

Supplementary Information

TORC1 Coordinates the Conversion of Sic1 from a Target to an Inhibitor of Cyclin-CDK-Cks1

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INVENTORY OF SUPPLEMENTARY INFORMATION

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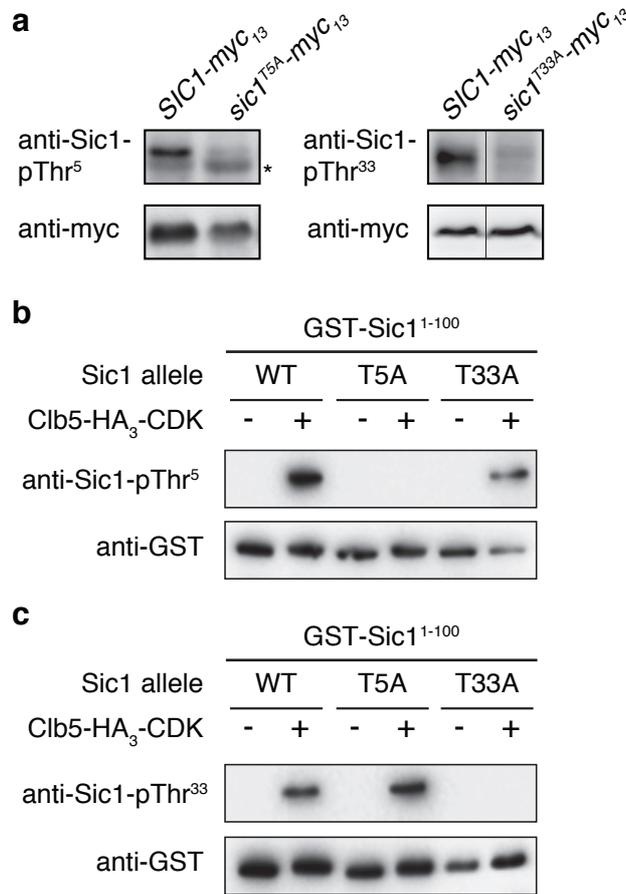


Figure S1 *In vivo* and *in vitro* specificity of the anti-Sic1-pThr⁵/pThr³³ antibodies. **(a)** Sic1-pThr⁵ and Sic1-pThr³³ levels were determined by immunoblot analyses using respective phospho-specific antibodies and extract of exponentially growing cells that expressed plasmid-encoded versions of myc₁₃-tagged Sic1 and Sic1^{T5A} (left panels), or myc₁₃-tagged Sic1 and Sic1^{T33A} (right panels). The levels of the Sic1-myc₁₃ variants were determined by using polyclonal anti-myc antibodies. The asterisk denotes an unspecific band. **(b, c)** Clb5-HA₃-CDK immunocomplexes from exponentially growing yeast cells were used for *in vitro* kinase assays in which the bacterially-purified, N-terminal parts (encompassing the first 100 amino acids) of Sic1 (WT), Sic1^{T5A} (T5A), and Sic1^{T33A} (T33A) served as substrates. Sic1-pThr⁵ (b) and Sic1-pThr³³ (c) levels were determined by immunoblot analyses using respective phospho-specific antibodies and the indicated GST-Sic1¹⁻¹⁰⁰ variants that have been phosphorylated (+), or not (-), by Clb5-HA₃-CDK. The input levels of the GST-Sic1¹⁻¹⁰⁰ variants were determined by using polyclonal anti-GST antibodies.

