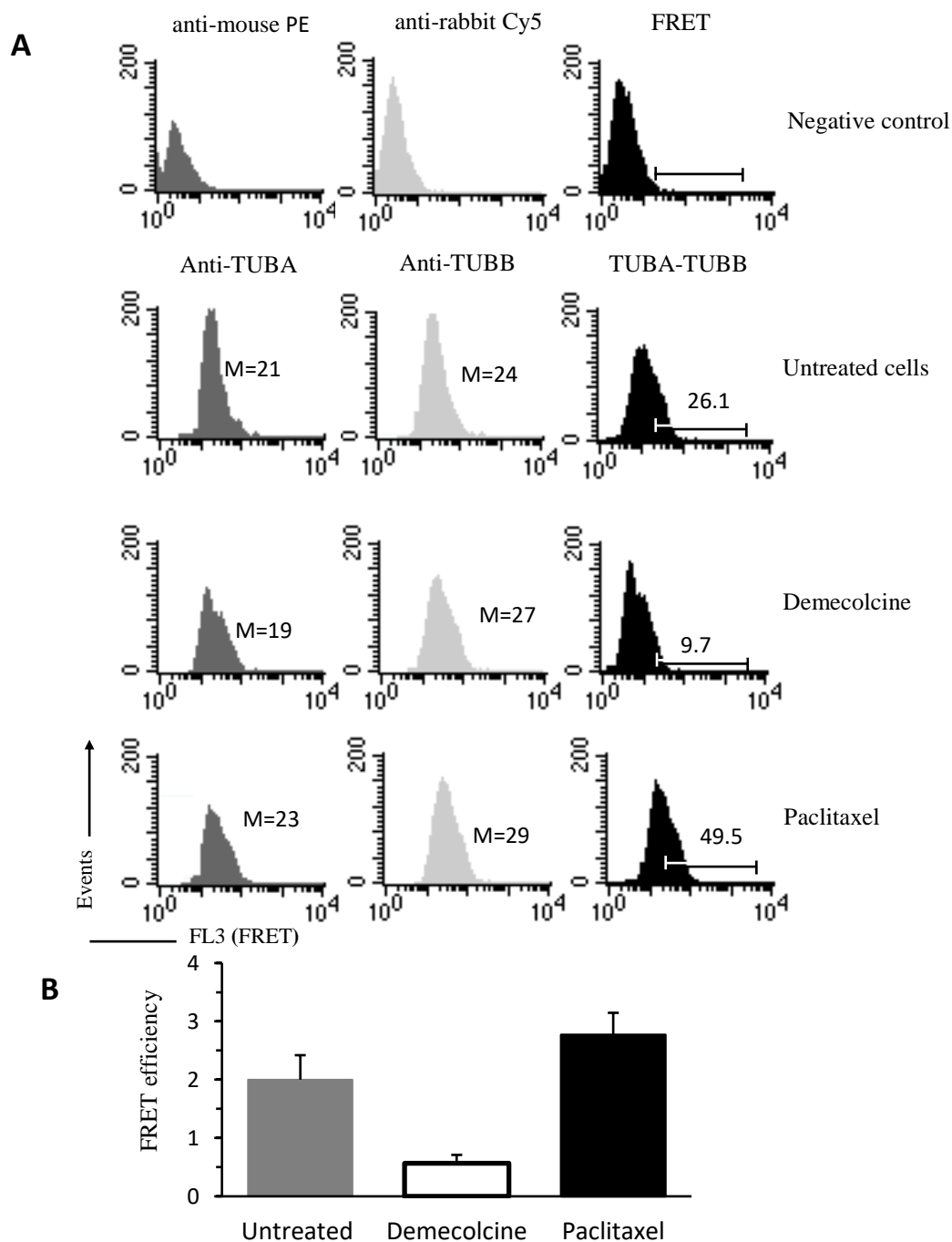
