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## Supplementary Materials for **Deciphering Protein Kinase Specificity Through Large-Scale Analysis of Yeast Phosphorylation Site Motifs**

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Data set S3. MUSCLE alignment of all predicted *S. cerevisiae* ORFs with orthologs from 12 other yeast species (clustal alignment files).  
Data set S4. Alignment of yeast kinases analyzed in this study (clustal alignment file).

## Materials and Methods

### Yeast strains

pDEST-*PRK1KD* was generated from pDEST-*PRK1WT* (1) by mutating Asp176 to Ala by the QuikChange method using the following primers: PRK1KDUP: 5'-TGTATAAAGTTGTGCTTTGGTTCCGTTTC-3', PRK1KDDN: 5'-GAAACGGAACCAAAAGCACAAACTTATACA-3'. Sequence-confirmed pDEST-*PRK1KD* was then transformed into yeast strain Y258 using standard LiAc transformation methods (2).

*prk1Δ ark1Δ* strains were generated by first introducing a *prk1-Δ::KANMX6* deletion cassette by transformation using the available Bem2 and Ede1 chromosomally TAP-tagged strains (3). These *MATα prk1-Δ::KANMX6* strains were then mated to a *MATα ark1-Δ::URA3* strain that was obtained by rescuing haploids from the marker swapped *ark1-Δ::KANMX6* heterozygote diploid strain generated by the YKO consortium (pM4758) (4, 5). Following sporulation and tetrad dissection, *MATα prk1Δ::KANMX6 ark1Δ::URA3* haploids that still contained the TAP-tagged candidate substrate were identified by growth on synthetic complete media lacking uracil (SC-ura), rich media containing geneticin (YPAD + G418), and synthetic complete media lacking histidine (SC-his) and positive mating to the a mating type tester strain.

*vhs1Δ::KANMX6 Sol2-TAP::HIS3MX6* was generated by transformation using the available Sol2 chromosomally TAP-tagged strain.

44 of the kinases were purified from yeast using either the N-terminal GST-His<sub>6</sub>-tagged overexpression ORF collection (6) or the C-terminal TAP-tagged overexpression ORF collection (1). Yeast strains were first grown as starter cultures in 4 ml of SC-ura media, and then diluted into 400 ml of SC-ura/raffinose media and allowed to grow ~16 hr at 30°C to an OD<sub>600</sub> of 0.6-0.8. Fusion protein expression was induced by adding 200 ml of 3X YEP-Gal (3% yeast extract, 6% peptone, 6% galactose) for 6 hr. Cells were harvested by centrifugation, washed with ice-cold water, aliquoted into four tubes, and stored at -80°C.

### Preparation of crude yeast extracts

To prepare crude extracts for affinity purification, cell pellets were thawed on ice, and 500 µl of 0.5 mm acid washed glass beads (Biospec) was added to each tube. Two rounds of lysis were performed: once with lysis buffer (50 mM Tris-HCl, pH 7.5, 1 mM EGTA, 0.1% Triton X-100, 10% glycerol, 0.5 mM DTT, 1 mM PMSF, 1 mM NaF, 12.5 mM β-glycerophosphate, 1X Complete protease inhibitors [Roche]) containing 150 mM NaCl (LB150) and once with lysis buffer containing 650 mM NaCl (LB650). 500 µl of LB150 was added to each tube, and cells were lysed by shaking for 6 min in a paint-shaker (5G-HD, Harvil) at 4°C followed by centrifugation at 20,000 X g for 5 min. Supernatants were combined together in a single 15 ml conical tube. 500 µl of LB 650 was then added to each tube containing the remaining cell debris. Cells were subjected to another round of paint-shaking, and second round lysates were collected as described above and combined with the first round lysates.

### Affinity purification of GST-His<sub>6</sub>-tagged kinases

To affinity purify GST-His<sub>6</sub>-tagged kinases from yeast, crude extracts were mixed with 500 µl of washed glutathione sepharose (GE Healthcare) resuspended in 4 ml of lysis buffer (without added NaCl) and incubated with agitation for 2 hr at 4°C. Beads were pelleted by centrifugation at 2,500 X g for 5 min and washed three times with 10 ml of cold wash buffer (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.1% Triton X-100, 10% glycerol). Beads were washed an additional two times with kinase buffer without MgCl<sub>2</sub> (20 mM HEPES, pH 7.4, 150 mM NaCl, 0.1% Tween 20, 25% glycerol, 1 mM DTT), and bound kinase was recovered by two rounds of elution with 200 µl of kinase buffer containing 30 mM reduced GSH (15 min, 4°C with agitation). Eluate was separated from beads by centrifugation through a 3 µm polycarbonate filter plate (5 min at 1,900 X g). The eluted kinase was snap frozen and stored at -80°C.

#### *Affinity purification of TAP-tagged kinases*

To affinity purify TAP-tagged kinases from yeast, crude extracts were mixed with 500 µl of washed IgG sepharose (GE Healthcare) resuspended in 4 ml of lysis buffer (without added NaCl) and incubated with agitation for 2 hr at 4°C. Beads were pelleted by centrifugation at 210 X g for 5 min and washed three times with 10 ml of cold wash buffer (above). Beads were washed an additional two times with kinase buffer without MgCl<sub>2</sub> (above). To elute bound kinase, beads resuspended in 400 µl of kinase buffer were incubated with 15 µl of GST-3C protease (expressed and purified as described (1)) on a nutator overnight at 4°C. Cleaved kinase was separated from beads by centrifugation through a 3 µm polycarbonate filter plate as described above. GST-3C protease was removed by mixing the flow-through with 250 µl of washed glutathione sepharose (1 hr, 4°C). Pure kinase was recovered by a second round of centrifugation through fresh wells of a filter plate. The purified kinase was snap frozen and stored at -80°C.

#### *Expression and purification of kinases from mammalian cells*

Coding sequences for Cak1, Cdc15 (residues 1 – 303), Gcn2 (residues 590 – 994), Prr1, Skm1 (residues 339 – 655), Vhs1, and Ykl171w (residues 436 – 913) were PCR amplified from yeast genomic DNA and ligated into the mammalian expression vector pEBG2, which produces proteins as N-terminal GST fusions. Kinases were expressed by transient transfection of HEK293T cells using Lipofectamine Plus (Invitrogen). Between 40 and 44 hr after the start of transfection, cells were washed with ice-cold PBS, and extracted into 293 lysis buffer (20 mM Tris, pH 7.5, 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1% Triton X-100, 1 mM DTT, 1 mM PMSF, 2.5 mM sodium pyrophosphate, 1 mM β-glycerophosphate, 1 mM Na<sub>3</sub>VO<sub>4</sub>, 10 µg/ml leupeptin, 2 µg/ml pepstatin A, 10 µg/ml aprotinin), 250 µl per 6 cm plate. Lysates were cleared by centrifugation (10 min at 16,000 X g, 4°C), and incubated with glutathione sepharose (8 µl per plate) for 2 hr at 4°C. Beads were pelleted and washed twice with 293 lysis buffer, twice with GSH wash buffer (50 mM HEPES, pH 7.4, 5 mM β-glycerophosphate, 2 mM DTT, 0.1 mM Na<sub>3</sub>VO<sub>4</sub>, 10 mM MgCl<sub>2</sub>), and then eluted with two rounds GSH wash buffer containing 20 mM reduced GSH and 10% glycerol (10 µl per plate each round for 30 min on ice). Eluted kinases were snap frozen in aliquots and stored at -80°C.

### *Production of remaining kinases*

The following kinases were prepared in active form as described: Cdc28-Cln1 (7), Cdc7 (8), Fus3 (9), Hog1 (10), Ipl1-Sli15 (11), Mek1 (12), Pho85-Pcl1/Pcl2/Pho80 (13), Rad53 (14), and Rim15 (15).