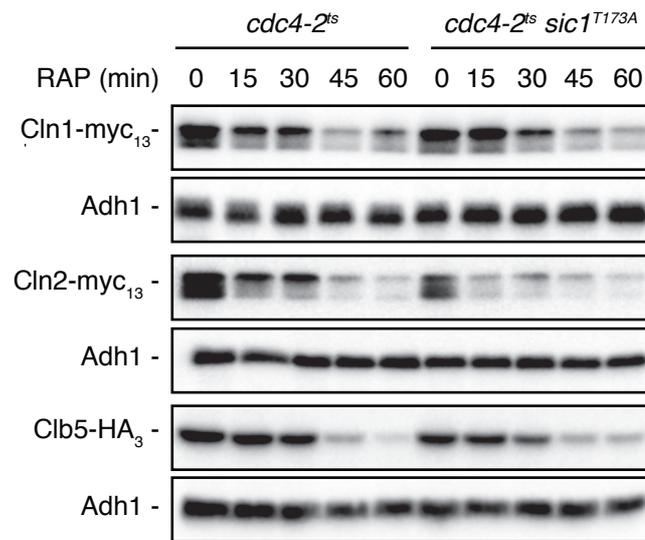


Figure S2 Sic1^{T173A}-expressing *cdc4-2^{ts}* cells are not defective in the clearance of Cln1, Cln2 or Clb5 when treated with rapamycin. *cdc4-2^{ts}* and *cdc4-2^{ts} sic1^{T173A}* strains expressing Cln1-myc₁₃, Cln2-myc₁₃, or Clb5-HA₃ were grown as in Fig. 1A. the levels of the tagged proteins were determined by immunoblot analyses using monoclonal anti-myc or anti-HA antibodies. Adh1 levels served as loading control.

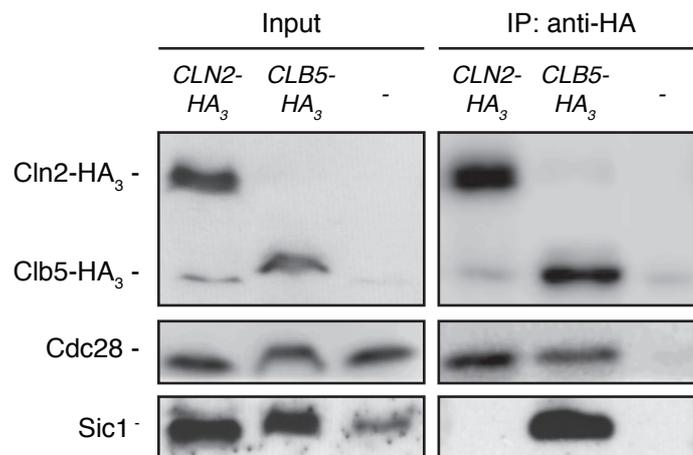


Figure S3 Cln2-HA₃ and Clb5-HA₃ interact with Cdc28 *in vivo*. Plasmid-expressed, HA₃-tagged Cln2 or Clb5 were immunoprecipitated (IPed) from extracts of exponentially growing wild-type cells. Cell lysates (Input) and anti-HA immunoprecipitates (IP: anti-HA) were analyzed by immunoblotting with anti-HA (top panels), anti-Cdc28 (panels in the middle), and anti-Sic1 (bottom panels) antibodies. Please note that both Cln2-HA₃ and Clb5-HA₃ interact with Cdc28, while only Clb5-HA₃ is able to bind Sic1 as expected. Neither Cdc28 nor Sic1 were recovered in anti-HA immunoprecipitates from extracts of cells that carried an empty plasmid (-; 3rd lanes in all panels).

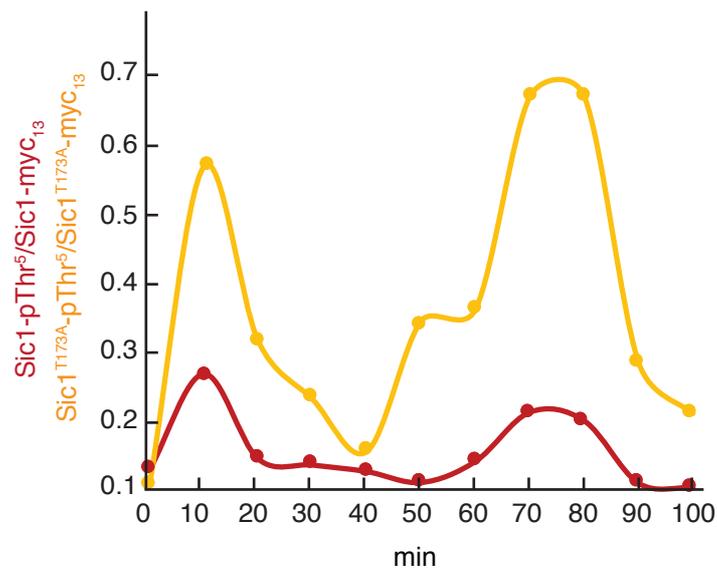


Figure S4 Proliferating, Sic1^{T173A}-expressing cells exhibit enhanced levels of Cln-/Clb5/6-CDK activity. For synchronization, exponentially growing WT cells expressing genomically-tagged Sic1-myc₁₃ or Sic1^{T173A}-myc₁₃ were treated for 2 h with α -factor (5 $\mu\text{g ml}^{-1}$). Following α -factor release, samples were collected at the indicated time points. The phosphorylation levels of Thr⁵ in Sic1 and Sic1^{T173A} were determined by using phosphospecific anti-Sic1-pThr⁵ antibodies and normalized with respect to the total levels (quantified by using anti-myc antibodies) of Sic1-myc₁₃ and Sic1^{T173A}-myc₁₃, respectively.

