⁹ In detail, our approach predicts that the solvent-to-lipid concentration of squalene in reported experimental studies²⁹ is below 10%, while hexadecane is present at concentrations slightly above 35%. Thus, our method suggests that lipid bilayers prepared with hexadecane, such as BLMs,¹⁴ DIBs,¹¹ and LAMBs,⁷ might have physical properties that are

significantly different from those of solvent-free lipid bilayers (e.g., the P–P distance as shown in Table 3). As such, our results indicate that data originating from these approaches must be interpreted with particular care for what pertains to membrane properties.

■ CONCLUSIONS

In this work, we present a methodology based on previous work by Stern and Feller⁴⁷ to estimate the membrane capacitance of lipid bilayers from MD simulations. We observe a very good correlation between the capacitance values computed using our approach and recent measurements on solvent-free lipid bilayers.²⁷ This allows us to provide insights into the electrical properties of lipid bilayers and to propose a protocol, based on calibration curves, to estimate the amount of residual solvent based on capacitance measurements in reconstituted lipid bilayers.

We found that a textbook uniform slab approximation works well for bilayer membranes and that the hydrophobic thickness determined from X-ray measurements does not provide a good estimation of the membrane capacitor thickness.

Our methodology is limited to systems composed of uncharged molecules. As such, the approach we propose is not suited to the study of lipid bilayers in the presence of charged lipids or ions. The good correlation we observe between our computational results in the absence of ions and experimental capacitance measurements in the presence of physiological ion concentrations,²⁷ however, seems to suggest that bilayer capacitance does not originate from variations in charge distribution due to ions (that take place close to the bilayer surface) but rather is solely due to the low permittivity of the hydrophobic layer. In addition, potential artifacts may arise from a possible unsatisfactory description of ion–membrane interactions in nonpolarizable atomistic force fields.⁶⁰

In order to include charged lipids and ions in the determination of membrane capacitance from MD simulations, we envision that alternative methodologies, possibly based on different electrical properties of lipid bilayers such as bilayer charge or electric field,^{61,62} will be required. However, the lack of further experimental data on charged lipid systems (only one lipid composition is currently available²⁷) is a major roadblock in this direction.

The main motivation of this work was to be able to infer, in solvent-rich lipid bilayers, the solvent concentration based on capacitance measurements and, consequently, to investigate structural and physical properties of lipid bilayers in the presence of residual solvent molecules. In fact, several reconstitution approaches are not solvent-free, but they generally represent very convenient methodologies for the investigation of complex lipid bilayers, including asymmetric ones,⁶³ in combination with proteins.⁶⁴ However, to our knowledge, the exact determination of solvent concentration in such reconstituted systems is not currently possible, and the difference in capacitance measurements between solvent-rich and solvent-free lipid bilayers can vary from almost zero to up to more than 30%.²⁷ While the low end of this spectrum is in good agreement with NMR measurements on oil concentration in small unilamellar vesicles (SUV)^{65,66} and with coarse-grained MD simulations mimicking droplet–interface bilayers,^{67,68} the concentration of solvent molecules in the high end of the spectrum (i.e., for capacitance values of around 0.6–0.7 $\mu\text{F}/\text{cm}^2$ are typical of alkanes (>10 carbons)^{19,69}) remains

unknown. Using the methodology developed in this work, we have been able to estimate the mol/mol concentration of hexadecane in DOPC lipid bilayers at approximately 30%. Remarkably, this corresponds to a phosphate-to-phosphate thickness increase of 0.5 nm and an increase in the area per lipid from 0.67 (± 0.01) to 0.71 (± 0.01) nm^2 in comparison with solvent-free DOPC lipid bilayers.

In summary, our methodology provides a viable strategy for a better interpretation of the experimental results in solvent-rich lipid bilayers while concomitantly allowing for more informed experimental design. In particular, we foresee that our approach could be particularly useful in guiding the design of the optimal lipid composition for the incorporation of integral membrane proteins in reconstituted membranes given the importance of hydrophobic matching between lipid bilayers and integral membrane proteins.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jpcc.0c03145>.