Full length *Kss1* coding sequence was cloned by PCR into the bacterial His<sub>6</sub> tag expression vector pET28. To produce active *Kss1*, BL21 *E. coli* were co-transformed with pET28-*Kss1* and pGEX4T-Ste7EE, which produces a constitutively active mutant of Ste7 that has its two activation loop phosphorylation sites mutated to Glu. Phosphorylated *Kss1* was expressed and purified as described for mammalian MAPKs (16).

*Rck2* purified from yeast was activated by incubation with Hog1 (15 µg/ml) in 20 mM HEPES, pH 7.4, 10 mM MgCl<sub>2</sub>, 1 mM DTT, 1 mM ATP for 30 min at 30°C immediately prior to peptide screening. Residual Hog1 activity was blocked during assay of *Rck2* by including 50 µM SB203580 in the reactions.

### *Characterization of yeast kinase preparations*

Purified kinases were analyzed by SDS-PAGE followed by Coomassie staining and immunoblotting against the GST or HA tag (anti-GST antibody, 1:1,000, Sigma or anti-HA16B12 antibody, 1:1,000, Covance) as appropriate. Kinase preparations deemed to be sufficiently pure (>90%) and of the correct fusion size were tested for kinase activity by radiolabel assay using a cocktail of myelin basic protein (MBP, Upstate) and histone H1 (Upstate) as substrates. Purified kinase (7 µl) was incubated for 1 hr at 30°C with 0.8 µg each of MBP and histone H1 in the presence of 100 µM ATP, 0.25 µCi γ-[<sup>33</sup>P]-ATP (Amersham), and 10 mM MgCl<sub>2</sub> in a total reaction volume of 10 µl. The reactions were stopped by adding 3X SDS-PAGE sample loading buffer (5 µl) followed by heating for 4 min at 95°C. Proteins were resolved by SDS-PAGE, and gels were dried and exposed to autoradiography film. Activity of each kinase preparation was assessed by analyzing the extent of both autophosphorylation and phosphorylation of the exogenous substrates as indicated by the amount of incorporated radiolabel.

### *Assay of individual peptide substrates*

Optimized peptide substrates and individual variants were synthesized using standard Fmoc chemistry and purified by reversed phase HPLC. Kinase assays were performed at 10 µM peptide concentration in kinase buffer with 50 µM γ-[<sup>33</sup>P]-ATP (1 µCi per 25 µl reaction) (10 - 15 min, 30°C). At 5 min time points, reactions were spotted onto P81 phosphocellulose membrane, which was washed extensively with 0.42% phosphoric acid, immersed briefly in acetone, and air-dried. Peptide phosphorylation signals were quantified by scintillation counting.

### *In vitro kinase assays of protein substrates*

*Prk1* WT kinase, *Prk1* KD kinase, and candidate substrates were purified from the C-terminal TAP-tagged overexpression ORF collection (1) on IgG sepharose beads as described above. Purified kinase (2.5 µl) was mixed with each purified candidate substrate (5 µl) and incubated for 1 hr at 30°C in the presence of 100 µM ATP, 0.25 µCi

$\gamma$ -[<sup>33</sup>P]-ATP (Amersham), and 10 mM MgCl<sub>2</sub>. The total volume of each reaction was 10  $\mu$ l. Reactions were stopped by adding 3X SDS-PAGE sample loading buffer (5  $\mu$ l) followed by heating for 4 min at 95°C. Proteins were fractionated by SDS-PAGE, and gels were dried and exposed to autoradiography film.

#### *Electrophoretic mobility shift analyses*

$1.8 \times 10^7$  cells grown in YPAD to OD<sub>600</sub> of 0.6 were harvested by centrifugation and washed with ice-cold water. TAP-tagged Prk1/Ark1 substrates were purified on IgG sepharose from yeast cell lysates as described above, except that proteins were not eluted from the beads. Beads were washed twice with  $\lambda$  protein phosphatase ( $\lambda$ -PPase) reaction buffer [New England Biolabs (NEB)], and resuspended in the same buffer (30  $\mu$ l). Samples were phosphatase treated where indicated by incubating each sample with 1.5 units each of calf intestinal phosphatase (NEB), protein phosphatase 1 (NEB), and  $\lambda$ -PPase (NEB) for 30 min at 30°C. Substrates were eluted from the beads in 1X SDS loading buffer by heating for 4 min at 95°C and resolved by SDS-PAGE, followed by immunoblotting with anti-TAP antibody (Open Biosystems, 1:3000).

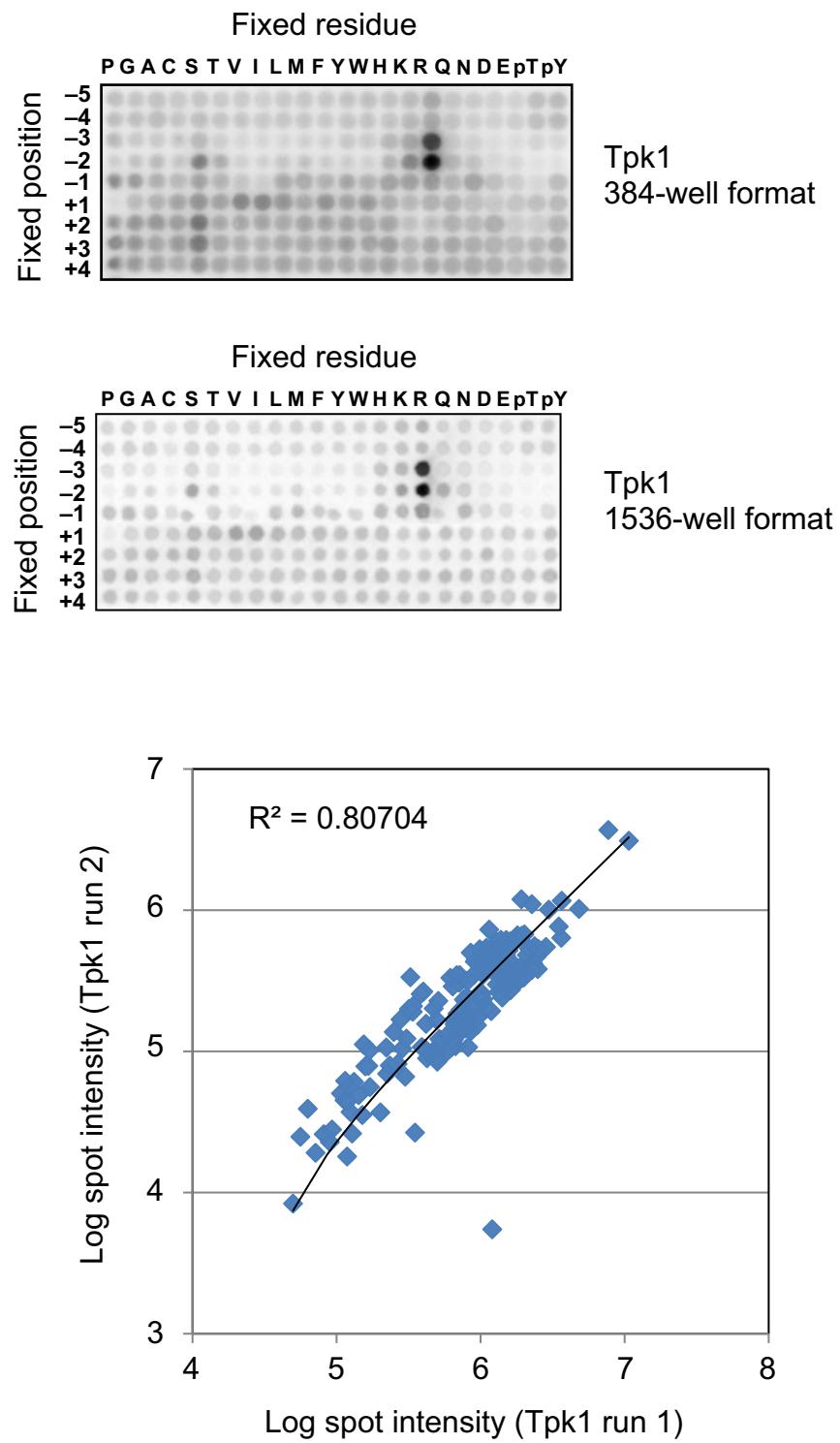
Electrophoretic mobility of Sol2 was determined from crude lysates. Cell pellets from WT Sol2-TAP and *vhs1Δ* Sol2-TAP strains were lysed directly in 250  $\mu$ l 1X SDS loading buffer (NEB) using the FastPrep®-24 System (6 m/s for 20 sec, MP Biomedicals) with 250  $\mu$ l of 0.5 mm acid washed glass beads (Biospec). The resulting crude extracts were heated for 4 min at 95°C and resolved on SDS-PAGE gels containing 25 mM Phos-tag (17) (NARD Institute, Ltd.) followed by immunoblotting with anti-TAP antibody as described above.

