

Measuring multiple neurochemicals and related metabolites in blood and brain of the rhesus monkey by using dual microdialysis sampling and capillary hydrophilic interaction chromatography–mass spectrometry

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Abstract In vivo measurement of multiple functionally related neurochemicals and metabolites (NMs) is highly interesting but remains challenging in the field of basic neuroscience and clinical research. We present here an analytical method for determining five functionally and metabolically related polar substances, including acetylcholine (quaternary ammonium), lactate and pyruvate (organic acids), as well as glutamine and glutamate (amino acids). These NMs are acquired from samples of the brain and the blood of non-human primates in parallel by dual microdialysis, and subsequently analyzed by a direct capillary hydrophilic interaction chromatography (HILIC)–mass

spectrometry (MS) based method. To obtain high sensitivity in electrospray ionization (ESI)–MS, lactate and pyruvate were detected in negative ionization mode whereas the other NMs were detected in positive ionization mode during each HILIC-MS run. The method was validated for linearity, the limits of detection and quantification, precision, accuracy, stability and matrix effect. The detection limit of acetylcholine, lactate, pyruvate, glutamine, and glutamate was 150 pM, 3 μ M, 2 μ M, 5 nM, and 50 nM, respectively. This allowed us to quantitatively and simultaneously measure the concentrations of all the substances from the acquired dialysates. The concentration ratios of both lactate/pyruvate and glutamine/glutamate were found to be higher in the brain compared to blood ($p < 0.05$). The reliable and simultaneous quantification of these five NMs from brain and blood samples allows us to investigate their relative distribution in the brain and blood, and most importantly paves the way for future non-invasive studies of the functional and metabolic relation of these substances to each other.

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Introduction

The physiological function of neurons and the correct information processing within the brain depend critically on neuroactive molecules and metabolites (NMs). These NMs consisting of neurotransmitters like glutamate or neuro-modulators like acetylcholine can be detected by appropriate techniques in the extracellular fluid of the brain. The concentrations of these NMs are tightly controlled and

stable within narrow boundaries. Many disorders of the central nervous system (CNS) such as Parkinson's disease [1], Alzheimer's disease [2], and schizophrenia [3], are correlated with or even caused by changes in the concentrations of such substances. NMs can be sampled from the extracellular brain fluid (EBF) which allows us to investigate their dynamic reflecting changes in neuronal activity. The EBF forms a complex, dynamic environment embedding all neural elements and serves as medium for the cell-to-cell communication and metabolite transport. Many of these NMs also exist in the blood system, but their concentrations are commonly different from those in the brain because of the differential permeability of the blood brain barrier (BBB) and of differences in absorption mechanisms, synthesis, and metabolism. Simultaneous and quantitative monitoring of these NMs from the brain and the blood system will therefore allow us to assess their relation between the brain and the blood system.

In the present study, we focused on a selection of NMs functionally and metabolically related and developed a direct and sensitive analytical method to monitor acetylcholine, lactate, pyruvate, glutamine, and glutamate in parallel from the brain and the blood. All these five substances are derivatives from glucose and the citrate cycles [4, 5]. Additionally, acetylcholine is able to modulate glutamatergic release and can interact with the brain energy metabolism [6–8]. To do so, we used microdialysis to collect samples from EBF and blood of anesthetized non-human-primates in parallel. This will allow us to investigate changes in their relation caused by functional and metabolic challenges or by pathologies in the future. However, their chemical and physical properties are different which makes it difficult to separate and detect these NMs together. To the best of our knowledge, there is currently no report describing the simultaneous determination of all five aforementioned substances by a single analysis. Indeed, so far, most analytical methods developed were only focusing on the determination of one or two of such targeted substances. For example, electrochemical detection (ECD) is commonly used to monitor these compounds [9–11], but an additional enzyme reactor is needed to generate electrochemically detectable hydrogen peroxide for acetylcholine detection [12], so it is difficult to monitor all these substances in a single analytical run using ECD as detector. Compared to the classical ECD, mass spectrometry (MS) is able to detect diverse NMs with high sensitivity and selectivity. Recent analytical advances have been made by using MS detection coupled with ion-pair reversed-phase liquid chromatography (RPLC) or ion exchange chromatography (IEC) as separation tools. For instance, acetylcholine has been analyzed by strong cation exchange (SCX)-MS [13] or ion-pair RPLC-MS [14, 15]. Pyruvate was analyzed by weak anion exchange (WAX)-MS [16], while

lactate and pyruvate together were detected by RPLC-ESI/MS (electro spray ionization, ESI), but a time-consuming derivatization process with 3-nitrophenyl-hydrazine was needed [17]. Glutamine and glutamate were determined by ion-pair RPLC-MS using an ion-pairing reagent which might cause ESI suppression [18].