Comparison between experimental measurements of hydrophobic thickness from X-ray experiments and predicted values using three definitions of bilayer thickness from atomistic MD trajectories using the slab model (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

- (1) Escribá, P. V.; González-Ros, J. M.; Goñi, F. M.; Kinnunen, P. K. J.; Vigh, L.; Sánchez-Magraner, L.; Fernández, A. M.; Busquets, X.; Horváth, I.; Barceló-Coblijn, G. Membranes: A Meeting Point for Lipids, Proteins and Therapies: Translational Medicine. *J. Cell. Mol. Med.* **2008**, *12* (3), 829–875.
- (2) van Meer, G.; Voelker, D. R.; Feigenson, G. W. Membrane Lipids: Where They Are and How They Behave. *Nat. Rev. Mol. Cell Biol.* **2008**, *9* (2), 112–124.
- (3) Cheng, X.; Smith, J. C. Biological Membrane Organization and Cellular Signaling. *Chem. Rev.* **2019**, *119* (9), 5849–5880.
- (4) Watson, H. Biological Membranes. *Essays Biochem.* **2015**, *59*, 43–70.
- (5) Lee, A. G. Biological Membranes: The Importance of Molecular Detail. *Trends Biochem. Sci.* **2011**, *36* (9), 493–500.
- (6) Harayama, T.; Riezman, H. Understanding the Diversity of Membrane Lipid Composition. *Nat. Rev. Mol. Cell Biol.* **2018**, *19* (5), 281–296.
- (7) Beltramo, P. J.; Van Hooghten, R.; Vermant, J. Millimeter-Area, Free Standing, Phospholipid Bilayers. *Soft Matter* **2016**, *12* (19), 4324–4331.
- (8) Seddon, A. M.; Curnow, P.; Booth, P. J. Membrane Proteins, Lipids and Detergents: Not Just a Soap Opera. *Biochim. Biophys. Acta, Biomembr.* **2004**, *1666* (1–2), 105–117.
- (9) Ritchie, T. K.; Grinkova, Y. V.; Bayburt, T. H.; Denisov, I. G.; Zolnerciks, J. K.; Atkins, W. M.; Sligar, S. G. Reconstitution of Membrane Proteins in Phospholipid Bilayer Nanodiscs. *Methods Enzymol.* **2009**, *464* (C), 211–231.
- (10) Richter, R. P.; Bérat, R.; Brisson, A. R. Formation of Solid-Supported Lipid Bilayers: An Integrated View. *Langmuir* **2006**, *22* (8), 3497–3505.
- (11) Bayley, H.; Cronin, B.; Heron, A.; Holden, M. A.; Hwang, W. L.; Syeda, R.; Thompson, J.; Wallace, M. Droplet Interface Bilayers. *Mol. BioSyst.* **2008**, *4* (12), 1191–1208.
- (12) Castellana, E. T.; Cremer, P. S. Solid Supported Lipid Bilayers: From Biophysical Studies to Sensor Design. *Surf. Sci. Rep.* **2006**, *61* (10), 429–444.
- (13) Mueller, P.; Rudin, D. O.; Ti Tien, H.; Wescott, W. C. Reconstitution of Cell Membrane Structure in Vitro and Its Transformation into an Excitable System. *Nature* **1962**, *194* (4832), 979–980.
- (14) Papahadjopoulos, D. Bilayer Lipid Membranes (BLM): Theory and Practice By H. Ti Tien. *Perspect. Biol. Med.* **1975**, *18* (4), 575–577.