#### **Supplemental Datasets**

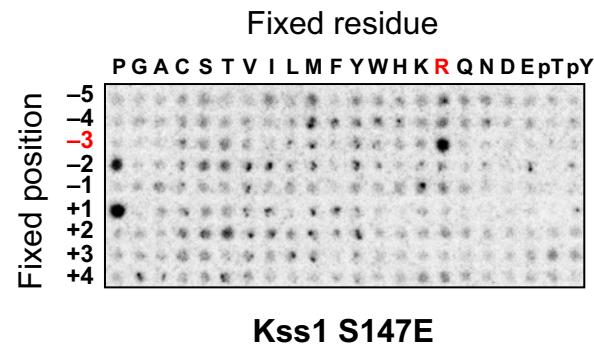
Dataset S1. Tab delimited files containing average PWMs for each of the 61 kinases assayed. PWM format follows spot array design where each row represents which position relative to the targeted phosphoacceptor site is fixed and each column represents which amino acid is fixed at that position. Spots intensities from the peptide arrays were quantified and normalized so that the total value within a single row (corresponding to a single position relative to the phosphorylation site), not including pThr and pTyr, was given a value of 20. Values greater than 1.0 thus represent positively selected residues, and values less than 1.0 are negatively selected.

Dataset S2. Tab delimited files containing MOTIPS output for each of the 61 kinases assayed. Phosphorylation sites in yeast proteins were ranked by match to the PWM and scored for predicted accessibility, probability that it falls in a disordered region, and sequence conservation across 13 yeast proteomes. An overall likelihood score is given that is the output of the Bayesian classifier of these four features. Separate columns in each file indicate whether the kinase and predicted substrate share a common GO cellular compartment, whether the substrate was identified through proteome chip screening by Ptacek, *et al.* (18), and whether a physical or genetic interaction has been reported (according to Biogrid). If the site appeared in a MS-based phosphoproteomic screen, the identified phosphopeptide is given.

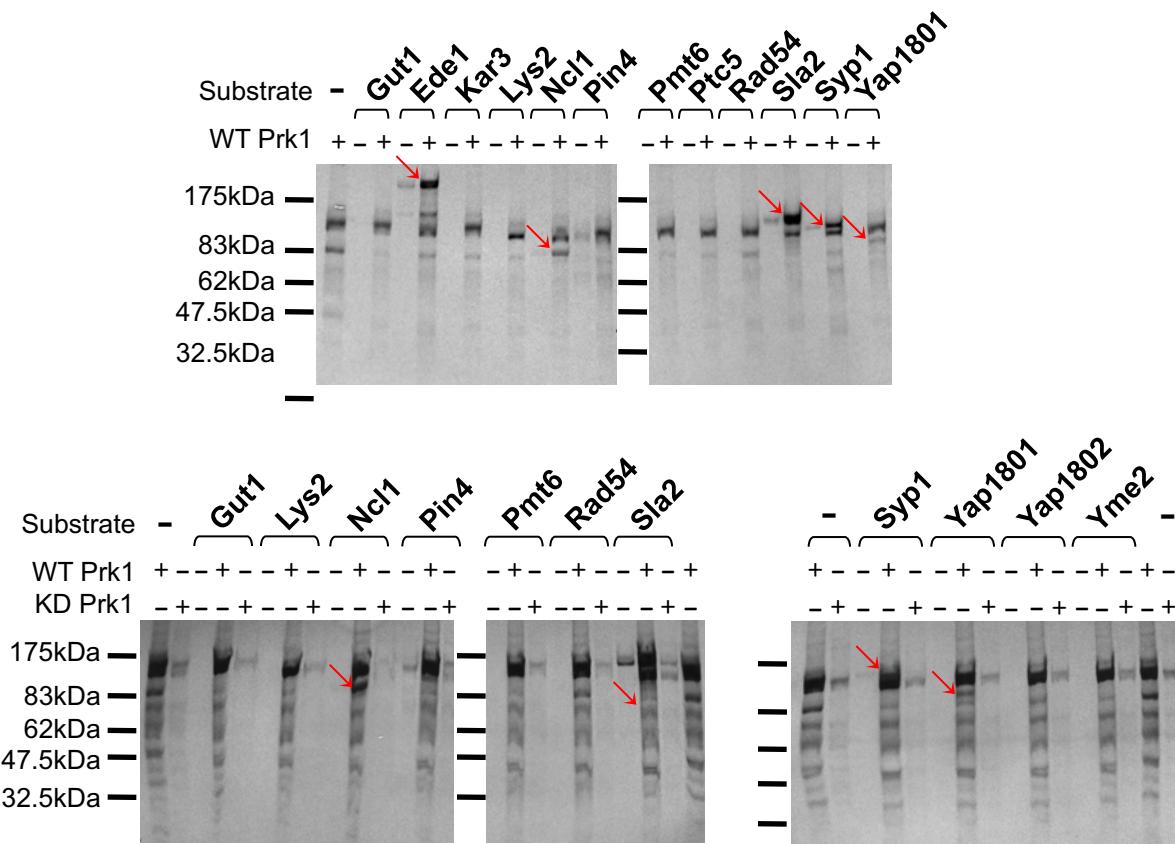
Dataset S3. Clustal files containing multiple sequence alignments of every *S. cerevisiae* protein sequence with its orthologs from 12 other yeasts.



**Figure S1.** Assay reproducibility. Tpk1 was assayed using both the 384-well plate format originally reported and the miniaturized 1536-well format. The graph at bottom shows the correlation between spot intensities corresponding to each peptide for two runs with Tpk1 using the miniaturized 1536-well format.