In contrast to ion-pair RPLC or IEC-based analytical approaches, hydrophilic interaction liquid chromatography (HILIC)-MS is an alternative technique that allows separating and determining multiple polar analytes irrespective of being a base or an organic acid [19, 20]. It already has proved its merits in determining peptides, proteins and small polar molecular compounds in complex matrices [21–24]. Recent reports demonstrated that HILIC-MS is well suited to determine polar neurotransmitters in EBF offering high sensitivity combined with time-saving procedures for sample preparation [23, 25].

In this study, we report the successful combination of capillary HILIC-ESI/MS with dual microdialysis sampling for simultaneous measurement of five NMs together in brain and blood dialysates of non-human primates. First, our approach allows us to compare the relation of these NMs to each other under different physiological and pathological states of the brain. Second, the simultaneous sampling from the brain and the blood can be used to extrapolate from blood measurements to brain concentrations of these NMs. Relation changes of these NMs detectable in the blood can serve as easy accessible biomarkers reflecting physiological or pathological states of the brain [26].

Experimental

Chemicals

HPLC-MS grade acetonitrile, formic acid, and ammonium formate were purchased from Sigma-Aldrich (Munich, Germany). [3-¹³C] lactate (as aqueous solution, 20% *w/w* in H₂O) (¹³C-lactate), glutamine-L-2, 3, 3, 4, 4-d₅ (glutamine-d₅), and glutamic acid-L-2, 3, 3, 4, 4-d₅ (glutamate-d₅) were purchased from Cambridge Isotope Laboratories (Andover, USA). Acetylcholine-*N,N,N*-trimethyl-d₉ chloride (acetylcholine-d₉) was bought from Medical Isotopes, Inc (Pelham, USA). Acetylcholine chloride, sodium lactate (lactate), sodium pyruvate, glutamine, glutamate, [1-¹³C] sodium pyruvate (¹³C-pyruvate), and the chemicals for preparing frits were all obtained from Sigma-Aldrich. Pure water was produced by a TKA superpure water system (Niederelbert, Germany). Artificial cerebral spinal fluid (a-CSF) for standard solutions consisted of 148 mM NaCl, 3.00 mM KCl, 0.80 mM MgCl₂, and 1.40 mM CaCl₂, pH 7.3.

Fabrication of capillary columns

The preparation of capillary columns was adopted and modified from Zhang et al. [23]. To prevent the packing material from spilling out of the column a frit was formed at one end by in situ free radical polymerization modified from Viklund et al. [27]. Compared to the frit formed by sintered reaction, our frit can easily withstand long exposure to high pressure (>3,000 psi) and minimize extra-column band broadening. After the reaction, unreacted material and other soluble compounds were removed from the pores by washing with ethanol followed by purging with helium for 15 min. The column was packed with acetone slurry (1.5 mL) containing 50 mg of 5 μm polyhydroxyethyl aspartamide particles (PolyLC, USA) with the help of a helium pressure cell with a stirring bar to maintain particle suspension running under a pressure of 100 bars for 30 min. The packed column was cut to 15 cm for use.

Preparation of calibration standards and internal standards

To increase the reproducibility of the quantitative analysis of acetylcholine, lactate, pyruvate, glutamine, and glutamate, acetylcholine- d_9 , ^{13}C -lactate, ^{13}C -pyruvate, glutamine- d_5 , and glutamate- d_5 were used as their respective internal standards (IS). Stock solutions of the analytes were prepared in 30% acetonitrile containing 0.1% formic acid and stored at 5 °C. To characterize the analytical performance of capillary HILIC-MS methods for the quantitative measurement of the analytes, the stock solution was serially diluted using a solution containing a-CSF/solution A (90% acetonitrile, 0.1% formic acid; 1:4, v/v). The calibration solutions of acetylcholine ranged from 0.5–250 nM and consisted of three levels (low, medium, and high) of quality control (QC) samples with 2.5, 25, and 125 nM. For lactate and pyruvate, the calibration solutions ranged from 10–5,000 μM , and QC samples were 50, 500, and 2500 μM . For glutamine, the calibration solutions ranged from 0.5–250 μM , and QC samples were 2.5, 25, and 125 μM . For glutamate the calibration solutions ranged from 0.25–125 μM , and QC samples were 1.25, 12.5, and 62.5 μM . Two IS stock solutions were prepared in water containing 0.1% formic acid: one (IS₁) is used for the calibration curve preparation and the blood microdialysate samples analysis, which contains 500 nM acetylcholine- d_9 , 10 mM ^{13}C -lactate, 10 mM ^{13}C -pyruvate, 500 μM glutamine- d_5 , and 250 μM glutamate- d_5 ; the other one (IS₂) is used for the monkey brain microdialysate samples analysis containing 300 nM acetylcholine- d_9 , 6 mM ^{13}C -lactate, 6 mM ^{13}C -pyruvate, 300 μM glutamine- d_5 , and 150 μM glutamate- d_5 . These internal standards were diluted ten times (IS₁) and six times (IS₂) on the day of