Supplementary Tables

Table S1. Strains used in this study

| Strain | Genotype | Source | Figure |
|------------|---|------------|-----------------------------|
| JK9-3D | <i>MATa</i> , <i>leu2</i> , <i>his4</i> , <i>trp1</i> , <i>ura3</i> , <i>rme1</i> , <i>GAL</i> , <i>HMLa</i> | Ref. (1) | 2A/B, 3B, 2C, 4G, S1B/C, S3 |
| RL343-E1 | [JK9-3D] <i>his3</i> , <i>HIS4</i> , <i>cdc28Δ</i> , pRS416- <i>cdc28^{as}</i> (F88G) | Ref. (2) | 4C |
| YMM114 | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> | Ref. (3) | 1A/C |
| YMM118-2D | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | Ref. (3) | 1A/C |
| YMM67-1C | [JK9-3D] <i>sic1Δ::kanMX</i> | Ref. (3) | 1A/B, S1A |
| YMM237-1A | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>cdc28Δ::kanMX</i> , pRS416- <i>cdc28^{as}</i> | This study | 1B/D |
| YMM250-10C | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>cdc28Δ::kanMX</i> , pRS416- <i>cdc28^{as}</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 1B/D |
| YMM246-1C | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>CLB5-HA₃::TRP1</i> , <i>CLN1-myc₁₃::kanMX</i> | This study | S2 |
| YMM247-4B | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>CLB5-HA₃::TRP1</i> , <i>CLN1-myc₁₃::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | S2 |
| YMM249-2B | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>CLB5-HA₃::TRP1</i> , <i>CLN2-myc₁₃::kanMX</i> | This study | S2 |
| YMM252-2A | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>CLB5-HA₃::TRP1</i> , <i>CLN2-myc₁₃::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | S2 |
| MJA4090 | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>CLB5-HA₃::TRP1</i> | This study | S2 |
| MJA4091 | [JK9-3D] <i>cdc4-2^{ts}::kanMX</i> , <i>CLB5-HA₃::TRP1</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | S2 |
| YMM91 | [JK9-3D] <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | Ref. (3) | 2A/B, 3B, |
| MJA490 | [JK9-3D] <i>CKS1-HA₃::kanMX</i> | This study | 2D-F, 4A |
| MJA491 | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 2D-E, 4A |
| MJA524-2B | [JK9-3D] <i>sic1^{R262A/L264A}-myc₁₃::kanMX</i> | This study | 3A |
| YMM63 | [JK9-3D] <i>SIC1-myc₁₃::kanMX</i> | Ref. (3) | 3A, 4B, S4 |
| YMM98 | [JK9-3D] <i>sic1^{T173A}-myc₁₃::kanMX</i> , <i>EMP46::natMX</i> | Ref. (3) | 3A, S4 |
| MJA523 | [JK9-3D] <i>sic1^{R262A/L264A}</i> , <i>EMP46::natMX</i> | This study | 3B |
| MJA536 | [JK9-3D] <i>cln1Δ::kanMX</i> , <i>cln2Δ::kanMX</i> | This study | 3B |
| MJA528 | [JK9-3D] <i>sic1^{T173A/R262A/L264A}</i> , <i>EMP46::natMX</i> | This study | 3B |
| YMM232-6A | [JK9-3D] <i>clb5Δ::kanMX</i> , <i>clb6Δ::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 3B |
| MJA544-2B | [JK9-3D] <i>cln1Δ::kanMX</i> , <i>cln2Δ::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 3B |
| YMM253-11A | [JK9-3D] <i>clb5Δ::kanMX</i> , <i>clb6Δ::kanMX</i> | This study | 3B |
| MJA547 | [JK9-3D] <i>clb5Δ::kanMX</i> , <i>cln1Δ::kanMX</i> , <i>cln2Δ::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 3B |
| MJA545 | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>CLB5-myc₁₃::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 3D |
| MJA546 | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>CLB5-myc₁₃::kanMX</i> | This study | 3D |
| MJA531 | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>CLN2-myc₁₃::kanMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 3E |
| MJA530 | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>CLN2-myc₁₃::kanMX</i> , | This study | 3E |
| YMM231-1A | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>mpk1Δ::kanMX</i> | This study | 4A |
| YMM230-8A | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>cdc55Δ::natMX</i> | This study | 4A |
| YMM233-3A | [JK9-3D] <i>CKS1-HA₃::kanMX</i> , <i>cdc55Δ::natMX</i> , <i>sic1^{T173A}</i> , <i>EMP46::natMX</i> | This study | 4A |
| MJA518 | [JK9-3D] <i>cdc28Δ</i> , pRS416- <i>cdc28^{as}</i> | This study | 4C |
| MJA519 | [JK9-3D] <i>cdc28Δ</i> , pRS416- <i>cdc28^{as}</i> , <i>rim15Δ::kanMX</i> | This study | 4C/H |
| YMM58-1B | [JK9-3D] <i>rim15Δ::kanMX</i> | This study | 4E/F |
| MM3D | [JK9-3D] <i>cdc28Δ</i> , pRS416- <i>cdc28^{as}</i> , <i>rim15Δ::kanMX</i> , <i>LEU2::CYC1p-HHF2-tDimer</i> | This study | 4D |