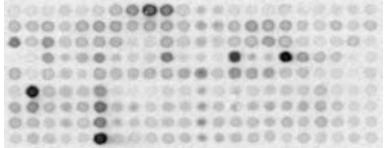
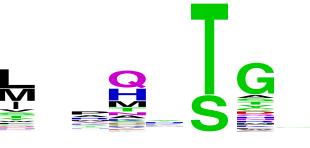
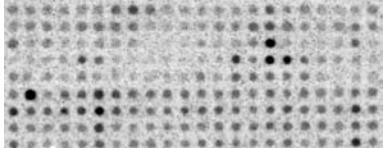
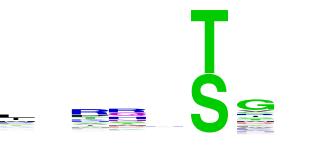
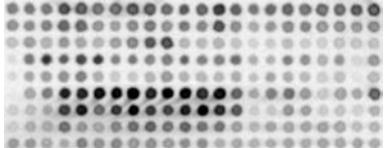
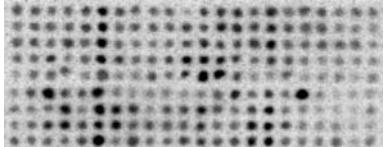
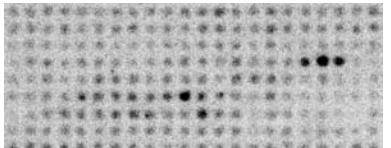
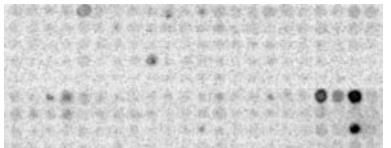


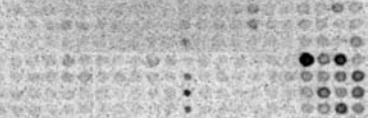
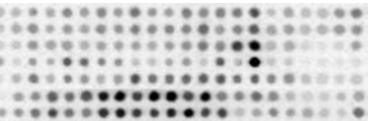
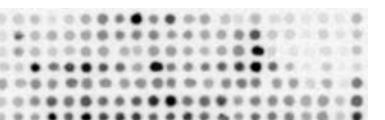
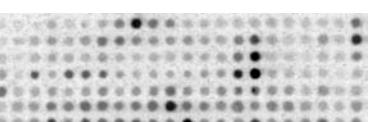
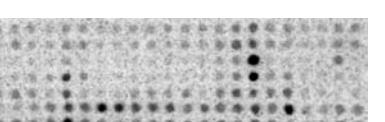
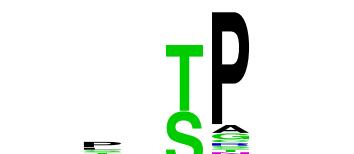
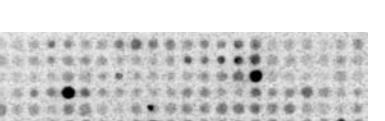
**Figure S2.** Representative peptide array screening results for the Kiss1 S147E mutant.

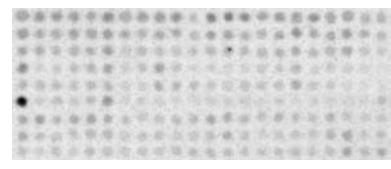
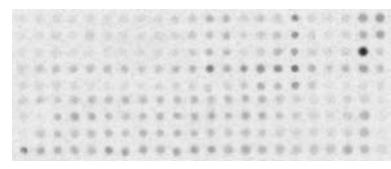
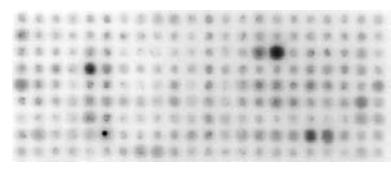
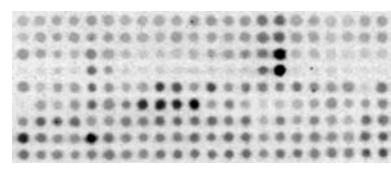
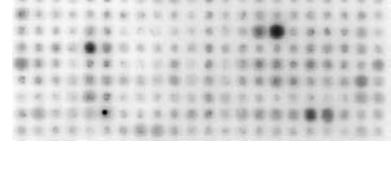
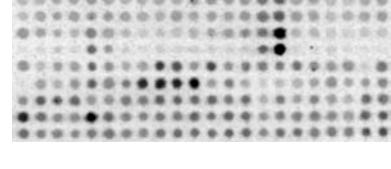
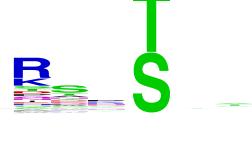
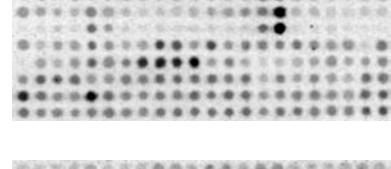
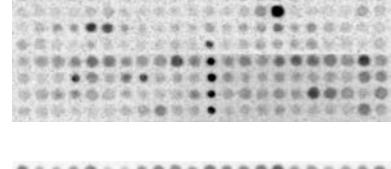
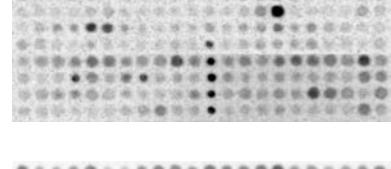
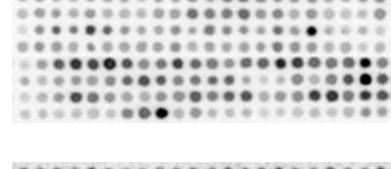
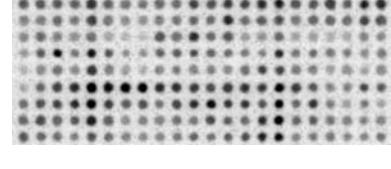
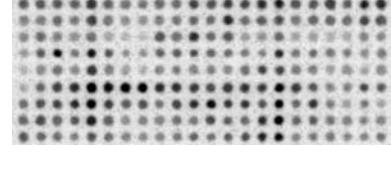
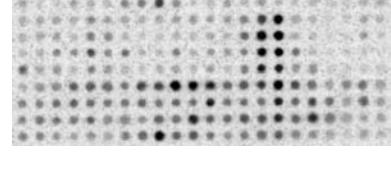
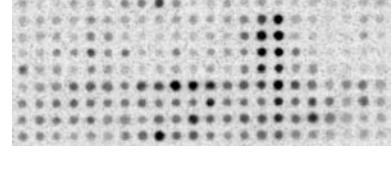


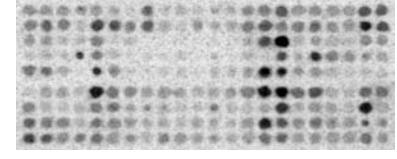
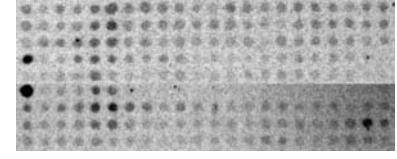
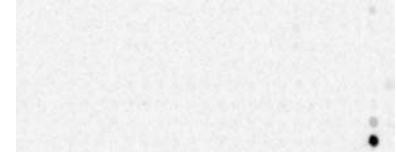
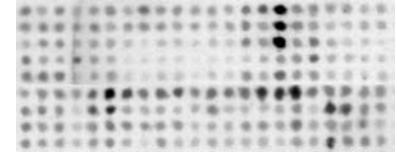
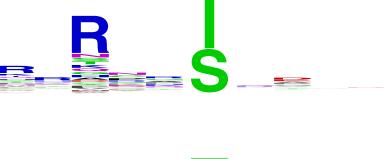
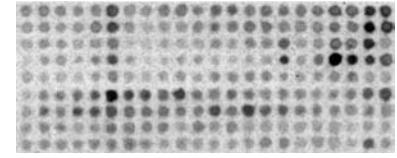
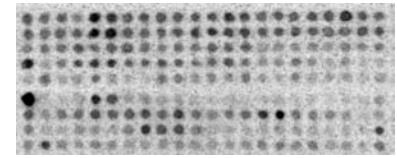
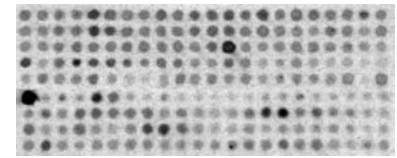
**Figure S3.** Representative Prk1 in vitro assays. Candidate substrates were purified and assayed in the presence of either wildtype Prk1 (WT Prk1) or a Prk1 kinase inactive mutant (KD Prk1, D176A), or in the absence of any kinase in 10  $\mu$ l reactions. Red arrows point to validated substrates.