the assay using a-CSF/solution A (90% acetonitrile, 0.1% formic acid; 1:4, v/v). The calibration curve was constructed based on the response ratio of the peak area ($P_{\text{analyte}}/P_{\text{IS}}$) versus the nominal standard concentrations by a least-squares regression using a weighting factor of $1/\text{concentration}^2$.

Microdialysis sampling

Two male rhesus monkeys (*Macaca mulatta*) were used with an age of 5 years weighing between 6 and 7 kg. The experimental procedures were approved by the local authorities (in German: “Regierungspraesidium Tuebingen”) and were in agreement with guidelines of the European Community (EUVD 86/609/EEC) for the care and use of laboratory animals. During the experiment, the monkeys were anesthetized with full monitoring of all vital parameters. Before the experiment, the monkeys were sedated with ketamine (15 mg/kg). Anesthesia was induced with fentanyl (31 $\mu\text{g}/\text{kg}$), thiopental (5 mg/kg), and succinyl chloride (3 mg/kg). Afterwards, the monkeys were intubated and ventilated with a Servo Ventilator 900 C (Siemens, Germany). The maintenance of anesthesia was attained with remifentanyl (0.2–1 $\mu\text{g}/\text{kg}/\text{min}$) and mivacurium chloride (4–7 mg/kg/h). A crystalloid solution (Ionosteril, Fresenius Kabi, Germany) with 2.5% glucose was infused at a rate of 10 ml/kg/h. During the experiment, body temperature was kept between 38.5 and 39.5 °C, SpO_2 above 95% and end tidal CO_2 at 35 mmHg.

The monkeys had a miniaturized PEEK chamber over the primary visual cortex that gave access to the brain. The probe was inserted through a small incision in the dura. The probe for blood sampling was placed into a superficial leg vein using an intravenous catheter.

For brain sampling, microdialysis probes CMA 12 with 2 mm PAES membrane of 0.5 mm outer diameter (o.d.) and a 20 kDa cut-off (CMA Microdialysis AB, Sweden) were used. The probe was flushed with a-CSF solution at a flow rate of 2 $\mu\text{L}/\text{min}$ for 15 min, and then conditioned at a flow rate of 600 nL/min for 2 h before insertion into the cortex of the monkey. For the blood sampling, a microdialysis catheter CMA 64, 20 mm PAES membrane, 0.6 mm o.d., 20 kDa cut-off (CMA Microdialysis AB, Sweden) was flushed with saline containing fragmin 25 IU/mL (Pfizer Pharma GmbH, Germany) at a flow rate of 2 $\mu\text{L}/\text{min}$ for 2 h before insertion. The collections of microdialysis samples were started 2 h after the insertion of the probes for blood sampling as well as for brain sampling.

The dialysates were collected in parallel by two refrigerated fraction collectors CMA 740 (CMA Microdialysis AB, Sweden) at 6 °C for 1 h. The samples were collected at an interval of 3 min for both brain and blood sampling. We collected simultaneously five brain and five

blood samples each time. The sampling experiments were repeated two times from each animal. Before sampling, the IS solution was added to the glass inserts used for sample collection and then put into the refrigerated fraction collectors. After the experiment, the brain microdialysate samples were concentrated by a vacuum concentrator (Eppendorf, Germany) under room temperature for 5 min and reconstituted by solution B (60% acetonitrile, 0.1% formic acid) for further analysis. The blood microdialysate samples were directly diluted by solution C (80% acetonitrile, 0.1% formic acid) and then analyzed.

In vitro recovery of CMA 12 probes has been tested in our lab, and it was 50.2–66.8% for all analytes with a reliable stability not exceeding 3.6% standard errors. The measured in vitro recovery of CMA 64 probes for all analytes was 42.5–49.3% with good reliable stability not exceeding 4.3% standard errors.

Capillary liquid chromatography and mass spectrometry

An Agilent capillary HPLC 1100 system coupled to a XCT plus ion-trap mass spectrometer (Agilent, Waldbronn, Germany) was used for analysis. Mobile phase A was an aqueous solution containing 50 mM ammonium formate and 1.0% formic acid (pH 2.87), and mobile phase B was acetonitrile. The analysis was performed using a gradient profile: 0.0–10.0 min, 85% B to 45% B, then 45% B was kept for 5 min. The column was reconditioned using an 85% solution of B for 10 min before further injection. The flow rate was 3.5 $\mu\text{L}/\text{min}$, and a sample of 0.3 μL was injected in the column each time.