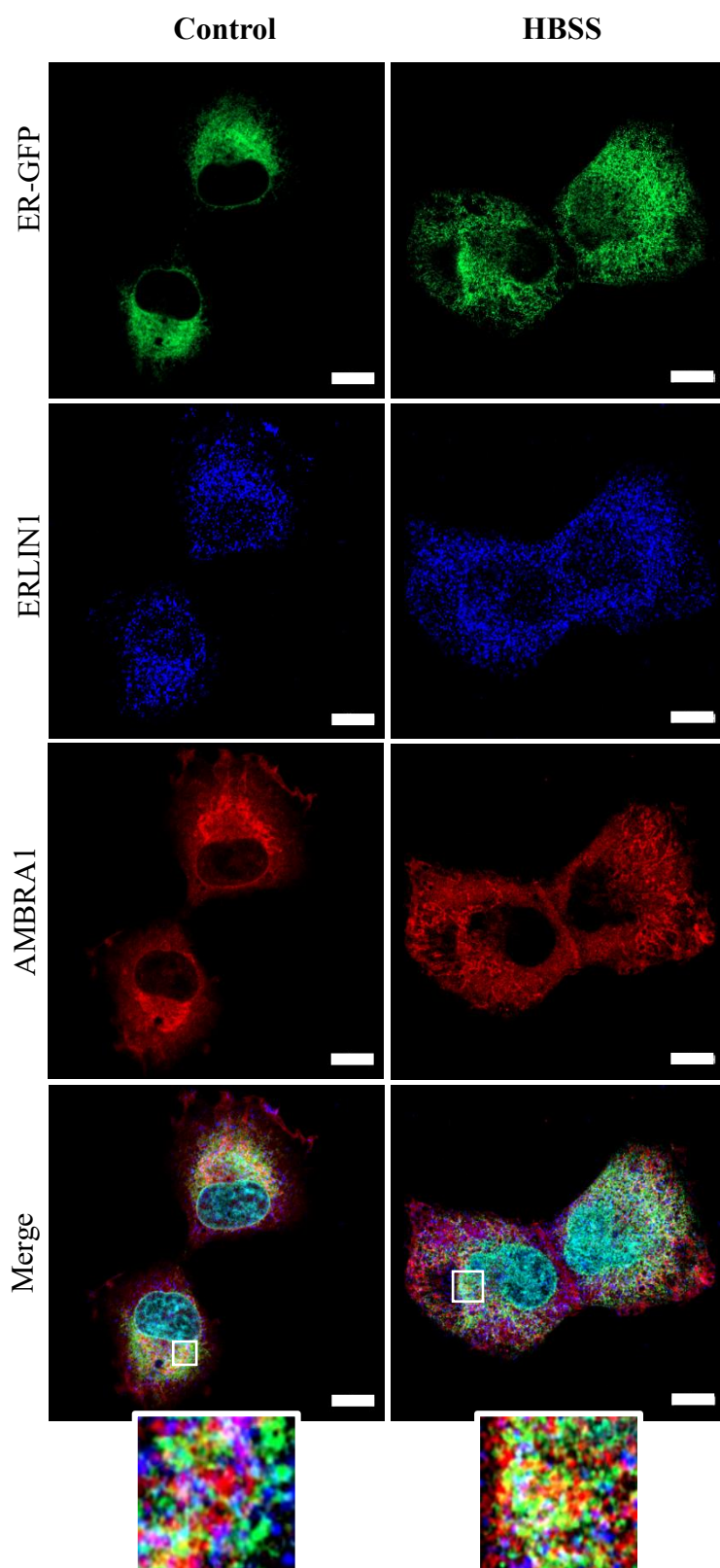