**Table S1.** Representative peptide array screening results and sequence logos for each of the 61 kinases assayed. Hsl1 was assayed in the original 384-well format which lacked the zero position control mixtures, and thus no information regarding its selectivity for Ser or Thr residues as the targeted phosphoacceptor site was obtained.

Gene Name	ORF Name	Array Image PGACSTVILMFYWHKRQNDEpTpY -5 -4 -3 -2 -1 +1 +2 +3 +4	0:- S T	Logo
<i>AKL1</i>	<i>YBR059C</i>			
<i>ARK1</i>	<i>YNL020C</i>			
<i>ATG1</i>	<i>YGL180W</i>			
<i>CAK1</i>	<i>YFL029C</i>			
<i>CDC15</i>	<i>YAR019C</i>			
<i>CDC28</i>	<i>YBR160W</i>			
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<i>CDC7</i>	<i>YDL017W</i>			

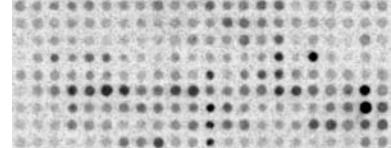
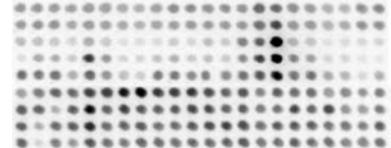
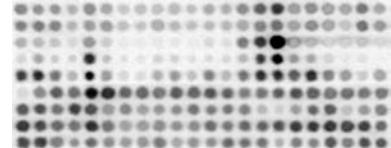
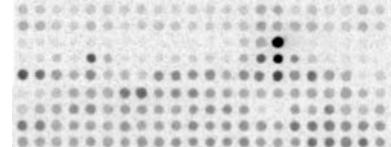
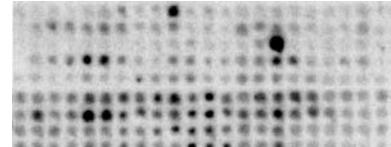
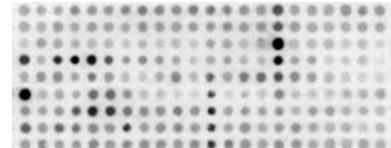
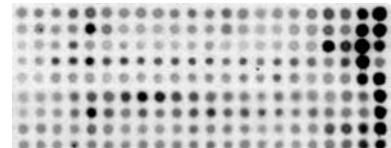
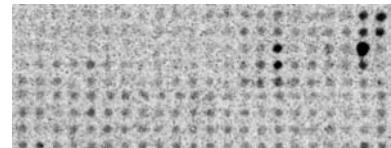
Gene Name	ORF Name	Array Image		Logo
		PGACSTVILMFYWHKRQNDEpTpY	0:- S T	
CKA1	YIL035C	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
CLA4	YNL298W	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
CMK1	YFR014C	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
CMK2	YOL016C	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
FMP48	YGR052W	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
FUS3	YBL016W	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
GCN2	YDR283C	 -5 -4 -3 -2 -1 +1 +2 +3 +4		
GIN4	YDR507C	 -5 -4 -3 -2 -1 +1 +2 +3 +4		

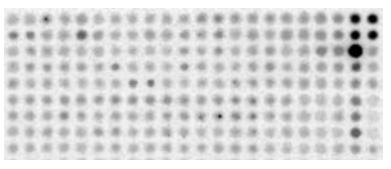
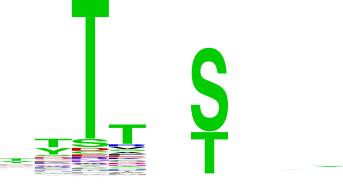
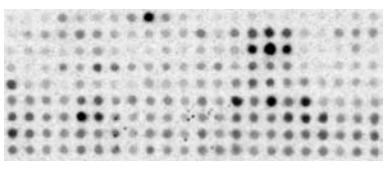
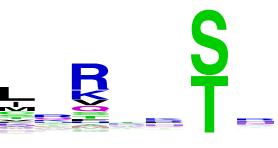
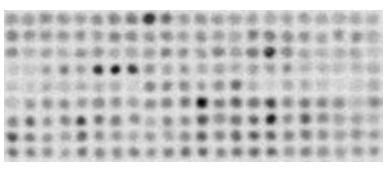
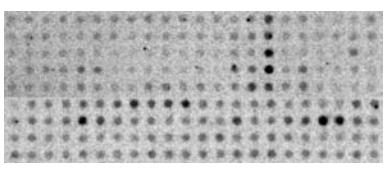
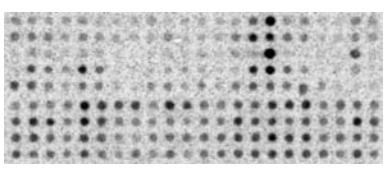
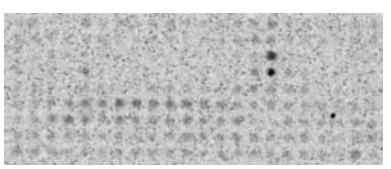
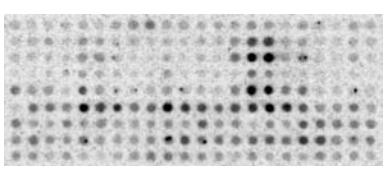
Gene Name	ORF Name	Array Image PGACSTVILMFYWHKRQNDEpTpY 0:- S T	Logo
<i>HOG1</i>	<i>YLR113W</i>	 	
<i>HRR25</i>	<i>YPL204W</i>	 	
<i>HSL1</i>	<i>YKL101W</i>	 	
<i>IPL1</i>	<i>YPL209C</i>	 	
<i>KCC4</i>	<i>YCL024W</i>	 	
<i>KIN1</i>	<i>YDR122W</i>	 	
<i>KIN3</i>	<i>YAR018C</i>	 	
<i>KIN4</i>	<i>YOR233W</i>	 	

Gene Name	ORF Name	Array Image PGACSTVILMFYWHKRQNDEpTpY	0:- S T	Logo
<i>KSP1</i>	<i>YHR082C</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>KSS1</i>	<i>YGR040W</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>MCK1</i>	<i>YNL307C</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>MEK1</i>	<i>YOR351C</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>MPS1</i>	<i>YDL028C</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>MRK1</i>	<i>YDL079C</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>PHO85-PCL1</i>	<i>YPL031C-YNL289W</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		
<i>PHO85-PCL2</i>	<i>YPL031C-YDL127W</i>	 <p>-5 -4 -3 -2 -1 +1 +2 +3 +4</p>		

Gene Name	ORF Name	Array Image			Logo
		PGAC	STVILM	FYWHKRQNDEpTpY	
<i>PHO85-PHO80</i>	<i>YPL031C-YOL001W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>PKH2</i>	<i>YOL100W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>PRK1</i>	<i>YIL095W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>PRRI</i>	<i>YKL116C</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>PSK2</i>	<i>YOL045W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>PTK2</i>	<i>YJR059W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>RAD53</i>	<i>YPL153C</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>RCK2</i>	<i>YLR248W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			

Gene Name	ORF Name	Array Image			Logo
		PGAC	S T V I L M F Y W H K R Q N D E p T p Y	0:- S T	
<i>RIM11</i>	<i>YMR139W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>RIM15</i>	<i>YFL033C</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>SCH9</i>	<i>YHR205W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>SKM1</i>	<i>YOL113W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>SKY1</i>	<i>YMR216C</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>SLT2</i>	<i>YHR030C</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>SNF1</i>	<i>YDR477W</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			
<i>STE20</i>	<i>YHL007C</i>	-5 -4 -3 -2 -1 +1 +2 +3 +4			