MS analysis was performed by switching between positive and negative ion mode. Ion-trap tandem mass spectrometry (MS/MS) experiments were applied to isolate and fragment the precursor ions. The mass spectrometer was operated in MRM mode (multiple-reaction-monitoring) for the quantitative analysis. Five scans were averaged for each spectrum with the scan range of m/z 50–2,000. MS scans were split into three segments considering the different MS behaviors of five compounds. In the first segment (0.0–3.5 min) acetylcholine and acetylcholine- d_9 were monitored under a positive mode with the capillary voltage of 3,500 V, while lactate, ^{13}C -lactate, pyruvate and ^{13}C -pyruvate were monitored under a negative mode in the second segment (3.5–8.6 min) with the capillary voltage of 2,541 V, and glutamine, glutamine- d_5 , glutamate, and glutamate- d_5 were monitored under a positive mode in the third segment (8.6–15.0 min) with capillary voltage of 3200 V. The MS parameters of five analytes and their corresponding internal standards are listed in Table 1. Lactate and pyruvate did not generate significant and stable product ions, so we could not use the product ions for lactate and pyruvate in MRM conditions. We used ion

fragmentation amplitude of 0.5 V and ion fragmentation cut-off of 58 which reduced the background noise keeping the lactate and pyruvate unaffected. This procedure could compensate for the lack of specificity which is achieved by monitoring the transfer from the precursor ion to the product ion. The MS settings such as capillary exit, skimmer and lens voltages were optimized and tuned by the data acquisition software during infusion of a standard solution for each compound.

Results and discussion

HILIC-MS analytical method development

Highest intensities for all the analytes were achieved when both the positive and the negative ionization mode of the mass spectrometer were used in the ESI experiments. Previous studies showed that chemicals can have different ESI/MS responses because of their different physical and chemical properties [28]. In our case, lactate and pyruvate, which are organic acids, had much higher intensities in negative ionization mode than in positive ionization mode. The other analytes, acetylcholine, glutamine and glutamate generated responses with high intensity only in positive ionization mode. For our experiments, we thus used positive ionization mode for detecting acetylcholine, glutamine, and glutamate, while negative ionization mode for lactate and pyruvate. In the analytical experiments, we selected HILIC as the method to separate the five polar analytes. Our initial tests showed that HILIC was able to separate them simultaneously (data not shown). The elution order was acetylcholine, lactate, pyruvate, glutamine, and glutamate. Because both positive and negative ionization mode were used for ESI/MS detection, the efficient separation of lactate and pyruvate from the other analytes has to be achieved to ensure that each analyte reliably matches the time window of the appointed ionization mode.

To achieve optimal separation and intensity of the five analytes on the HILIC-MS, we examined the influence of pH value, water content and buffer concentration of the mobile phase on the retention behavior and MS signal of each analyte. Figure 1a shows that the pH value had clearly stronger effects on lactate and glutamate compared to the other three analytes when we varied the pH of mobile phase A from 2.7–4.2 by adjusting formic acid proportions from 0.05–1.50%. In particular, the elution order of lactate and pyruvate reversed when pH value ranged from 3.9–2.7. This may be because lactate is ionized at higher pH values compared to its $\text{p}K_a$ of 3.85. Figure 1d indicates that the MS intensity of the five analytes was also affected by the pH value. Their intensities were on average higher at lower pH value; 1.0% formic acid (pH 2.9) was finally used for

Table 1 MS parameters for five analytes and their internal standards

Compounds	MRM transition	Width	Frag. cut-off (m/z)	Frag. amplitude (V)	ICC smart target (m/z)	ICC accumulation time (ms)
Ach	146→87	1.0	62	1.38	200,000	200
Ach-d ₉	155→87	1.0	62	1.38	200,000	200
Lac	89→89	1.0	58	0.50	200,000	100
¹³ C-Lac	90→90	1.0	58	0.50	200,000	100
Pyr	87→87	1.0	58	0.50	200,000	100
¹³ C-Pyr	88→88	1.0	58	0.50	200,000	100
Gln	147→130	1.0	69	1.20	50,000	200
Gln-d ₅	152→135	1.0	69	1.20	50,000	200
Glu	148→130	1.0	60	0.98	50,000	200
Glu-d ₅	153→135	1.0	60	0.98	50,000	200

the following optimization warranting efficient separation combined with high intensity.