Table S2. Plasmids used in this study

| Plasmid | Genotype | Source | Figure |
|----------|--|------------|-------------------------------|
| pRS415 | CEN, <i>LEU2</i> | Ref. (5) | |
| pMJA2881 | [pRS415] <i>ADH1p-SIC1-myc₁₃</i> | This study | S1A |
| pMJA3173 | [pRS415] <i>ADH1p-sic1^{T5A}-myc₁₃</i> | This study | S1A |
| pMJA3174 | [pRS415] <i>ADH1p-sic1^{T33A}-myc₁₃</i> | This study | S1A |
| pMJA2995 | [pRS415] <i>ADH1p-SIC1¹⁵⁰⁻²⁸⁵-myc₁₃</i> | This study | 2F |
| pMJA2996 | [pRS415] <i>ADH1p-sic1^{150-285-T173A}-myc₁₃</i> | This study | 2F |
| pRS416 | CEN, <i>URA3</i> | Ref. (5) | S3 |
| pMJA3038 | [pRS416] <i>ADH1p-CLB5-HA₃</i> | This study | 2B/C, 3A, 4F/G, S3, S1B/C, S3 |
| YCplac33 | CEN, <i>URA3</i> | Ref. (6) | |
| JCE456 | [YCplac33] <i>ADH1p-CLN2-HA₃</i> | Ref. (7) | 2A/C, 4D, 4F/G, S3 |
| pAS2654 | [YCplac33] <i>ADH1p-LST7-HA₃</i> | Ref. (8) | 4F |
| pGEX | <i>GST</i> | Ref. (9) | 4G, S1B/C |
| pMMT2629 | [pGEX] <i>GST-SIC1</i> | This study | 2C |
| pMMT2630 | [pGEX] <i>GST-sic1^{T173A}</i> | This study | 2C |
| pMJA3029 | [pGEX] <i>GST-sic1^{R262A/L264A}</i> | This study | 2C |
| pMJA3037 | [pGEX] <i>GST-SIC1¹⁻¹⁰⁰</i> | This study | 2C, 3A, 4G, S1B/C |
| pMJA3219 | [pGEX] <i>GST-sic1^{T5A(1-100)}</i> | This study | S1B/C |
| pMJA3220 | [pGEX] <i>GST-sic1^{T33A(1-100)}</i> | This study | S1B/C |
| pVW995 | [pGEX] <i>GST-RIM15⁹⁴⁴⁻¹¹⁴⁹</i> | Ref. (10) | 4G |
| pVW827 | CEN, <i>LEU2</i> , <i>ADH1p-GST-RIM15</i> | Ref. (10) | 4F |
| pVW904 | 2 μ , <i>LEU2</i> , <i>TDH3p-RIM15-myc₁₃</i> | Ref. (10) | 4H |
| pVW910 | 2 μ , <i>LEU2</i> , <i>TDH3p-rim15^{T1075A}-myc₁₃</i> | Ref. (10) | 4H |
| pFD1008 | CEN, <i>TRP1</i> , <i>ADH1p-rim15^{K823Y}-GFP</i> | Ref. (10) | 4D/E |