Gene Name	ORF Name	Array Image PGACSTVILMFYWHKRQNDEpTpY -5 -4 -3 -2 -1 +1 +2 +3 +4	0:- S T	Logo
<i>TOS3</i>	<i>YGL179C</i>			
<i>TPK1</i>	<i>YJL164C</i>			
<i>TPK2</i>	<i>YPL203W</i>			
<i>TPK3</i>	<i>YKL166C</i>			
<i>VHS1</i>	<i>YDR247W</i>			
<i>YAK1</i>	<i>YJL141C</i>			
<i>YCK1</i>	<i>YHR135C</i>			
<i>YCK2</i>	<i>YNL154C</i>			

Gene Name	ORF Name	Array Image PGACSTVILMFYWHKRQNDEpTpY -5 -4 -3 -2 -1 +1 +2 +3 +4	0:- S T	Logo
<i>YCK3</i>	<i>YER123W</i>			
<i>YDL025C</i>	<i>YDL025C</i>			
<i>YKL171W</i>	<i>YKL171W</i>			
<i>YNR047W</i>	<i>YNR047W</i>			
<i>YPK1</i>	<i>YKL126W</i>			
<i>YPK2</i>	<i>YMR104C</i>			
<i>YPL141C</i>	<i>YPL141C</i>			

**Table S2.** Protein kinases analyzed in this study. Mammalian orthologs are indicated where known. References for reported phosphorylation motifs are listed where either the yeast kinase itself or the mammalian ortholog has been previously characterized.

Kinase	ORF	Category	Ortholog	Reported motif
Akl1	YBR059C	Other	BiKE	None
Ark1	YNL020C	Other	BiKE	None
Atg1	YGL180W	Other	ULK2	None
Cak1	YFL029C	Other	None	None
Cdc15	YAR019C	Other	None	(19)
Cdc28	YBR160W	Proline	CDK2	(20)
Cdc5	YMR001C	Acidophilic	PLK1	(21)
Cdc7	YDL017W	Acidophilic	CDC7	None
Cka1	YIL035C	Acidophilic	CK2	(22)
Cla4	YNL298W	Basophilic	PAK1	(23)
Cmk1	YFR014C	Basophilic	CAMK1	(24)
Cmk2	YOL016C	Basophilic	CAMK1	(24)
Fmp48	YGR052W	Basophilic	None	None
Fus3	YBL016W	Proline	ERK	(22)
Gcn2	YDR283C	Acidophilic	GCN2	None
Gin4	YDR507C	Basophilic	BRSK1	None
Hog1	YLR113W	Proline	p38- $\alpha$	(25)
Hrr25	YPL204W	Acidophilic	CK1- $\delta$	(22)
Hsl1	YKL101W	Basophilic	BRSK1	None
Ipl1	YPL209C	Basophilic	Aurora kinase C	(26)
Kcc4	YCL024W	Basophilic	BRSK1	None
Kin1	YDR122W	Other	MARK2/Par1b	None
Kin3	YAR018C	Other	NEK2	(27)
Kin4	YOR233W	Basophilic	None	None
Ksp1	YHR082C	Basophilic	None	None
Kss1	YGR040W	Proline	ERK	(22)
Mck1	YNL307C	Acidophilic	GSK3	(28)
Mek1	YOR351C	Basophilic	None	None
Mps1	YDL028C	Acidophilic	TTK/MPS1	None
Mrk1	YDL079C	Acidophilic	GSK3	(28)
Pho85	YPL031C	Proline	None	None
Pkh2	YOL100W	Basophilic	PDK1	None
Prk1	YIL095W	Other	AAK1	None
Prr1	YKL116C	Basophilic	None	None
Psk2	YOL045W	Basophilic	PASK	None
Ptk2	YJR059W	Basophilic	None	None
Rad53	YPL153C	Basophilic	None	None
Rck2	YLR248W	Basophilic	MK2	(25)

Table S2, continued

Rim11	YMR139W	Acidophilic	GSK3	(28)
Rim15	YFL033C	Other	None	None
Sch9	YHR205W	Basophilic	Akt/PKB	(29)
Skm1	YOL113W	Basophilic	PAK1	(23)
Sky1	YMR216C	Basophilic	SRPK2	None
Slt2	YHR030C	Proline	None	None
Snf1	YDR477W	Basophilic	AMPK	(30)
Ste20	YHL007C	Basophilic	PAK1	(23)
Tos3	YGL179C	Other	LKB1	(31)*
Tpk1	YJL164C	Basophilic	PKA	(20)
Tpk2	YPL203W	Basophilic	PKA	(20)
Tpk3	YKL166C	Basophilic	PKA	(20)
Vhs1	YDR247W	Basophilic	None	None
Yak1	YJL141C	Proline	DYRK4	(32)
Yck1	YHR135C	Acidophilic	CK1- $\gamma$ 2	(22)
Yck2	YNL154C	Acidophilic	CK1- $\gamma$ 2	(22)
Yck3	YER123W	Acidophilic	CK1- $\gamma$ 3	(22)
Ydl025c	YDL025C	Basophilic	None	None
Ykl171w	YKL171W	Basophilic	None	None
Ynr047w	YNR047W	Basophilic	None	None
Ypk1	YKL126W	Basophilic	SGK2	None
Ypk2	YMR104C	Basophilic	SGK2	None
Ypl141c	YPL141C	Basophilic	None	None

\*Our observed motif for Tos3 differs from that reported for its human ortholog LKB1.

**Table S3.** Pairwise correlation coefficients for each of four genomic features and the Scansite match score. The PRK1 PWM was run on MOTIPS to obtain the top 2,000 predicted binding sites with values for all the five features: hits per protein, match score, disorder score, accessibility score, and conservation score. Multiple hits on a single open reading frame (ORF) were combined and the best set of scores was chosen, resulting in a list of 1,583 hits with no redundant protein targets. Pairwise Spearman correlation was then performed for all the features using the statistical software R.

	Hits per ORF	Scansite score	Accessibility score	Disorder score	Conservation score
Hits per ORF	1.00	-0.25	0.06	0.17	-0.02
Scansite score	-0.25	1.00	-0.01	-0.03	-0.01
Accessibility score	0.06	-0.01	1.00	0.57	-0.42
Disorder score	0.17	-0.03	0.57	1.00	-0.27
Conservation score	-0.02	-0.01	-0.42	-0.27	1.00