The water content had strong influence on the chromatographic behaviors and MS signal intensities of all five analytes. When the water content was decreased to 30% or lower, lactate and pyruvate could be very well separated from their neighboring peaks of acetylcholine and glutamine in the chromatogram (Fig. 1b). Figure 1e illustrates that the reduced water content (from 60–30%) could enhance the MS intensity, which is the benefit of the

increased electrospray ionization efficiency in a high organic phase. On the other hand, a MS signal decline was observed when the water content ranged from 30% to 20%. This might result from the low solubility of the polar compounds in the high organic phase mobile.

During the method development, we found that the salt content in the buffer solution had a great impact on the peak shapes and MS signal of the analytes (Fig. 1f), but less effect on retention time (Fig. 1c). Sharp peak shapes and high reproducible retention times were obtained for all

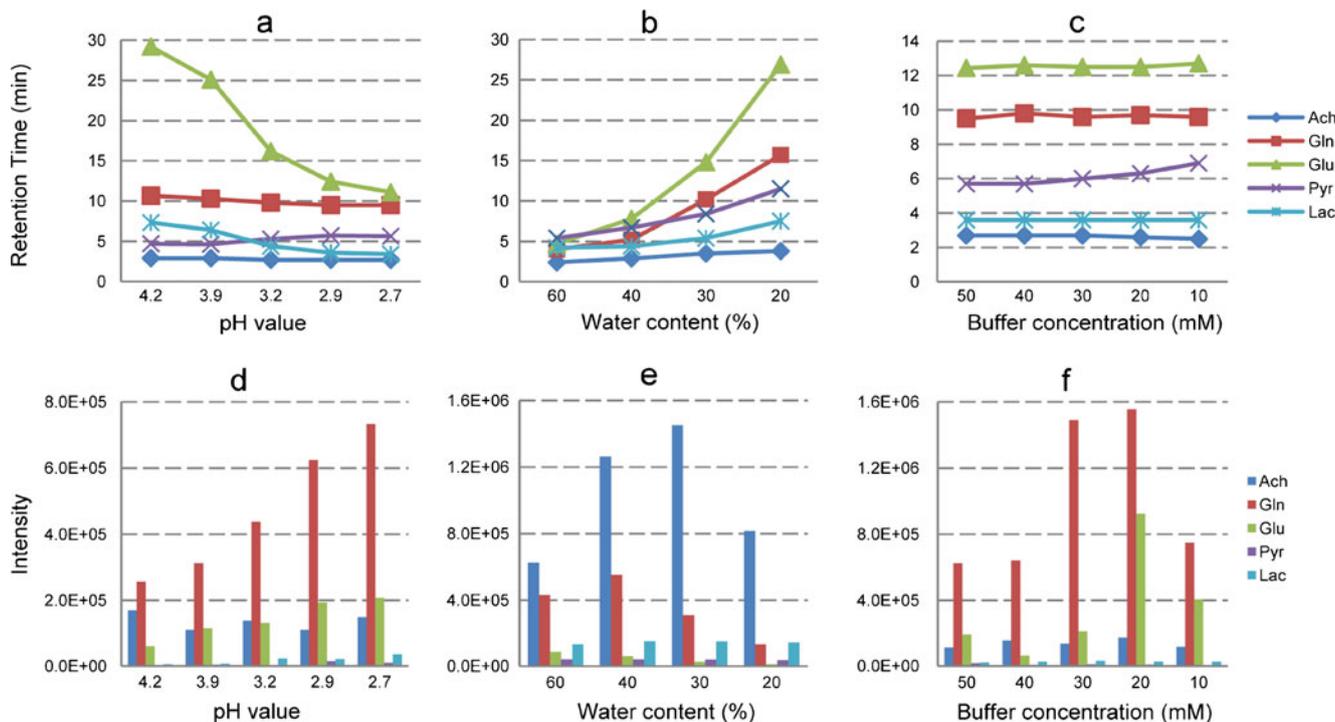


Fig. 1 Influences of mobile phase on retention behavior and MS signal of the five analytes. **a** Effects of pH value on the retention time. **b** Effects of water content on the retention time. The aqueous phase containing 50 mM ammonium formate and 1.0% formic acid. **c** Effects of buffer concentration on the retention time. Mobile phase A containing 1.0% formic acid. **d** Effects of pH value on MS signal. **e**

Effects of water content on MS signal. **f** Effects of buffer concentration on MS signal. An isocratic elution of mobile phase A: mobile phase B (15:75, v/v) was used to test the influence of pH value and buffer concentration. *Acetylcholine (*Ach*), Lactate (*Lac*), Pyruvate (*Pyr*), Glutamine (*Gln*), Glutamate (*Glu*)