Table S3. Antibodies used in this study

| Name | Dilution | Source |
|--------------------------------------|-----------|-------------------------|
| Anti-Sic1 | 1:1'000 | sc-50441 Santa Cruz |
| Anti-Adh1 | 1:200'000 | Calbiochem |
| Anti-Sic1-pThr ¹⁷³ | 1:1'000 | GenScript |
| Anti-Sic1-pThr ⁵ | 1:1'000 | GenScript |
| Anti-Sic1-pThr ³³ | 1:1'000 | GenScript |
| Anti-myc | 1:3'000 | 9E10; sc-40; Santa Cruz |
| Anti-HA | 1:1'000 | Enzo Life Sciences |
| Anti-GST | 1:3'000 | Bethyl Laboratories |
| Anti-Igo1-pSer ⁶⁴ | 1:1'000 | GenScript |
| Anti-Igo1 | 1:1'000 | Eurogentec |
| Anti-Cln2 | 1:1'000 | Santa Cruz |
| Anti-Cdc28 | 1:300 | Santa Cruz |
| Anti-Clb5 | 1:1'000 | Santa Cruz |
| Anti-Sch9-pThr ⁷³⁷ | 1:1'0000 | GenScript |
| Anti-Sch9 | 1:1'000 | GenScript |
| Anti-Rim15-pThr ¹⁰⁷⁵ | 1:10'000 | Eurogentec |
| Goat anti-rabbit IgG HRP | 1:3'000 | Biorad |
| Goat anti-mouse HRP | 1:3'000 | Biorad |
| Donkey anti-goat HRP | 1:5'000 | Abcam |
| Goat anti-mouse IgG-Fcy HRP | 1:5'000 | Jackson Immunoresearch |
| Goat anti-mouse IgG, light chain HRP | 1:5'000 | Jackson Immunoresearch |

Supplementary References

1. Heitman J, Movva NR, & Hall MN (1991) Targets for cell cycle arrest by the immunosuppressant rapamycin in yeast. *Science* 253(5022):905-909.
2. Bodenmiller B, *et al.* (2010) Phosphoproteomic analysis reveals interconnected system-wide responses to perturbations of kinases and phosphatases in yeast. *Sci. Signal.* 3(153):rs4.
3. Moreno-Torres M, Jaquenoud M, & De Virgilio C (2015) TORC1 controls G₁-S cell cycle transition in yeast via Mpk1 and the greatwall kinase pathway. *Nat. Commun.* 6:8256.
4. Pedruzzi I, *et al.* (2003) TOR and PKA signaling pathways converge on the protein kinase Rim15 to control entry into G₀. *Mol. Cell* 12(6):1607-1613.
5. Brachmann CB, *et al.* (1998) Designer deletion strains derived from *Saccharomyces cerevisiae* S288C: a useful set of strains and plasmids for PCR-mediated gene disruption and other applications. *Yeast* 14(2):115-132.
6. Gietz RD & Sugino A (1988) New yeast-*Escherichia coli* shuttle vectors constructed with *in vitro* mutagenized yeast genes lacking six-base pair restriction sites. *Gene* 74(2):527-534.
7. Quilis I & Igual JC (2012) Molecular basis of the functional distinction between Cln1 and Cln2 cyclins. *Cell Cycle* 11(16):3117-3131.
8. Péli-Gulli MP, Sardu A, Panchaud N, Raucci S, & De Virgilio C (2015) Amino Acids Stimulate TORC1 through Lst4-Lst7, a GTPase-Activating Protein Complex for the Rag Family GTPase Gtr2. *Cell Rep* 13(1):1-7.
9. Smith DB & Johnson KS (1988) Single-step purification of polypeptides expressed in *Escherichia coli* as fusions with glutathione S-transferase. *Gene* 67(1):31-40.
10. Wanke V, Pedruzzi I, Cameroni E, Dubouloz F, & De Virgilio C (2005) Regulation of G₀ entry by the Pho80-Pho85 cyclin-CDK complex. *EMBO J.* 24(24):4271-4278.