- (15) Sharpe, H. J.; Stevens, T. J.; Munro, S. A Comprehensive Comparison of Transmembrane Domains Reveals Organelle-Specific Properties. *Cell* **2010**, *142* (1), 158–169.
- (16) Clay, L.; Caudron, F.; Denoth-Lippuner, A.; Boettcher, B.; Frei, S. B.; Snapp, E. L.; Barral, Y. A Sphingolipid-Dependent Diffusion Barrier Confines ER Stress to the Yeast Mother Cell *eLife* **2014**, *3*, DOI: 10.7554/eLife.01883.
- (17) White, S. H. Phase Transitions in Planar Bilayer Membranes. *Biophys. J.* **1975**, *15* (2), 95–117.
- (18) Taylor, G. J.; Venkatesan, G. A.; Collier, C. P.; Sarles, S. A. Direct in Situ Measurement of Specific Capacitance, Monolayer Tension, and Bilayer Tension in a Droplet Interface Bilayer. *Soft Matter* **2015**, *11* (38), 7592–7605.
- (19) Gross, L. C. M.; Heron, A. J.; Baca, S. C.; Wallace, M. I. Determining Membrane Capacitance by Dynamic Control of Droplet Interface Bilayer Area. *Langmuir* **2011**, *27* (23), 14335–14342.
- (20) Nagle, J. F.; Tristram-Nagle, S. Structure of Lipid Bilayers. *Biochim. Biophys. Acta, Rev. Biomembr.* **2000**, *1469* (3), 159–195.
- (21) Wesolowska, O.; Michalak, K.; Maniewska, J.; Hendrich, A. B. Giant Unilamellar Vesicles - a Perfect Tool to Visualize Phase Separation and Lipid Rafts in Model Systems. *Acta Biochim. Polym.* **2009**, *56* (1), 33–39.
- (22) Everitt, C. T.; Haydon, D. A. Electrical Capacitance of a Lipid Membrane Separating Two Aqueous Phases. *J. Theor. Biol.* **1968**, *18* (3), 371–379.
- (23) Gramse, G.; Dols-Perez, A.; Edwards, M. A.; Fumagalli, L.; Gomila, G. Nanoscale Measurement of the Dielectric Constant of Supported Lipid Bilayers in Aqueous Solutions with Electrostatic Force Microscopy. *Biophys. J.* **2013**, *104* (6), 1257–1262.
- (24) Raudino, A.; Mauzerall, D. Dielectric Properties of the Polar Head Group Region of Zwitterionic Lipid Bilayers. *Biophys. J.* **1986**, *50* (3), 441–449.
- (25) Panuganti, S. R.; Wang, F.; Chapman, W. G.; Vargas, F. M. A Simple Method for Estimation of Dielectric Constants and Polarizabilities of Nonpolar and Slightly Polar Hydrocarbons. *Int. J. Thermophys.* **2016**, *37* (7), 75.
- (26) Scaife, W. G. The Relative Permittivity of the n Alkanes from n Pentane to n Decane as a Function of Pressure and Temperature. *J. Phys. A: Gen. Phys.* **1972**, *5* (6), 897–903.
- (27) Garten, M.; Mosgaard, L. D.; Bornschlöggl, T.; Dieudonné, S.; Bassereau, P.; Toombes, G. E. S. Whole-GUV Patch-Clamping. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114* (2), 328–333.
- (28) Lewis, B. A.; Engelman, D. M. Lipid Bilayer Thickness Varies Linearly with Acyl Chain Length in Fluid Phosphatidylcholine Vesicles. *J. Mol. Biol.* **1983**, *166* (2), 211–217.