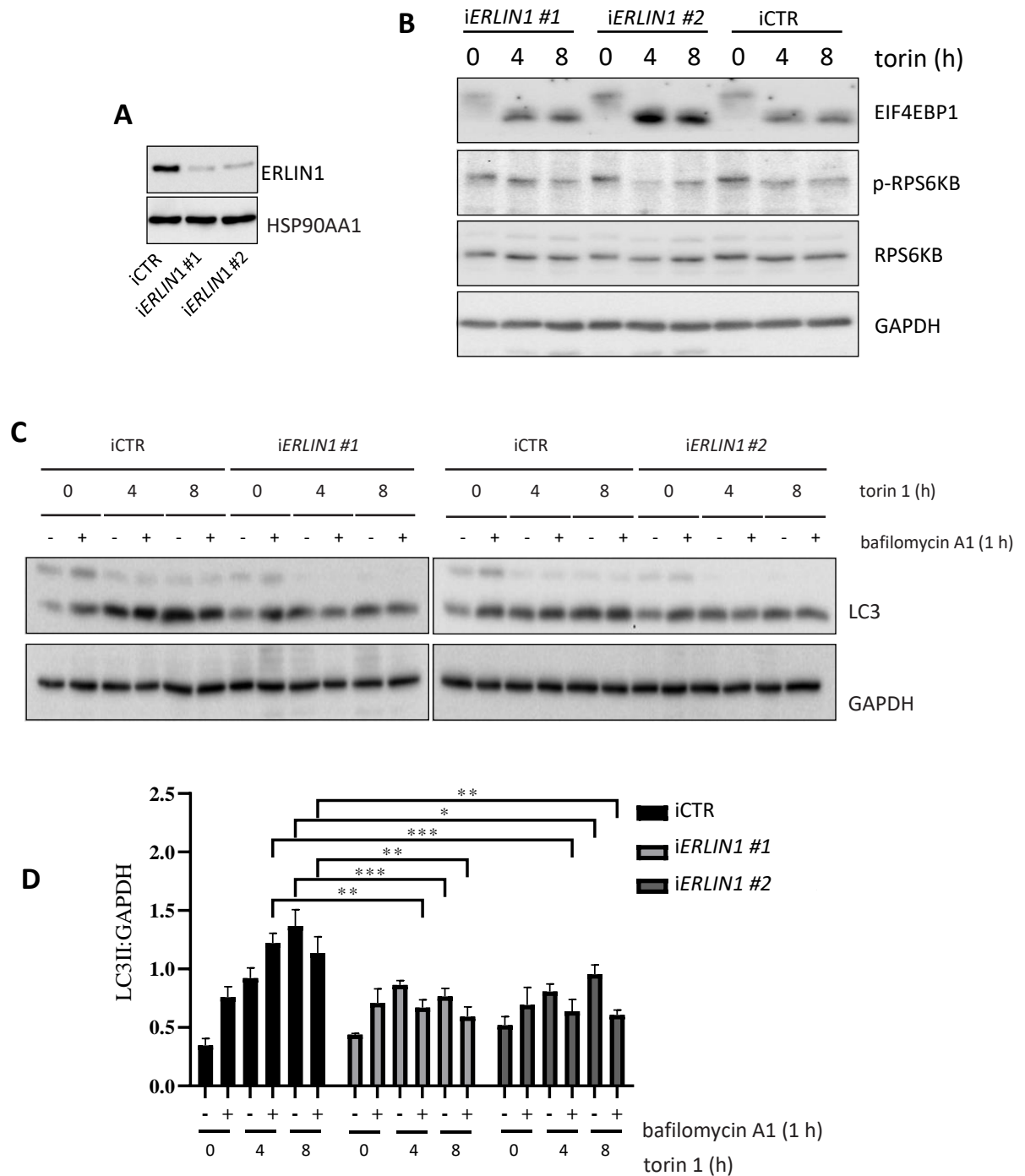
**Figure S1.** Autophagy evaluation performed by flow and static cytometry in 2FTGH cells under different experimental conditions. 2FTGH fibroblasts were treated or not with 50 nM rapamycin (Enzo Life Sciences, BML-A275-0025) for 4 h, 10 mM 3-MA (Sigma-Aldrich, M9281) overnight, or 1  $\mu$ M bafilomycin (Sigma-Aldrich, B1793) for 4 h. At the end of treatment cells were harvested and analyzed by flow cytometry after single staining with Cyto-ID autophagy detection kit (Enzo Life Sciences, ENZ-51031-K200, left panel), or double staining, after fixation and permeabilization, with anti-LC3 (mouse, Invitrogen, GT3612) and anti-SQSTM1 (rabbit, Abcam, ab91526) primary antibodies followed by anti-mouse Alexa Fluor 488 (Invitrogen, A32723, red in figure) and anti-rabbit Alexa Fluor 594-conjugated (Invitrogen, A32740, green in figure) secondary antibodies (central panels), as reported in Material and Methods section. For fluorescence microscopy cells were fixed, permeabilized and stained directly on lenticular slides (pictures in the right panel). In (A) are shown results obtained in a representative experiment. Numbers represent the median fluorescence intensity values of the histograms. Bar graphs in (B) report the mean  $\pm$  SD of the results obtained in three independent experiments.