**Table S4.** Alignment of yeast kinases analyzed in this study. Dataset S4 contains this alignment in clustal format.

	1	10	20	30	40	50
TOS3_Scer/1-295	FEILATLGNGQYGKVKLARDLGT				GALVAIKILNR	-----
TPK3_Scer/1-255	FQILRTLGTGSFGRVHLIRSNHN				GRFYALKTLKKHTIVK	
TPK1_Scer/1-255	FQILRTLGTGSFGRVHLIRSRHN				GRYYAMKVLKKEIVVR	
TPK2_Scer/1-255	FQIMRTLGTGSFGRVHLVRSVHN				GRYYAIKVLKKQQVVK	
YPK1_Scer/1-256	FDLLKVIGKGSGFKVMQVRKKDT				QKVYALKAIRKSYIVS	
YPK2_Scer/1-256	FDLLKVIGKGSGFKVMQVRKKDT				QKIYALKALRKAYIVS	
SCH9_Scer/1-260	FEVLRLLGKGTFGQVYQVKKKD				QRIYAMKVLSSKKVIVK	
YNR047W_Scer/1-282	FEKIRLLGQGDVGKVFLVREKKT				NRVYALKVLSKDEM	IK
RIM15_Scer/1-269	YDILKPISKGAYGSVYLARKKLT				GDYFAIKVLRKSDMIA	
IPL1_Scer/1-252	FELGKKLGKGKFGKVYCVRHRST				GYICALKVMEKEEIK	
PKH2_Scer/1-265	FKFGSVIDGDAYSTVMLATSIDT				KKRYAAKVLNKEYLIR	
KCC4_Scer/1-265	WKLGETLGFGSTGKVQLAQHERT				GHRTAVKVISKSIFNN	
GIN4_Scer/1-271	WKLGETLGLGSTGKVQALARNGST				GQEAAVKVISKA	VFT
HSL1_Scer/1-289	WKLGKTLGKGSSGRVRLAKNMET				GQAAIKIVPKKKAFV	
SNF1_Scer/1-252	YQIVKTLGECSFGKVLAYHTT				GQKVALKIINKVLA	K
KIN1_Scer/1-279	WEFVETVGAGSMGKVKLAKHRYT				NEVCAVKIVNRATKAF	
KIN4_Scer/1-268	YIIGSTLGEGEFGKVKGWPKNFSNNS				VPKQVAIKLIRRDTIKK	
YPL141C_Scer/1-273	YILGSTLGEGEFGKVKGWPKNFSNNS				TFDFPKQVAIKLIKRD	SISN
CMK1_Scer/1-263	YVFGKTLGAGTFGVVRQAKNTET				GEDVAVKILIKKALKG	
CMK2_Scer/1-263	YIFGRTLGA	GGSFGVVQRQAKLST			NEDVAIKILLKKALOG	
RAD53_Scer/1-269	SIIDEVVGQGAFATVKKAIERTT				KTFAVVIISKRKVIG	
RCK2_Scer/1-316	YKLINKIGEGAFSKVFR	FAIPAKNNSNEFL			TKNYKAVA	VIKKADLSS
APG1_Scer/1-302	YTAEKEIGKGSFATVYRGHLTS	DK			SQHVAIKEVSRA	LKN
MEK1_Scer/1-284	EITNRIVGN	GNTFGHVLITHNSKERDE	DVCYH		PENYAVKIIKLK	---
YKL171W_Scer/1-283	HRLGK	IIGFGAWGIIRECFDI	ET		GVGRV	IKIVKF
CDC5_Scer/1-256	YHRGHFLGEGGFARCFQ	-IKDD	S		GEIFA	AKTVAKASIKS
PSK2_Scer/1-259	FTILQVMGE	GEGAYGKVNLCIHN			REHYIVVI	KMIFKERILV
YDL025C_Scer/1-274	GIPGRKLGE	GEGASGSVSVVER			TDGKLFACK	MFRKPHLN
SKY1_Scer/1-312	YILVRKLGW	GWFSTVWLAKDMVNN			THVAMKIV	RGDKVY
YAK1_Scer/1-257	YLVL	DILGQGTFGQVVKCQNL	LT		EILAVKVV	KSRT
PTK2_Scer/1-305	DTDNKPIG	SGGSSEVRKVKS			YRQKD	VYALKKLN
PHO85_Scer/1-291	FKQLEKLN	GNTYATVYKGLNKTT			--MIYH	
CDC28_Scer/1-288	YKRLEKV	GEETYGVVYKALDLRPGQG			G	VYVALKEVKLD
SLT2_Scer/1-296	FQLIKEIGHG	AYGIVCSARFAAEE			V	RVVALKKIRLES
KSS1_Scer/1-301	YKLVDL	IGEGAYGTCSAHIKPS			DT	TVAIKKVTNVFSKT
FUS3_Scer/1-297	FQLKSLL	GEGAYGVVCSATHKPT			GI	KAIIQ-PFSKK
HOG1_Scer/1-280	YNDLN	PVGMAFGLVCSATDTLT			VA	IIKKIE-PFDKP
RIM11_Scer/1-284	FPTTE	EVVGHGSFGVV	FATVIQET		SQ	PVAIKKIMKF
MRK1_Scer/1-284	YPTTE	EVVGHGSFGVV	VTTVIET		P	A
MCK1_Scer/1-293	VKEYRK	IGRGA	FGTVVQAYLTQDKKNWL		EK	VAIKKVLQDKRF
CKA1_Scer/1-324	YEIEN	KVGRGKYSE	VQGVKLD		NQK	VAIKKVLQDR
CDC7_Scer/1-272	YKLID	KIGEGTFSSVY	KAKDITGKITKKFASHFW		V	YK
YCK1_Scer/1-261	YKIGK	KIGEGSGF	GVLFEGTNMIN		GP	FAIKKVA
YCK2_Scer/1-261	YKIGK	KIGEGSGF	GVLFEGTNMIN		AI	KKVPA
YCK3_Scer/1-296	YAVGP	KIGEGSGF	VIFEGENILHSCQ		KE	Y
HRR25_Scer/1-259	FRIGRK	I	GSGFDIYHGTLNIS		SR	YYAIKKIR
PRR1_Scer/1-317	WKKVRP	IGSGNF	STVLLYELMDQSN		Y	TEK-
MPS1_Scer/1-277	YEKI	ELLGRGGSSRVY	KVKGSGN		Y	YV
GCN2_Scer/1-268	FEEIAVL	QGQAFQVV	KARNALD		Y	YAIKKR
KIN3_Scer/1-319	YQVLEE	IGRGSFG	SVRKVHIPT		Y	YGHMNS
PRK1_Scer/1-277	AKII	IKYLTSGG	FAQVYTA		Y	Y
ARK1_Scer/1-277	VEII	IKYLTSGG	FAQVYSA		Y	Y
AKL1_Scer/1-285	VEVV	NYLAEGG	FAQIYVV	KFL	Y	Y
VHS1_Scer/1-321	YLITSQ	IGEGAY	GLVYRALDIRTD		Y	Y
SKM1_Scer/1-280	FOLVEK	AGQGAS	GAVYLSKRIKLP		Y	Y
CLA4_Scer/1-280	FKVIEK	AGQGAS	GSVYLAERTH	IP	Y	Y
STE20_Scer/1-252	YANLV	KIGQGAS	GGVYTAYEIGT		Y	Y
CDC15_Scer/1-248	YHLKQV	IGRGSY	GVVYKAINKHT		Y	Y
FMP48_Scer/1-285	YTKLRSI	QSGTF	STVYKAWSTTHN		Y	Y
KSP1_Scer/1-260	YQKIEDI	SEGSY	GYVSLAKDVREK		Y	Y
CAK1_Scer/1-304	-MKLDS	IDI	THCQLVKSTR	TARIYR	-SD	TYAIKCLALDF

