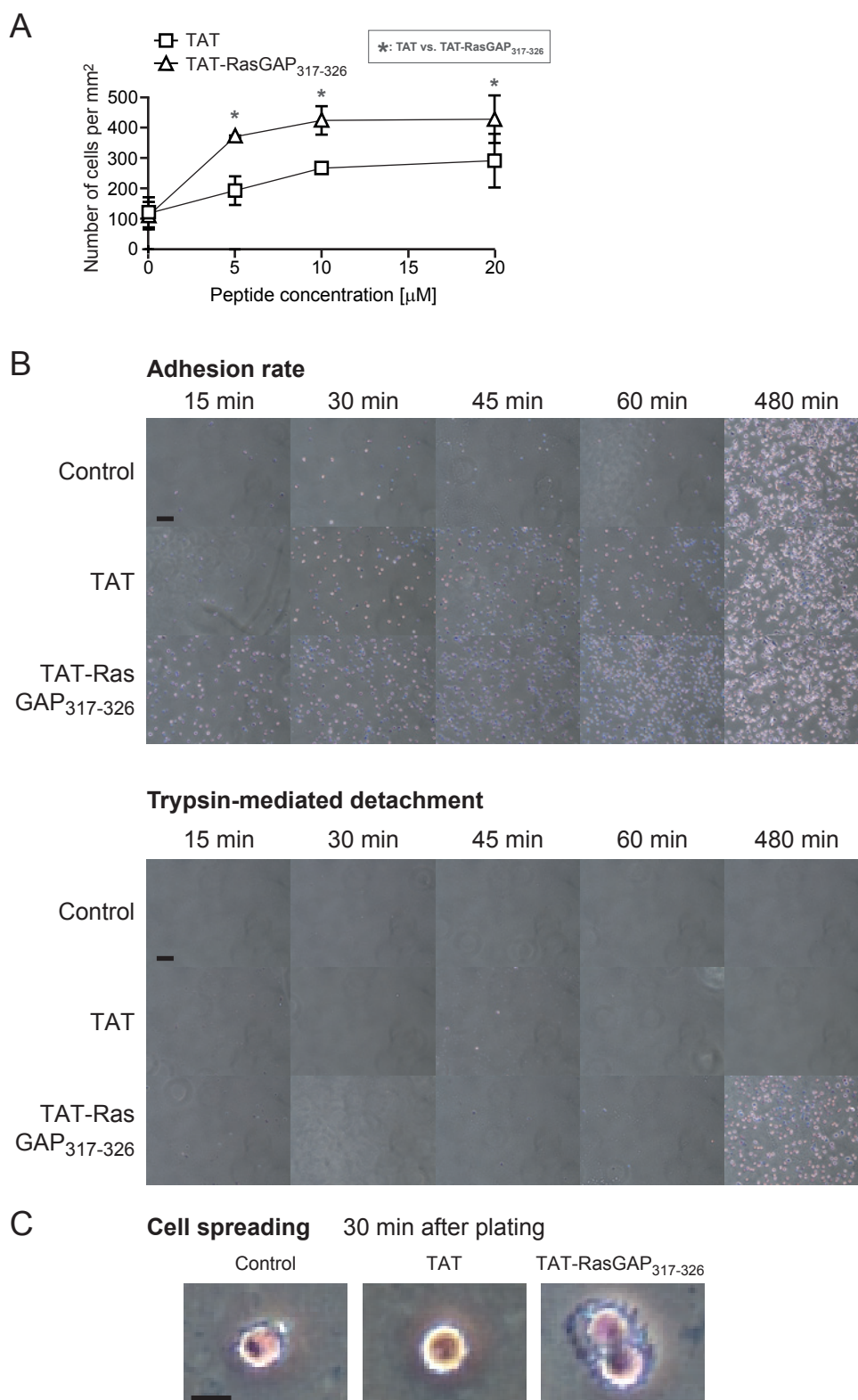


Figure S1

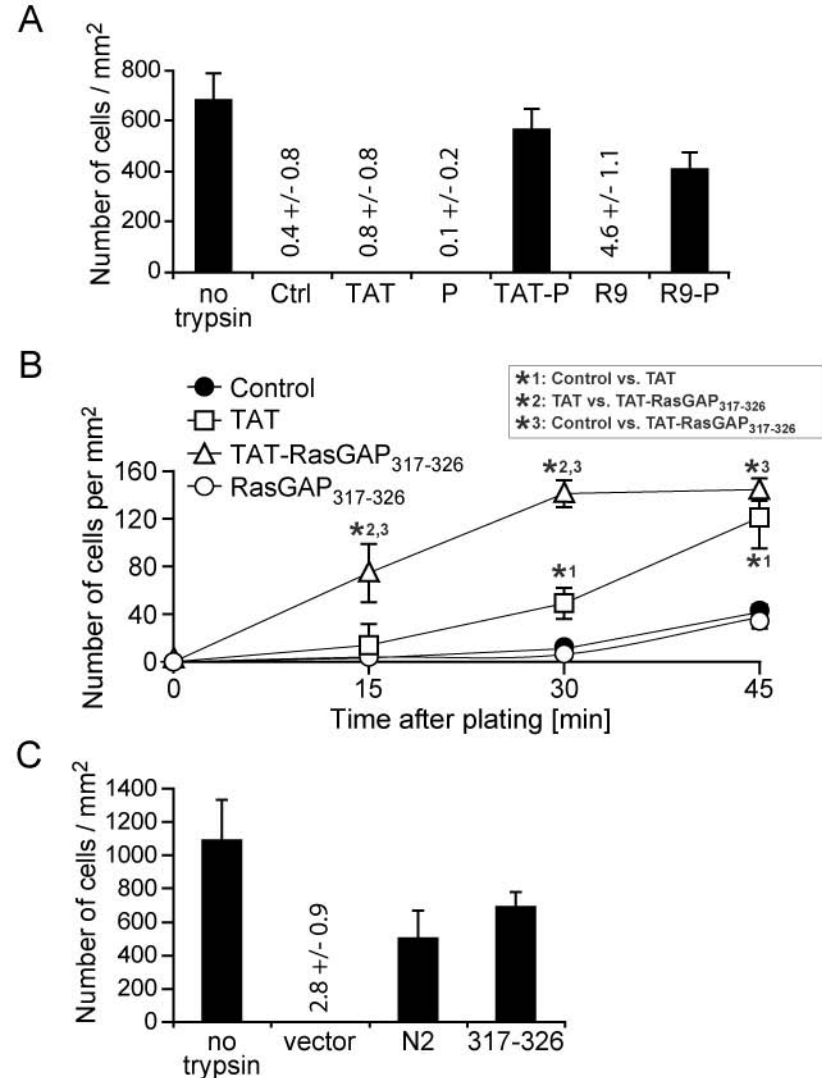


**Figure S1.** Kinetics of TAT-RasGAP<sub>317-326</sub>-mediated adhesiveness increase.

**A.** U2OS cells were subjected to an adhesion assay for 30 minutes in presence of the indicated concentrations of TAT and TAT-RasGAP<sub>317-326</sub> (the statistical significance of the observed differences was assessed by one-way ANOVA).