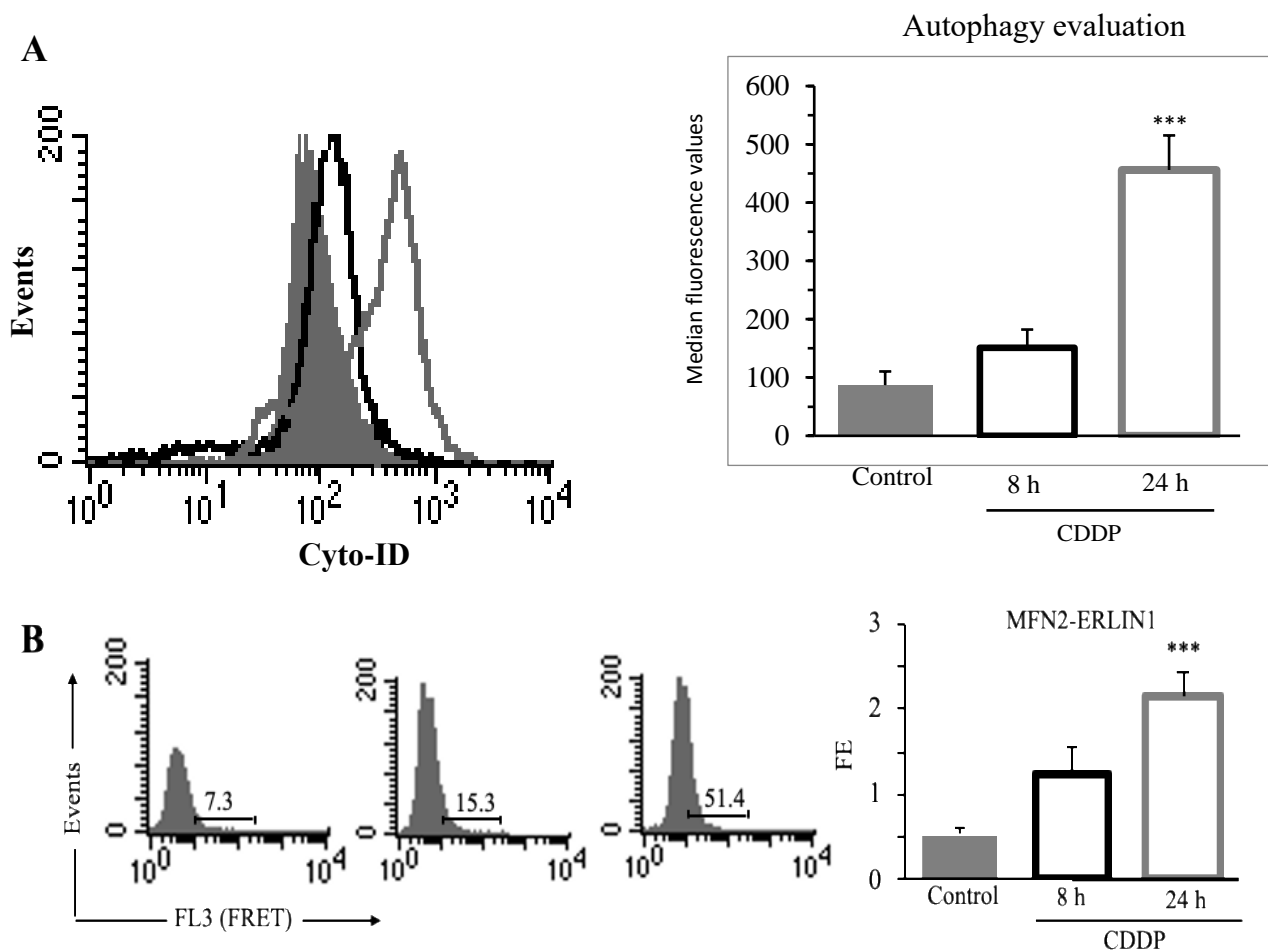
**Figure S2.** As positive control for FRET method, 2FTGH fibroblasts were treated overnight with Demecolcine (0.05  $\mu\text{g/ml}$ , SIGMA-Aldrich, D7385) or with Paclitaxel (10 nM, Sigma-Aldrich, T7191). At the end of treatment cells were double stained with an anti-TUBA monoclonal antibody (Sigma-Aldrich, T9026) and an anti-TUBB polyclonal antibody (Thermo Fisher Scientific, PA1-41331) followed by a goat anti-rabbit IgG H&L (Cy5) preadsorbed (Abcam, ab6564), and a goat F(ab')<sub>2</sub> anti-mouse IgG-Fc (PE) pre-adsorbed (Abcam, ab5881) to quantify molecular association TUBA-TUBB by quantitative FRET. As expected, treatment with Demecolcine, able to depolymerizes microtubules, significantly reduced the molecular association TUBA-TUBB compared to untreated cells. In contrast, Paclitaxel, known to stabilize microtubule polymer, induced a significant increase in the molecular association between TUBA and TUBB. Importantly, the treatment with either drugs did not induce a substantial alteration of the expression of the two proteins (see numbers in A), but only modified their degree of association, as revealed by quantification of FRET efficiency by Riemann's algorithm (bar graph at the bottom). (A) FACS histograms showing results obtained in representative experiments. Numbers represent median fluorescence intensity (in first and second columns) or the percentage of FL3-positive cells (in third column). (B) Bar graph reporting the mean  $\pm$  SD of the results obtained from three independent acquisitions.