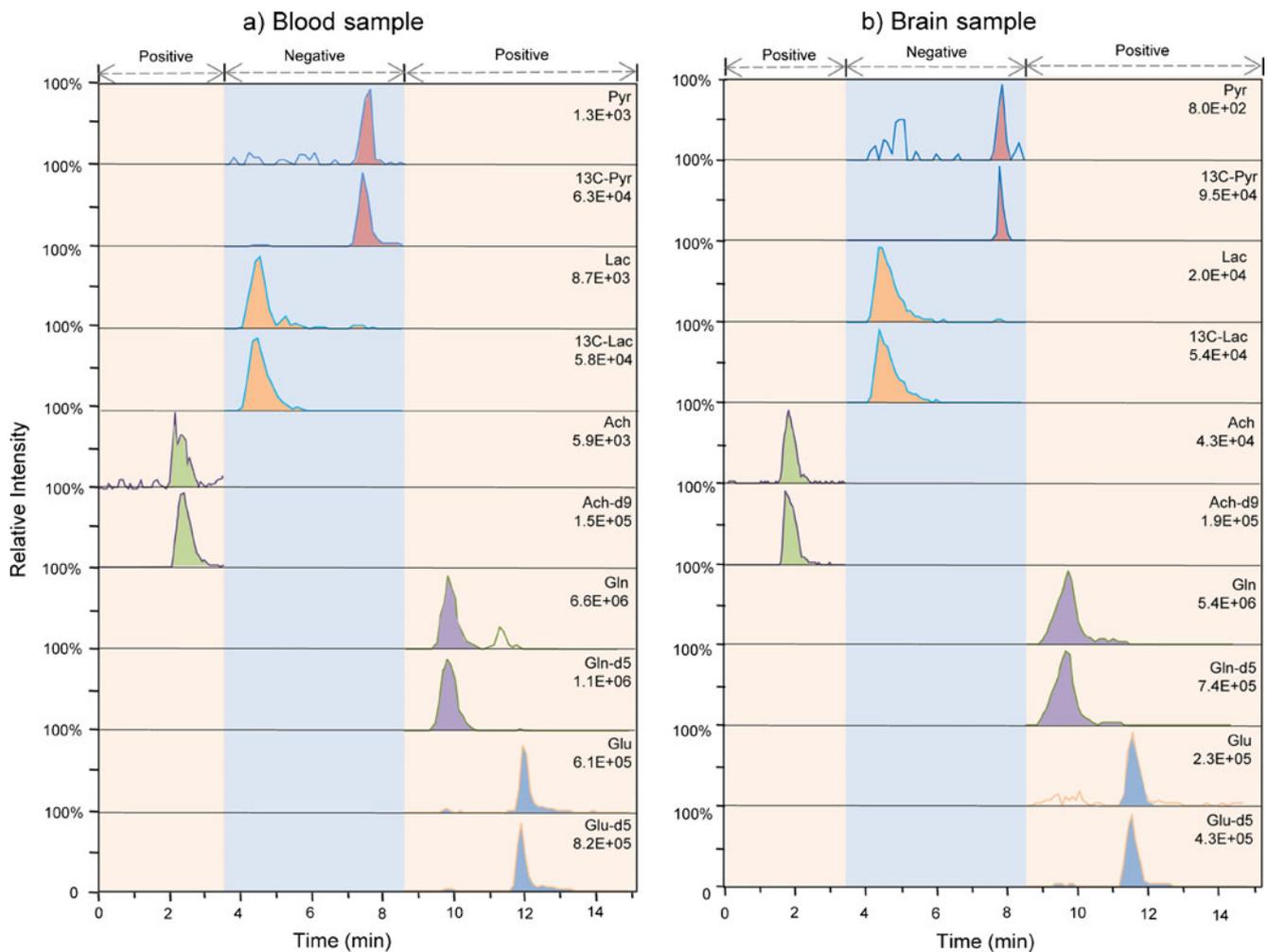


Fig. 2 Representative HILIC-ESI/MS chromatograms of five NMs under the optimized conditions **a** blood sample **b** brain sample. Both positive and negative ion mode was adopted for the detection of five analytes during each run

analytes when the ammonium formate concentration was increased from 10–50 mM. Therefore, a higher concentration of buffer solution was used for the elution, although ion suppression could be observed at this level.