**B.** Representative images of the adhesion assay performed in Figure 1E. Scale bar: 100 μm.

**C.** Higher magnification of cells 30 minutes after plating. Scale bar: 40 μm.



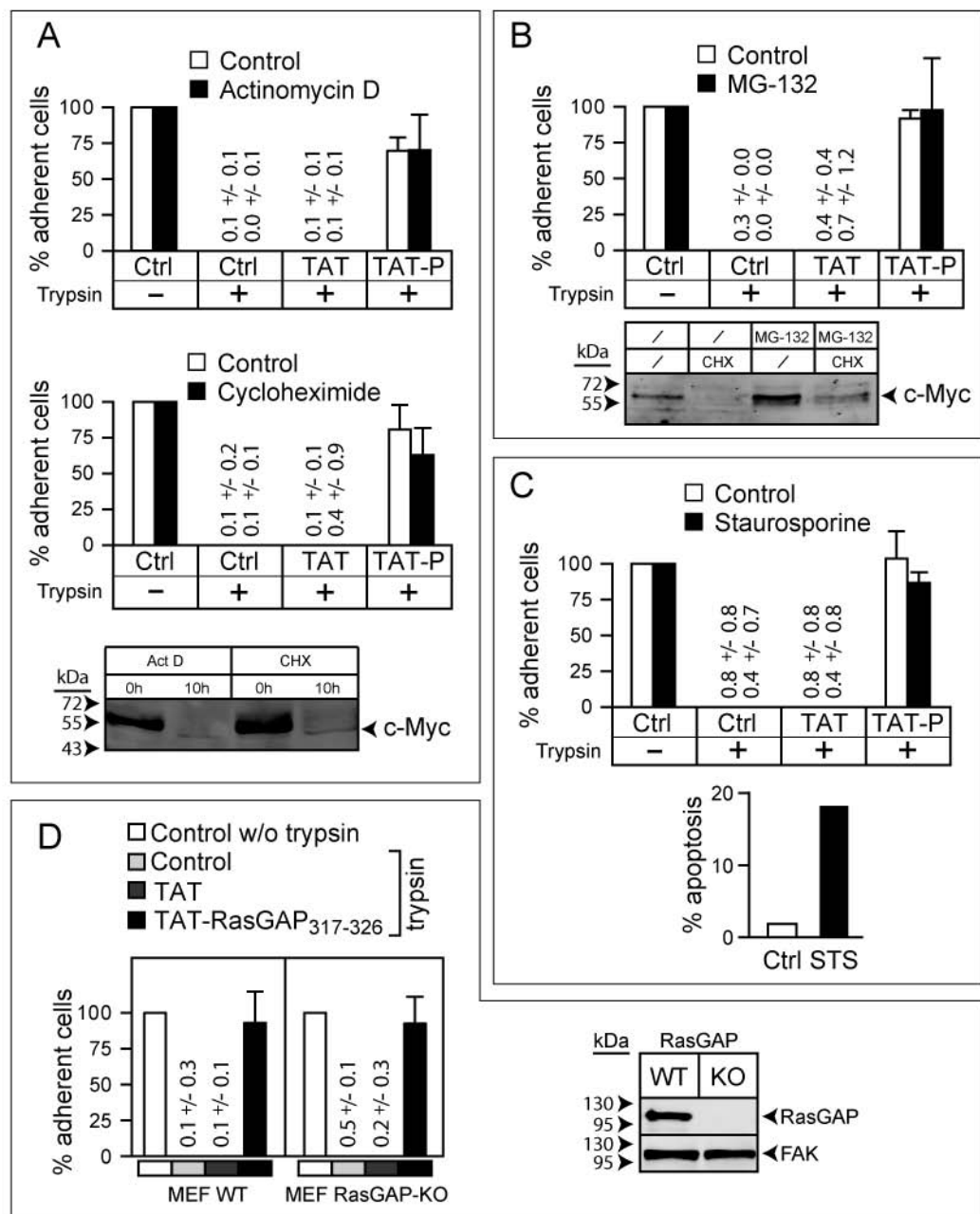
**Figure S2.** RasGAP<sup>317-326</sup> acts from inside cells.

**A.** U2OS cells were treated for 8 hours with 20  $\mu$ M TAT, 20  $\mu$ M RasGAP<sup>317-326</sup> (P), 20  $\mu$ M TAT-RasGAP<sup>317-326</sup> (TAT-P), 20  $\mu$ M R9, 20  $\mu$ M R9-RasGAP<sup>317-326</sup> (R9-P) or left untreated (Ctrl). A trypsin/EDTA-mediated detachment assay was then performed. A control without trypsin was also performed.

**B.** An adhesion assay was performed to assess the adhesion rate of U2OS cells in the presence of a RasGAP<sup>317-326</sup> peptide lacking the TAT cell permeable peptide. Treatment of cells with 20  $\mu$ M TAT-RasGAP<sup>317-326</sup> increased significantly the speed of adhesion compared to cells treated with 20  $\mu$ M TAT, 20  $\mu$ M RasGAP<sup>317-326</sup> or not treated. The statistical significance of the observed differences (asterisks) was assessed by one-way ANOVA.

**C.** HEK 293T cells were transfected with pcDNA3 (vector) or pcDNA3 encoding the HA-tagged forms of the N2 fragment of RasGAP (N2) or the 317-326 RasGAP sequence (317-326). A trypsin-mediated detachment assay was then performed but without tapping the plates. No trypsin, cells transfected with pcDNA3 that were not trypsinized.