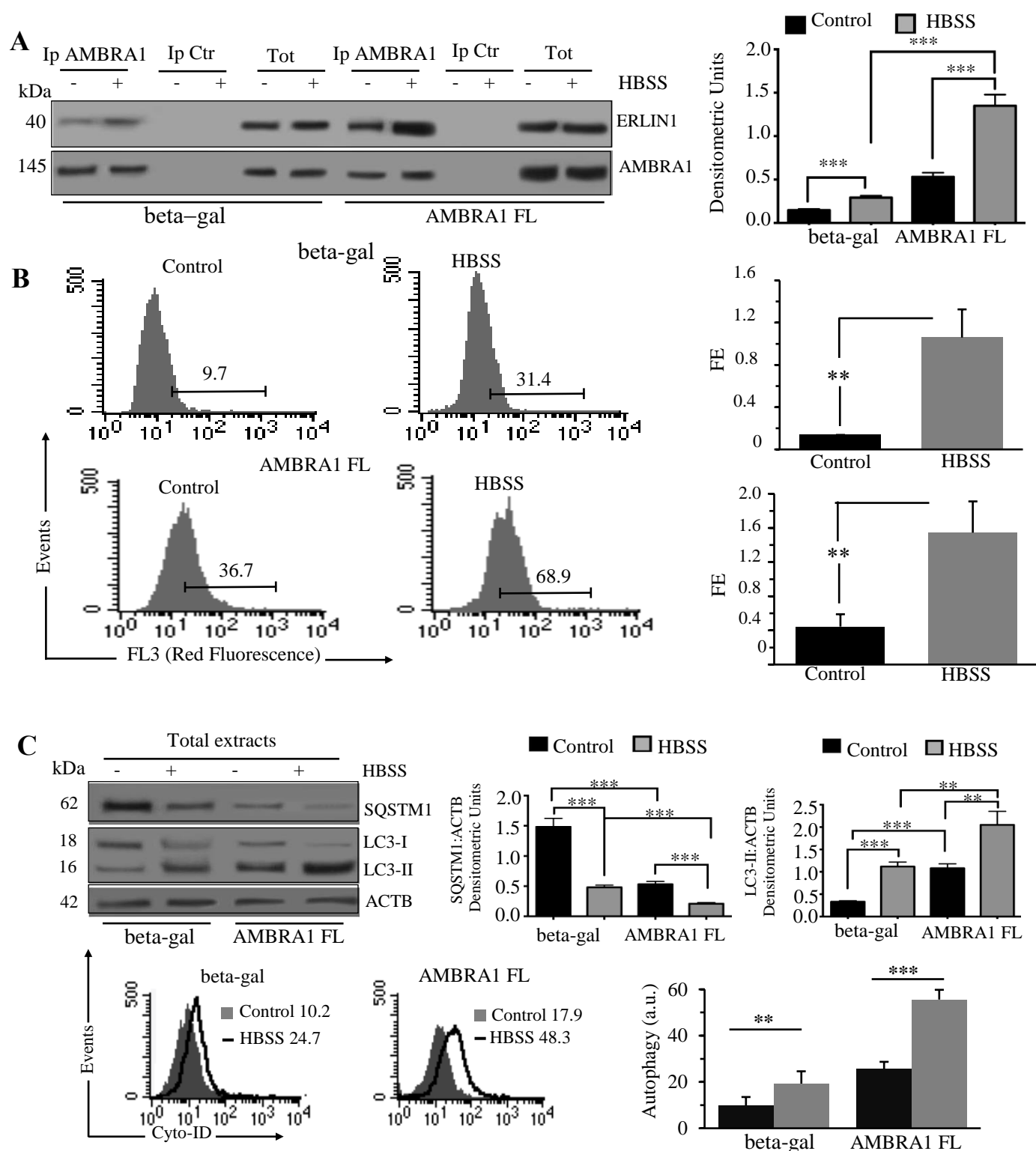
**Figure S3.** AMBRA1-ERLIN1-ER association during autophagy induction. 2F cells expressing ER-GFP (green in figure) were treated or not with HBSS 1 h and stained with anti-AMBRA1 (Cy3, red in figure), anti-ERLIN1 (Alexa Fluor 647, blue in figure) and with DAPI (Turquoise). Images were acquired using a LSM 900, Airyscan SR Zeiss confocal microscopy and the co-localization of ERLIN1 with AMBRA1 and ER-GFP was measured using ZEN 3.0 Blue edition software and expressed as  $\text{mm}^2$  per cell. A minimum of 30 cells/ sample was analyzed and the statistical analysis was performed using Student's t-test. *n.s.*, not significant. Scale bar: 10  $\mu\text{m}$ .