Taken together, the aqueous phase consisted of a buffer solution containing 50 mM ammonium formate and 1.0%

formic acid using a gradient profile that started at 85% acetonitrile for separation of the targeted analytes. This optimized separation method ensured that each analyte was matching the appointed time window of the chosen ionization mode without any interferences. The typical HILIC-ESI/MS chromatograms containing the five NMs

Table 2 Validation results of the HILIC-ESI/MS method for the analysis of the five neurochemical analytes

Compounds	LOD	LOQ	Linear range	r^2	Accuracy (%)	Precision ^a	
						Intra-day RSD%, $n=5$	Inter-day RSD%, $n=3$
ACh	150 pM	450 pM	0.5–250 nM	0.9989	82.6–109.0	11.6	12.8
Lac	3 μ M	10 μ M	10–5000 μ M	0.9990	86.0–106.8	9.6	15.2
Pyr	2 μ M	6 μ M	10–5000 μ M	0.9989	85.6–117.4	10.5	14.3
Gln	5 nM	15 nM	0.5–250 μ M	0.9990	81.5–117.8	10.3	13.3
Glu	50 nM	150 nM	0.25–125 μ M	0.9988	86.0–106.9	14.6	15.0

^a Intra-day and Inter-day precisions were analyzed by one-way ANOVA analysis.

Table 3 Concentrations of acetylcholine, lactate, pyruvate, glutamine, and glutamate in the dialysates of brain and blood

Compounds	Ach (nM)	Lac (μ M)	Pyr (μ M)	Gln (μ M)	Glu (μ M)
Brain	4.0 \pm 1.4	220.4 \pm 90.9	21.3 \pm 8.3	50.4 \pm 21.9	1.1 \pm 0.2
Blood	10.3 \pm 4.4	511.3 \pm 78.7	91.5 \pm 37.6	176.4 \pm 45.3	26.7 \pm 5.3

Anesthetized monkeys, resting status. Values are given as mean \pm standard error of the mean (S.E.M.); $n=4$.

(acetylcholine, lactate, pyruvate, glutamine, and glutamate) are shown in Fig. 2.

Analytical performance of the capillary HILIC-ESI/MS method

The capillary HILIC-ESI/MS method was validated for the quantitative measurements of acetylcholine, lactate, pyruvate, glutamine, and glutamate concentrations from in vivo dialysates. We characterized a series of parameters like the limits of detection (LODs), limits of quantification (LOQs), linearity, selectivity, accuracy, precision and stability by analyzing different levels of standard mixtures of the target NMs while using optimized MRM transitions and positive/negative ionization modes. The validated results of the analytical performance are listed in Table 2. The LODs of lactate and pyruvate in negative mode were 3 and 2 μ M, respectively, whereas the LODs of acetylcholine, glutamine, and glutamate in positive mode were 150 pM, 5 nM, and 50 nM, respectively. Although the LODs of lactate and pyruvate were much higher than those of the other three analytes (acetylcholine, glutamine, and glutamate) we still can measure them, because their basal concentrations in the brain as well in the blood are much higher than the other three. The matched MS/MS pattern between the endogenous NMs and their standards allowed confirming the high selectivity of the HILIC-MS method. Additionally, the retention time of the endogenous NMs always matched well with those of internal standards under different gradient elution conditions, which also proved the high selectivity of the method. The accuracy, precision and stability have been tested by replicate analysis (five determinations per concentration) of three different levels of QCs (see the section “Preparation of calibration standards and internal standards”). It is shown in Table 2 that the results of the accuracy, intra- and inter-day precisions were all in good agreement with the requirements of biological analysis [29, 30]. The stability test results indicated that the stock solution was stable for 1 month at -20°C with the RSD < 12.5%, and the QCs were stable for about 2 weeks in the autosampler at 4°C with the RSD < 15.1%. Additionally, the freeze (-20°C) and thaw stability has also been tested for these analytes. After two freeze-thaw cycles, the QCs were analyzed on the third cycle, and the results showed a good stability with the RSD < 13.1%.

In addition, we adopted the post-extraction spike method proposed by Matuszewski et al. [31] to evaluate the matrix effect of the acquired in vivo samples in our study. To reliably determine the endogenous concentration of the five analytes we analyzed them according our developed method and then checked the same brain and blood samples by spiking with appropriate standard solutions. The spiked concentration of acetylcholine, lactate, pyruvate, glutamine, and glutamate was 8 nM, 150 μ M, 10 μ M, 40 μ M, and 1 μ M respectively for the brain samples, while 20 nM, 400 μ M, 60 μ M, 300 μ M, and 30 μ M for the blood samples. The total concentrations of the five analytes in the spiked samples and the endogenous concentrations in the non-spiked samples were then determined and used to calculate the recovery of each analyte. We used the following formula for calculating the recovery: (measured value–endogenous value)/added value \times 100. The obtained recoveries for the five analytes fell in the range of 93–112%.