**Figure S3**

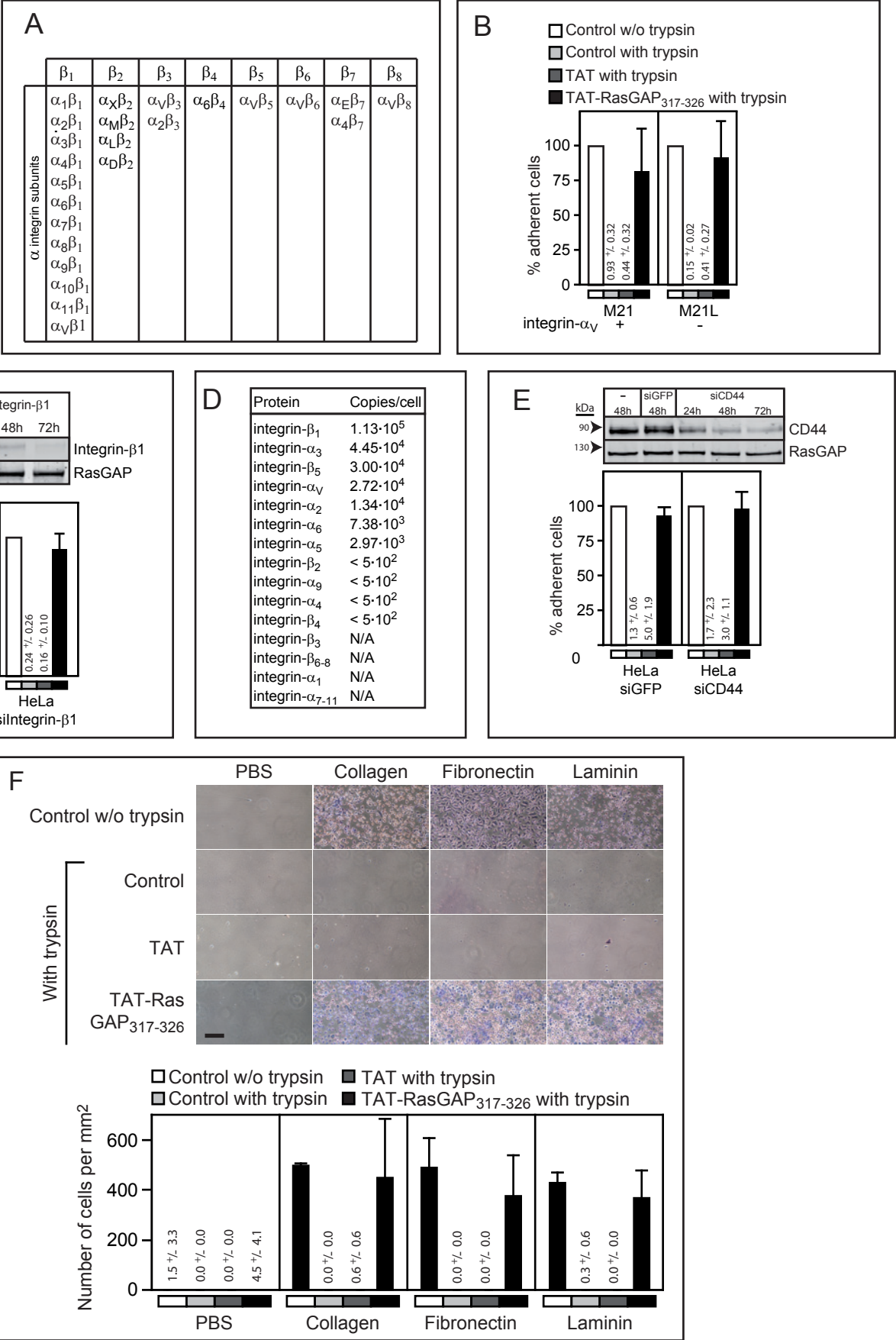


**Figure S3. TAT-RasGAP<sub>317-326</sub> does not require transcription, translation, proteasome-mediated degradation, staurosporine-inhibitable kinases, and full-length RasGAP to increase adhesion.**

**A-B-C.** U2OS cells were pre-incubated 2 hours with either 1  $\mu$ g/ml actinomycin D (Act D) or 30  $\mu$ g/ml cycloheximide (CHX) (panel A), with 10  $\mu$ M MG-132 for 16 hours (panel B), with 10 nM staurosporine (STS) for 16 hours (panel C), then treated 8 hours, in the presence of the inhibitors, with 20  $\mu$ M TAT (TAT), 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> (TAT-P) or left untreated (Ctrl). The cells were then subjected to a trypsin-mediated detachment assay. The lower panels in A and B depict c-Myc expression levels after Act D, CHX and MG-132 treatments (25  $\mu$ g of loaded proteins). The activity of staurosporine was controlled by evaluating its capacity to induce apoptosis, determined by measuring the percentage of cells with pycnotic nuclei (panel C, lower graph).