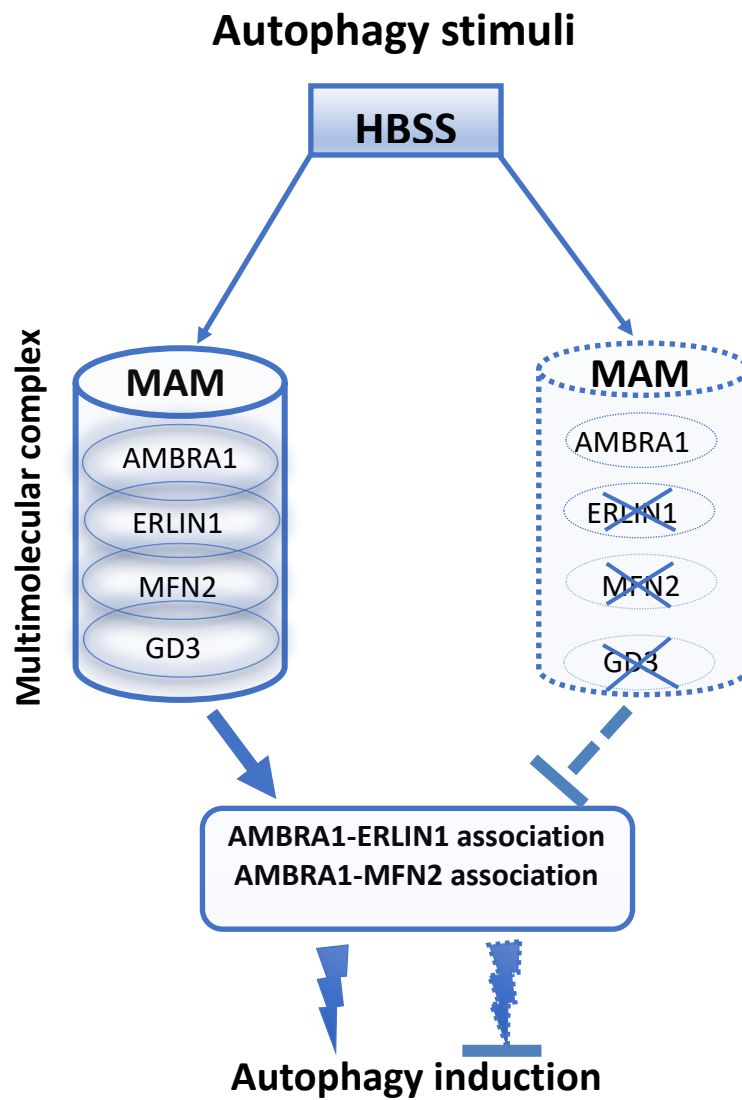
**Figure S4.** Autophagy flux evaluation in ERLIN1 downregulated cells treated with 100 nM torin 1. **(A)** ERLIN1 expression was downregulated by siRNA using two different oligonucleotides in 2F cells and western blotting was performed to evaluate ERLIN1 levels from three independent experiments; \* $p \leq 0.05$ . **(B)** Inhibition of MTOR activity by ERLIN1 downregulated 2F cells was evaluated by western blotting using anti p-RPS6KB1, anti-RPS6KB1 and anti-EIF4EBP1 antibodies. **(C)** siERLIN1 cells were treated with torin 1 (100 nM) for the indicated time points and incubated with the lysosome inhibitor bafilomycin A<sub>1</sub> or vehicle for 1 h before lysis. LC3 lipidation was detected by immunoblotting using a specific antibody; GAPDH was incubated as a loading control. **(D)** The graph reports mean  $\pm$  SD of LC3-II - GAPDH from three independent experiments; \* $p \leq 0.05$ , \*\* $p \leq 0.01$ , \*\*\* $p \leq 0.001$  using ANOVA 2-way test for repeated samples.