- (29) Beltramo, P. J.; Scheidegger, L.; Vermant, J. Toward Realistic Large-Area Cell Membrane Mimics: Excluding Oil, Controlling Composition, and Including Ion Channels. *Langmuir* **2018**, *34* (20), 5880–5888.
- (30) Berendsen, H. J. C.; van der Spoel, D.; van Drunen, R. GROMACS: A Message-Passing Parallel Molecular Dynamics Implementation. *Comput. Phys. Commun.* **1995**, *91* (1–3), 43–56.
- (31) Van Der Spoel, D.; Lindahl, E.; Hess, B.; Groenhof, G.; Mark, A. E.; Berendsen, H. J. C. GROMACS: Fast, Flexible, and Free. *J. Comput. Chem.* **2005**, *26* (16), 1701–1718.
- (32) Jo, S.; Kim, T.; Im, W. Automated Builder and Database of Protein/Membrane Complexes for Molecular Dynamics Simulations. *PLoS One* **2007**, *2* (9), e880.
- (33) Huang, J.; Rauscher, S.; Nawrocki, G.; Ran, T.; Feig, M.; De Groot, B. L.; Grubmüller, H.; MacKerell, A. D. CHARMM36m: An Improved Force Field for Folded and Intrinsically Disordered Proteins. *Nat. Methods* **2017**, *14* (1), 71–73.
- (34) MacKerell, A. D.; Bashford, D.; Bellott, M.; Dunbrack, R. L.; Evanseck, J. D.; Field, M. J.; Fischer, S.; Gao, J.; Guo, H.; Ha, S.; Joseph-McCarthy, D.; et al. All-Atom Empirical Potential for Molecular Modeling and Dynamics Studies of Proteins. *J. Phys. Chem. B* **1998**, *102* (18), 3586–3616.
- (35) Darden, T.; York, D.; Pedersen, L. Particle Mesh Ewald: An N-log(N) Method for Ewald Sums in Large Systems. *J. Chem. Phys.* **1993**, *98* (12), 10089–10092.
- (36) Hess, B.; Bekker, H.; Berendsen, H. J. C.; Fraaije, J. G. E. M. LINC: A Linear Constraint Solver for Molecular Simulations. *J. Comput. Chem.* **1997**, *18* (12), 1463–1472.
- (37) Miyamoto, S.; Kollman, P. A. Settle: An Analytical Version of the SHAKE and RATTLE Algorithm for Rigid Water Models. *J. Comput. Chem.* **1992**, *13* (8), 952–962.
- (38) Evans, D. J.; Holian, B. L. The Nose-Hoover Thermostat. *J. Chem. Phys.* **1985**, *83* (8), 4069–4074.
- (39) Parrinello, M.; Rahman, A. Polymorphic Transitions in Single Crystals: A New Molecular Dynamics Method. *J. Appl. Phys.* **1981**, *52* (12), 7182–7190.
- (40) Tajkhorshid, E. MD Simulation of the integral purple membrane of H. Salinarum http://www.ks.uiuc.edu/~emad/emad_pm.html http://www.ks.uiuc.edu/~emad/emad_pm.html.
- (41) Vermaas, J. V.; Hardy, D. J.; Stone, J. E.; Tajkhorshid, E.; Kohlmeyer, A. TopoGromacs: Automated Topology Conversion from CHARMM to GROMACS within VMD. *J. Chem. Inf. Model.* **2016**, *56* (6), 1112–1116.
- (42) Scopes, R. K. *Protein Purification: Principles and Practices*; Springer Verlag: New York, 1984.
- (43) Kornyshev, A. A.; Schmickler, W.; Vorotyntsev, M. A. Nonlocal Electrostatic Approach to the Problem of a Double Layer at a Metal-Electrolyte Interface. *Phys. Rev. B: Condens. Matter Mater. Phys.* **1982**, *25* (8), 5244–5256.