**D.** Mouse embryonic fibroblasts (MEF) or RasGAP-knock-out MEF (KO) cells were treated for 8 hours with 20  $\mu$ M TAT, 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> or left untreated. A trypsin-mediated detachment assay was performed as described for panel A-B. RasGAP expression was checked in wild-type and knock-out MEFs by immunoblotting (blot on the right-hand side of the panel).

Figure S4



**Figure S4.** Integrins, CD44 and caspases are not required for TAT-RasGAP<sub>317-326</sub>-mediated adhesion increase.

**A.** The twenty-four known integrin heterodimers are represented (8).

**B.** Wild-type (M21) or  $\alpha_V$ -integrin-null (M21L) melanoma cells were treated 8 hours with 20  $\mu$ M TAT, 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> or left untreated. The cells were then subjected to a trypsin-mediated detachment assay.

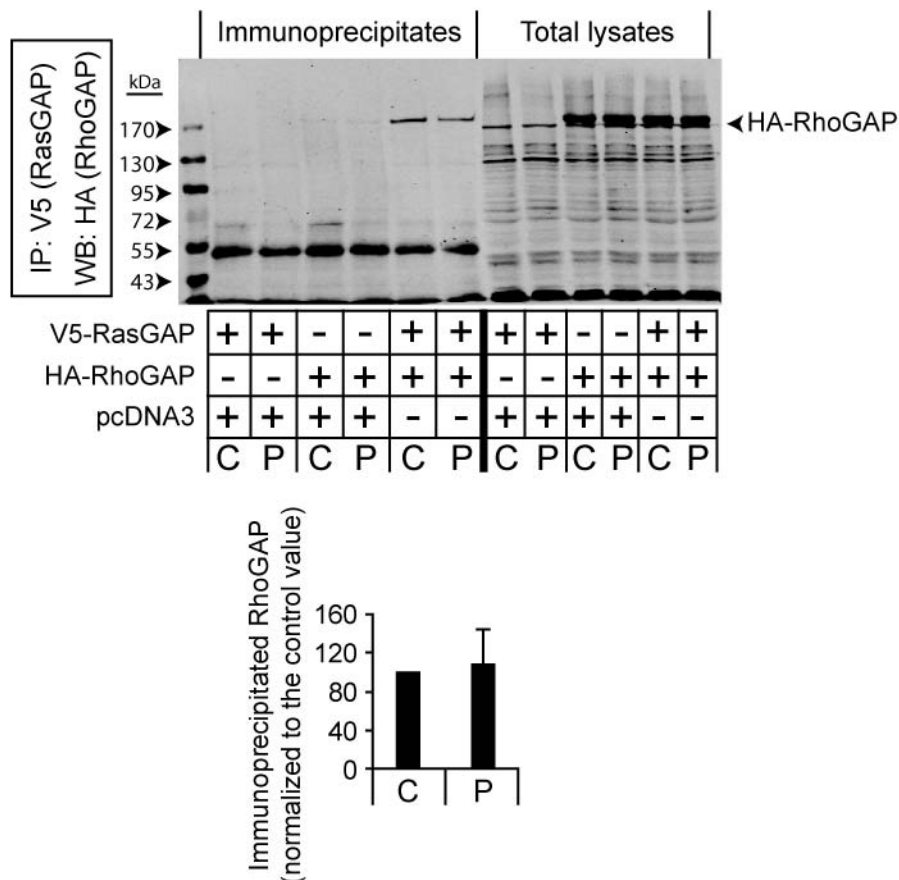
**C.** HeLa cells were transfected with siRNAs targeted against GFP (siGFP) or  $\beta_1$ -integrin (siIntegrin- $\beta_1$ ). Forty-eight hours later, the cells were incubated 8 hours with 20  $\mu$ M TAT, 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> or left untreated and were subjected to a trypsin-mediated detachment assay. The upper panel depicts the loss of  $\beta_1$ -integrin expression after siRNA transfection.

**D.** Expression levels of the indicated integrin family members in U2OS cells [data taken from (9)].

**E.** CD44 was silenced in HeLa cells with siRNAs. Forty-eight hours later, the cells were incubated 8 hours with 20  $\mu$ M TAT, 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> or left untreated. The cells were then subjected to a trypsin-mediated detachment assay. The upper panel depicts the loss of CD44 expression after siRNA transfection.