Concentrations of acetylcholine, lactate, pyruvate, glutamine, and glutamate in the brain and blood microdialysates

We applied capillary HILIC-ESI/MS method together with simultaneous blood and brain microdialysis sampling to measure the concentrations of five NMs from two anesthe-

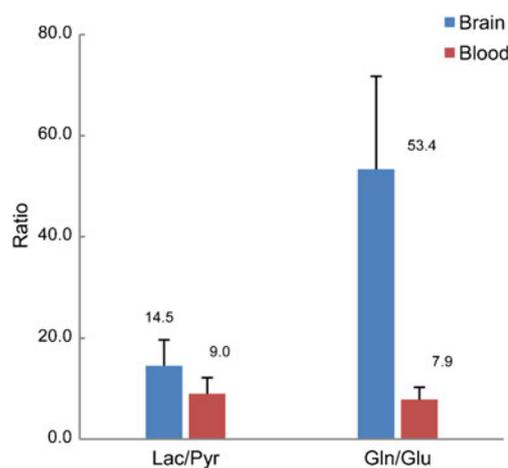


Fig. 3 Comparison of lactate/pyruvate (*Lac/Pyr*) and glutamine/glutamate (*Gln/Glu*) in primary visual cortex and the blood system of rhesus monkey. Ratios of *Lac/Pyr* and *Gln/Glu* in monkey primary visual cortex were significantly higher than the blood system ($p < 0.05$)

tized male rhesus monkeys. The microdialysis probe in the brain was placed into the primary visual cortex (V1). The concentrations of acetylcholine, lactate, pyruvate, glutamine, and glutamate in dialysates from primary visual cortex and blood are shown in Table 3. The values we monitored were in good agreement with the Human Metabolome Database Version 2.5 (www.hmdb.ca), and the values reported in the literature [18, 23, 32–39].

Comparison of lactate/pyruvate and glutamine/glutamate ratios in the brain and the blood

To further understand the distribution differences of NMs across the BBB, respectively, between the brain and the blood, we examined and compared the lactate/pyruvate and glutamine/glutamate ratios between the brain and blood dialysates. The results showed that the concentration ratio of lactate/pyruvate and glutamine/glutamate was significantly different between the brain and the blood. The values of lactate/pyruvate and glutamine/glutamate were calculated and are shown in Fig. 3. The mean lactate/pyruvate ratio in the brain was 14.5, but only reached 9.0 in the blood system. The mean ratio of glutamine/glutamate in the brain was 53.4, significantly higher than the blood value of 7.9. One-way ANOVA analysis confirmed the statistical significance of these findings ($p < 0.05$).

The lactate/pyruvate ratio in the human brain is currently considered a specific, sensitive marker that could potentially indicate whether glucose and oxygen supply meet the energy requirements of the brain tissue [40]. Such marker, that hint upon the adequacy of energy and metabolite supply are of great importance for the detection of onset of neurodegenerative diseases [41]. In the human brain, a lactate/pyruvate ratio above 40 indicates an “energy crisis” [42]. Additionally it was shown that lactate can increase the cerebral blood flow which correlates with an increase of the lactate/pyruvate ratio in the plasma [36]. A decreased interstitial glutamine/glutamate ratio has been observed in acute and chronic brain damage such as traumatic brain injury [43], hypoxia [44], and epilepsy [45]. Although such findings are still preliminary for diagnosis, we consider the simultaneous monitoring of blood and brain concentrations of important NMs a necessary step in this direction to get a clear understanding of their chemical relation across the BBB. On the basis of this data, extrapolations from the blood concentrations to brain concentrations of pathologically relevant NMs might be envisioned.

Conclusion

We developed a capillary HILIC-ESI/MS method for the simultaneous determination of multiple chemicals including

acetylcholine, lactate, pyruvate, glutamine and glutamate from the brain (EBF) and blood system of anesthetized non-human primates. To achieve highest intensity of these NMs, positive and negative ionization modes were used for MS detection. The optimized chromatographic separation allowed us to switch between the two detection modes and to simultaneously measure these NMs in a single run. A simple and fast sample treatment was carried out after collection by microdialysis without any additional sample purification or derivatization. We have successfully demonstrated that our method can reliably quantify these NMs in dialysates collected from the blood and the primary visual cortex of non-human primates. Additionally we found that the concentration ratio of lactate/pyruvate and glutamine/glutamate was significantly different between the brain and the blood, reflecting the active transports and different metabolic processes between the blood and the nervous system respectively across the BBB. Quantified coupling parameters between the blood and the nervous system of NMs are of utmost importance due to their possible use in the diagnosis of pathological processes in the brain. To further investigate the correlation of these NMs between the brain and the blood systems, we will use the developed analytical method to test the dynamic change of these NMs across the BBB by pharmacologically simulating dysfunctional states of the brain.

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