- (44) Kornyshev, A. A.; Vorotyntsev, M. A. Nonlocal Dielectric Response of the Electrode/Solvent Interface in the Double Layer Problem. *Can. J. Chem.* **1981**, *59* (13), 2031–2042.
- (45) Bonthuis, D. J.; Gekle, S.; Netz, R. R. Dielectric Profile of Interfacial Water and Its Effect on Double-Layer Capacitance. *Phys. Rev. Lett.* **2011**, *107* (16), 166102.
- (46) Buchoux, S. FATSLiM: A Fast and Robust Software to Analyze MD Simulations of Membranes. *Bioinformatics* **2017**, *33* (1), 133–134.
- (47) Stern, H. A.; Feller, S. E. Calculation of the Dielectric Permittivity Profile for a Nonuniform System: Application to a Lipid Bilayer Simulation. *J. Chem. Phys.* **2003**, *118* (7), 3401–3412.
- (48) Zhou, F.; Schulten, K. Molecular Dynamics Study of a Membrane - Water Interface. *J. Phys. Chem.* **1995**, *99* (7), 2194–2207.
- (49) Ballenegger, V.; Hansen, J. P. Dielectric Permittivity Profiles of Confined Polar Fluids. *J. Chem. Phys.* **2005**, *122* (11), 114711.
- (50) Nymeyer, H.; Zhou, H. X. A Method to Determine Dielectric Constants in Nonhomogeneous Systems: Application to Biological Membranes. *Biophys. J.* **2008**, *94* (4), 1185–1193.
- (51) Shushkov, P. G.; Tzvetanov, S. A.; Ivanova, A. N.; Tadjer, A. V. Dielectric Properties Tangential to the Interface in Model Insoluble Monolayers: Theoretical Assessment. *Langmuir* **2008**, *24* (9), 4615–4624.
- (52) McGibbon, R. T.; Beauchamp, K. A.; Harrigan, M. P.; Klein, C.; Swails, J. M.; Hernández, C. X.; Schwantes, C. R.; Wang, L. P.; Lane, T. J.; Pande, V. S. MDTraj: A Modern Open Library for the Analysis of Molecular Dynamics Trajectories. *Biophys. J.* **2015**, *109* (8), 1528–1532.
- (53) Hoerl, A. E.; Kennard, R. W. Ridge Regression: Biased Estimation for Nonorthogonal Problems. *Technometrics* **1970**, *12* (1), 55–67.
- (54) Golub, G. H.; Heath, M.; Wahba, G. Generalized Cross-Validation as a Method for Choosing a Good Ridge Parameter. *Technometrics* **1979**, *21* (2), 215.
- (55) P, D. Book Review: CRC Handbook of Chemistry and Physics. *J. Mol. Struct.* **1992**, *268* (1–3), 320.
- (56) Beltramo, P. J.; Van Hooghten, R.; Vermant, J. Millimeter-Area, Free Standing, Phospholipid Bilayers. *Soft Matter* **2016**, *12* (19), 4324–4331.
- (57) Pan, J.; Mills, T. T.; Tristram-Nagle, S.; Nagle, J. F. Cholesterol Perturbs Lipid Bilayers Nonuniversally. *Phys. Rev. Lett.* **2008**, *100* (19), 198103.
- (58) Tristram-Nagle, S.; Kim, D. J.; Akhuzada, N.; Kuerka, N.; Mathai, J. C.; Katsaras, J.; Zeidel, M.; Nagle, J. F. Structure and Water Permeability of Fully Hydrated DiphytanoylPC. *Chem. Phys. Lipids* **2010**, *163* (6), 630–637.
- (59) Andersen, O. S.; Koeppe, R. E. Bilayer Thickness and Membrane Protein Function: An Energetic Perspective. *Annu. Rev. Biophys. Biomol. Struct.* **2007**, *36*, 107–130.
- (60) Melcr, J.; Martinez-Seara, H.; Nencini, R.; Kolafa, J.; Jungwirth, P.; Ollila, O. H. S. Accurate Binding of Sodium and Calcium to a POPC Bilayer by Effective Inclusion of Electronic Polarization. *J. Phys. Chem. B* **2018**, *122* (16), 4546–4557.
- (61) Ding, W.; Palaiokostas, M.; Wang, W.; Orsi, M. Effects of Lipid Composition on Bilayer Membranes Quantified by All-Atom Molecular Dynamics. *J. Phys. Chem. B* **2015**, *119* (49), 15263–15274.
- (62) Melcr, J.; Bonhenry, D.; Timr, S.; Jungwirth, P. Transmembrane Potential Modeling: Comparison between Methods of Constant Electric Field and Ion Imbalance. *J. Chem. Theory Comput.* **2016**, *12* (5), 2418–2425.
- (63) Marquardt, D.; Geier, B.; Pabst, G. Asymmetric Lipid Membranes: Towards More Realistic Model Systems. *Membranes (Basel, Switz.)* **2015**, *5* (2), 180–196.
- (64) Shen, H. H.; Lithgow, T.; Martin, L. L. Reconstitution of Membrane Proteins into Model Membranes: Seeking Better Ways to Retain Protein Activities. *Int. J. Mol. Sci.* **2013**, *14* (1), 1589–1607.
- (65) Hamilton, J. A.; Vural, J. M.; Carpentier, Y. A.; Deckelbaum, R. J. Incorporation of Medium Chain Triacylglycerols into Phospholipid Bilayers: Effect of Long Chain Triacylglycerols, Cholesterol, and Cholesteryl Esters. *J. Lipid Res.* **1996**, *37* (4), 773–782.
- (66) Kamp, F.; Westerhoff, H. V.; Hamilton, J. A. Movement of Fatty Acids, Fatty Acid Analogues, and Bile Acids across Phospholipid Bilayers. *Biochemistry* **1993**, *32* (41), 11074–11085.
- (67) Zoni, V.; Campomanes, P.; Khaddaj, R.; Thiam, A. R.; Schneider, R.; Vanni, S. Lipid Droplet Biogenesis Is a Liquid Phase Separation Spatially Regulated by Seipin and Membrane Curvature. *bioRxiv* **2019**, 777466.
- (68) Ben M'barek, K.; Ajjaji, D.; Chorlay, A.; Vanni, S.; Foret, L.; Thiam, A. R. ER Membrane Phospholipids and Surface Tension Control Cellular Lipid Droplet Formation. *Dev. Cell* **2017**, *41*, 591–604.
- (69) Bartlett, P. N. *Bioelectrochemistry: Fundamentals, Experimental Techniques and Applications*; John Wiley & Sons, 2008.