**F.** U2OS cells were grown overnight on bacteriological Petri dishes that had been coated beforehand with collagen, fibronectin or laminin. The cells were then treated or not with 20  $\mu$ M TAT or 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> for 8 hours. A trypsin-based detachment assay was finally performed. Representative images of the cells remaining attached to the plates after trypsin treatment are shown. Scale bar: 100  $\mu$ m.

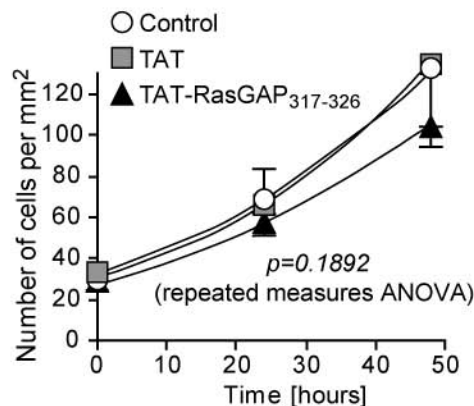
**Figure S5**



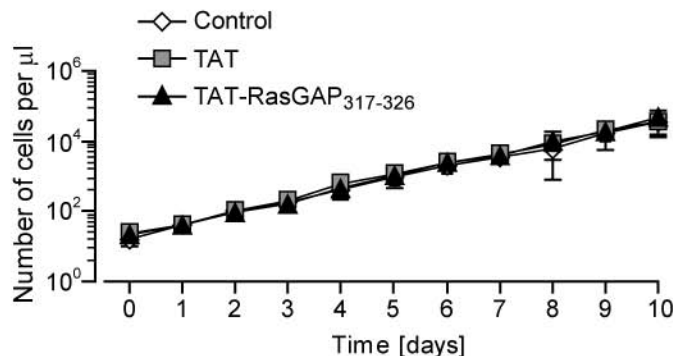
**Figure S5.** TAT-RasGAP<sub>317-326</sub> does not disrupt p120RasGAP:p190RhoGAP binding. HEK 293T cells were transfected with the indicated combinations plasmids encoding V5-RasGAP or HA-RhoGAP. Seven hundred  $\mu$ g of lysates from cells treated with 20  $\mu$ M TAT-RasGAP<sub>317-326</sub> for 24 hours (P) or left untreated (C) were immuno-precipitated with an anti-V5 antibody to precipitate exogenous p120RasGAP. Western blot against the HA-tag was performed to detect pull-down p190RhoGAP. Forty  $\mu$ g of total lysates were loaded as control. The graph represents the quantitation of the immuno-precipitated p190RhoGAP (normalized to the total lysate).

Figure S6

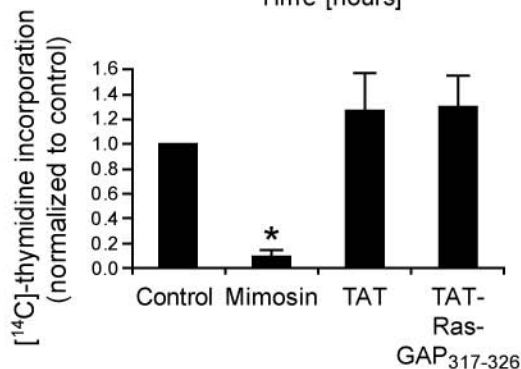
A



B



C



**Figure S6. TAT-RasGAP<sub>317-326</sub> does not affect cell proliferation**

**A.** U2OS cells were treated with 20 µM TAT, 20 µM TAT-RasGAP<sub>317-326</sub> or left untreated. The cell density was determined 0, 24 and 48 hours later. The curves were not found to differ significantly as assessed by repeated measures ANOVA.

**B.** Jurkat cells were treated with 20 µM TAT, 20 µM TAT-RasGAP<sub>317-326</sub> or left untreated, and were tested for their ability to proliferate in suspension for 10 days. The graph displays the number of cells per ml (cell splitting was performed when appropriate).

**C.** U2OS cells were treated 16 hours with 400 µM mimosine, 20 µM TAT, 20 µM TAT-RasGAP<sub>317-326</sub> or left untreated, and were tested for their ability to incorporate radioactive thymidine during 2.5 hours. Mimosine significantly altered DNA synthesis, while TAT and TAT-RasGAP<sub>317-326</sub> did not.