**Figure S5.** CDDP cell treatment induced autophagy and MFN2-ERLIN1 association. **(A)** Flow cytometry analysis of autophagy in untreated cells (Control, full gray curves) and after treatment with 8  $\mu$ M CDDP for 8 h (empty black curve) or 24 h (empty grey curve) performed with a Cyto-ID Autophagy Detection kit. Numbers represent the mean  $\pm$  SD of the median fluorescence intensity obtained in three independent experiments. A representative experiment among three is shown. Bar graph shows the mean  $\pm$  SD obtained from three independent experiments. \*\*\*  $p \leq 0.001$  vs Control. **(B)** Quantitative evaluation of MFN2, considered a marker of MAM, and ERLIN1 molecular association by FRET technique in cells untreated (Control) or treated with CDDP 8  $\mu$ M for 8 h or 24 h, as revealed by flow cytometry analysis. Numbers indicate the percentage of FL3-positive events (FRET channel), obtained in one experiment representative of three. Bar graphs on the right show the FRET efficiency, calculated according to the Riemann's algorithm. Data are reported as mean  $\pm$  SD from three independent experiments.



**Figure S6.** AMBRA1-ERLIN1 association during autophagy induction in AMBRA1 FL cells. **(A)** 2F cells were infected with retroviral vectors encoding MYC-tagged AMBRA1 FL or (MYC-tagged beta-gal), untreated or treated with HBSS for 1 h, were lysed in lysis buffer. Protein extracts were subjected to IP using an anti-AMBRA1 PAb. A rabbit IgG isotypic control (IpCtr) was employed. The immunoprecipitates were analyzed for the presence of ERLIN1 by western blot analysis using anti-ERLIN1 MAb. A representative experiment among 3 is shown. \*\*\* $p \leq 0.001$ . As a control, the immunoprecipitates were assessed by immunoblot with anti-AMBRA1 MAb. **(B)** Quantitative evaluation of AMBRA1-ERLIN1 association by FRET technique in both MYC-tagged AMBRA1 FL and MYC-tagged beta-gal, untreated or treated with HBSS, as revealed by flow cytometry analysis. Numbers in the first and second panels indicate the percentage of FL3-positive events (FRET channel), obtained in one experiment representative of 3. Bar graphs on the right show the FRET efficiency, calculated according to the Riemann's algorithm, of AMBRA1 and ERLIN1 molecular association. Data are reported as mean  $\pm$  SD from 3 independent experiments. **(C)** Upper panels, autophagy analysis by western blot, using rabbit anti-LC3 PAb or rabbit anti-SQSTM1 MAb. Loading control was evaluated using anti-ACTB MAb. Bar graph on the right panel shows densitometric analysis. Lower panels, semiquantitative flow cytometry analysis performed with a Cyto-ID Autophagy detection kit; on the right panel shows densitometric analysis. Results represent the mean  $\pm$  SD from 3 independent experiments. \*\* $p < 0.01$  and \*\*\* $p < 0.001$



**Figure S7.** A summary scheme showing as the knocking down MFN2, ST8SIA1 or ERLIN1 expression impairs the interaction AMBRA1-ERLIN1 and AMBRA1-MFN2 hindering autophagy.

**Table S1.** The tables (A-D) reports the identification of ERLIN1 as AMBRA1 interacting protein by tandem affinity purification and LC/Orbitrap.

**A**

Gene Names	Protein Descriptions	Uniprot	Proteins	Peptides (seq)	Sequence Coverage [%]	PEP
AMBRA1;KIAA1736	Isoform 1 of BECN1; Isoform3 of BECN1	Q9C0C71;Q9C0C7; B4DZ97;Q9C0C7-3;Q9C0C7-2	3	69	58.3	0
ERLIN1;C10orf69;KE04;KEO4; SPFH1;RP11-316M21.1-003	ER lipid raft associated 1	O75477;B2RDK6; B4DPN7;B0QZ43	2	9	25.6	4.01E-90

**B**

Exp1a Ratio Ambra FLAG-HA/Ambra untagged	Exp1a Ratio Ambra FLAG-HA/Ambra untagged	Exp1a Ratio Ambra FLAG-HA/Ambra untagged	Exp1a Ratio Ambra FLAG-HA/Ambra untagged	Exp1a Ratio Ambra FLAG-HA/Ambra untagged	Exp1a Ratio Ambra FLAG-HA/Ambra untagged
7.26	135.70	1233	7.74	139.53	1232
1.46	59.58	37	1.62	50.59	37

**C**

Exp2a Ratio Ambra FLAG-HA/Ambra untagged	Exp2a Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp2a Ambra FLAG-HA/Ambra untagged Count	Exp2b Ratio Ambra FLAG-HA/Ambra untagged	Exp2b Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp2b Ambra FLAG-HA/Ambra untagged Count
77.32	100.91	144	1.24	24.18	219
2.19	9.68	3			1

**D**

Exp3a Ratio Ambra FLAG-HA/Ambra untagged	Exp3a Ratio Ambra FLAG-HA/Ambra untagged	Exp3a Ratio Ambra FLAG-HA/Ambra untagged	Exp3a Ratio Ambra FLAG-HA/Ambra untagged	Exp3a Ratio Ambra FLAG-HA/Ambra untagged	Exp3a Ratio Ambra FLAG-HA/Ambra untagged
21.36	65.25	70	0.67	41.12	106
2.54	11.90	3	1.06	0.82	2

Experiments were performed using SILAC-labeled cells as described [Antonioli et al. 2014]. Results from three independent experiments performed in duplicate are reported.



**Table S2.** Cell lines used in the study

<b>2F cell lines</b>	<b>Retroviral constructs</b>
beta-gal	MYC-beta-galactosidase
AMBRA1 FL	MYC-AMBRA1 aa 1-1296
AMBRA1 F1	MYC-AMBRA1 aa 1-532
AMBRA1 F2	MYC-AMBRA1 aa 533-751
AMBRA1 F3	MYC-AMBRA1 aa 767-1296

"Table S1: The table reports the identification of Erlin 1 as AMBRA1 interacting protein by tandem affinity pufication and LC/Orbitrap. Experiments were performed using SILAC-labeled cells as described (Antonioli et al. 2014). Results from three independent experiments performed in duplicate are reported."

Gene Names	Protein Descriptions	Uniprot	Proteins	Peptides (seq)	Sequence Coverage [%]	PEP	Exp1a Ratio Ambra FLAG-HA/Ambra untagged	Exp1a Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp1b Ratio Ambra FLAG-HA/Ambra untagged	Exp1b Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp1b Ambra FLAG-HA/Ambra untagged Count	Exp2a Ratio Ambra FLAG-HA/Ambra untagged	Exp2a Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp2a Ambra FLAG-HA/Ambra untagged Count	Exp2b Ratio Ambra FLAG-HA/Ambra untagged	Exp2b Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp2b Ambra FLAG-HA/Ambra untagged Count	Exp3a Ratio Ambra FLAG-HA/Ambra untagged	Exp3a Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp3a Ambra FLAG-HA/Ambra untagged Count	Exp3b Ratio Ambra FLAG-HA/Ambra untagged	Exp3b Ratio Ambra FLAG-HA/Ambra untagged Variability [%]	Exp3b Ambra FLAG-HA/Ambra untagged Count	
AMBRA1;KIAA1736	Isoform 1 of Activating molecule in BECN1-regulated autophagy protein 1;Isoform 3 of Activating molecule in BECN1-regulated autophagy protein 1;Isoform 2 of Activating molecule in BECN1-regulated autophagy protein 1	Q9C0C7-1;Q9C0C7:B4DZ97;Q9C0C7-3;Q9C0C7-2	3	69	58.3	0	7.26	135.70	1233	7.74	139.53	1232	77.32	100.91	144	1.24	24.18	219	21.36	65.25	70	0.67	41.12	106
ERLIN1;C10orf69;KEO4;KEO4;SPFH1;RP11-316M21.1-003	ER lipid raft associated 1;ER lipid raft associated 1	O75477;B2RDK6;B4DPN7;B0QZ43	2	9	25.6	4.01E-90	1.46	59.58	37	1.62	50.59	37	2.19	9.68	3	1	2.54	11.90	3	1.06	0.82	2		