Table S4, continued

	61	70	80	90	100	110
TOS3_Scer/1-295				FEKRSG	-YSLQLKV-ENPRVNQEIE	
TPK3_Scer/1-255					-LKQVEHTNDERR	
TPK1_Scer/1-255					-LKQVEHTNDERL	
TPK2_Scer/1-255					-MKQVEHTNDERR	
YPK1_Scer/1-256					-KSEVTHTLAERT	
YPK2_Scer/1-256					-KCEVTHTLAERT	
SCH9_Scer/1-260					-KNEIAHTIGERN	
YNR047W_Scer/1-282					-RNKIKRVLTEQE	
RIM15_Scer/1-269					-KNQVTNVKSERA	
IPL1_Scer/1-252					-YNLQKQFRREVE	
PKH2_Scer/1-265					-QKKVKYVSIEKT	
KCC4_Scer/1-265	NGN				-HSNDDSVLPYNIEREIV	
GIN4_Scer/1-271	GNVSGTSI				-VGSTTPDALPYGIEREII	
HSL1_Scer/1-289	HCSNNNGTVPNSYSSSMVTSNVSSPSIA				-SREHSNHSQTNPYGIEREIV	
SNF1_Scer/1-252	SDMQG				-RIEREIS	
KIN1_Scer/1-279	LHKEQMLPPPKN EQDVLER				-QKKLEKEISRDKRTIREAS	
KIN4_Scer/1-268	D				-ADKEIKIYREIN	
YPL141C_Scer/1-273	D				-YRKEVKIYREIN	
CMK1_Scer/1-263	NKVQL				-EALYDELD	
CMK2_Scer/1-263	NNVQL				-QMLYEELS	
RAD53_Scer/1-269	N				-MDGVTRLE	
RCK2_Scer/1-316	INGDHRKKDK				-GKDSTKTSSRDQVLKEVA	
APG1_Scer/1-302	KKLL				-ENLEIEIA	
MEK1_Scer/1-284					-PNKFDKEAR	
YKL171W_Scer/1-283					-IKKHVLREVA	
CDC5_Scer/1-256	EKTR				-KKLLSEIQ	
PSK2_Scer/1-259	DT				-WVRDRKLGTIPSEIQ	
YDL025C_Scer/1-274	EGTN				-QSQLANYSKKVTTEFC	
SKY1_Scer/1-312					-EAAEDEIK	
YAK1_Scer/1-257					-TQSITEAK	
PTK2_Scer/1-305	E				-SPEKFYKRCSKEFI	
PHO85_Scer/1-291					-EGTPSTAIREIS	
CDC28_Scer/1-288					-EGVPSTAIREIS	
SLT2_Scer/1-296					-LLCKRSLRELK	
KSS1_Scer/1-301					-LFVTRTIREIK	
FUS3_Scer/1-297					-LFALRTLREIK	
HOG1_Scer/1-280					-VLA KRTYRELK	
RIM11_Scer/1-284					-NRELE	
MRK1_Scer/1-284					-NRELE	
MCK1_Scer/1-293					-SRELO	
CKA1_Scer/1-324					-IKREIK	
CDC7_Scer/1-272					-RIYNELN	
YCK1_Scer/1-261					-QLRDEYK	
YCK2_Scer/1-261					-QLKDEYR	
YCK3_Scer/1-296					-QLRDEFR	
HRR25_Scer/1-259					-QLDYESR	
PRR1_Scer/1-317	NVEQIN				-TSLRYKETLSRLENSLTRELO	
MPS1_Scer/1-277	S				-SIDGFKGEIE	
GCN2_Scer/1-268					-LSTILSEVM	
KIN3_Scer/1-319	KE				-RQQLIAECS	
PRK1_Scer/1-277					-LNTLRAEVD	
ARK1_Scer/1-277					-LNTLRAEVD	
AKL1_Scer/1-285					-LNEMRNEVE	
VHS1_Scer/1-321	KEADMGN DIHKNSVKLQKKLAKLFKESKNVVRVPSIDLESIENMSEEDFKKLPHYKEIS					
SKM1_Scer/1-280					-KQLIMNELL	
CLA4_Scer/1-280					-KELIVNEIL	
STE20_Scer/1-252					-KELIINEIL	
CDC15_Scer/1-248					-LNDIMAEIS	
FMP48_Scer/1-285					-ANMKNEYD	
KSP1_Scer/1-260					-AMYEV	
CAK1_Scer/1-304					-PHNAKFEVS	

Table S4, continued

	121	130	140	150	160	170
TOS3_Scer/1-295	VM-----				KRCHHENVVELYEILNDPE	
TPK3_Scer/1-255	MLSIVS-----				-HPFIIRMWGTFO-----	
TPK1_Scer/1-255	MLSIVT-----				-HPFIIRMWGTFO-----	
TPK2_Scer/1-255	MLKLVE-----				-HPFLIRMWGTFO-----	
YPK1_Scer/1-256	VALARVD-----				-CPFIVPLKFSFQ-----	
YPK2_Scer/1-256	VALARVD-----				-CPFIVPLKFSFQ-----	
SCH9_Scer/1-260	ILVTTASKS-----				-SPFIVGLKFSFQ-----	
YNR047W_Scer/1-282	ILATSN-----				-HPFIVTLYHSFQ-----	
RIM15_Scer/1-269	IMMVQSD-----				-KPYVARLFASFQ-----	
IPL1_Scer/1-252	IQTSLN-----				-HPNLTKSYGYFH-----	
PKH2_Scer/1-265	ALQKLNN-----				-PSVVRLFSTFQ-----	
KCC4_Scer/1-265	IMKLLS-----				-HPNVLSLYDVWE-----	
GIN4_Scer/1-271	IMKLLN-----				-HPNVLRLYDVWE-----	
HSL1_Scer/1-289	IMKLIS-----				-HTNVMALFEVWE-----	
SNF1_Scer/1-252	YLRLLR-----				-PHPIIKLYDVIK-----	
KIN1_Scer/1-279	LGQILY-----				-PHPICRLFEMCT-----	
KIN4_Scer/1-268	ALKHLT-----				-HPNIYIYLEEVLO-----	
YPL141C_Scer/1-273	ALKHLS-----				-HPNIVVKLEEVLO-----	
CMK1_Scer/1-263	ILQRLH-----				-HPNIVAFKDWF-----	
CMK2_Scer/1-263	ILQKLS-----				-HPNIVSFKDWF-----	
RAD53_Scer/1-269	VLQQLN-----				-HPRIVRLKGFYE-----	
RCK2_Scer/1-316	LHKTVSAG-----				-CSQIVAFIDFQE-----	
APG1_Scer/1-302	ILKKIK-----				-PHHVGLIDCER-----	
MEK1_Scer/1-284	ILLRLD-----				-HPNIIKVYHTFC-----	
YKL171W_Scer/1-283	IWRTLK-----				-HNRLPPLLWKLDNN-----	
CDC5_Scer/1-256	IHKSMS-----				-HPNIVQFIDCFE-----	
PSK2_Scer/1-259	IMATLNKN-----				-SQENILKLLDFEDDD-----	
YDL025C_Scer/1-274	IGSTLH-----				-HENIVETLDMLETEG-----	
SKY1_Scer/1-312	LLQRVNDADN-----				-TKEDSMGANHILKLLDHFNHKG-----	
YAK1_Scer/1-257	ILELLNQK-----				-IDPTNKHHFLRMYDSFV-----	
PTK2_Scer/1-305	IAKHLSHN-----				-VHITNTFYLLKVPTTY-----	
PHO85_Scer/1-291	LMKELK-----				-HENIVRLYDVIHTEM-----	
CDC28_Scer/1-288	LLKELK-----				-DDNIVRLYDIVHSDA-----	
SLT2_Scer/1-296	LLRHFR-----				-GHKNITCLYDMDIVFY-----	
KSS1_Scer/1-301	LLRYFH-----				-EHENIISILDKVRPVS-----	
FUS3_Scer/1-297	ILKHFK-----				-HENIITIFNIQRPDS-----	
HOG1_Scer/1-280	LLKHLR-----				-HENLICLQDIFLS-----	
RIM11_Scer/1-284	IMKMLS-----				-HINIIDLKYFFY-----	
MRK1_Scer/1-284	TMKMLC-----				-HPNTVGLQYYFY-----	
MCK1_Scer/1-293	ILRIAD-----				-HPNIVKLQYFFT-HL-----	
CKA1_Scer/1-324	ILTDLNEKVPPPTLFPQKDQYYTNQKEVLKFIRPYIFDQPHNGHANIHLFDIICKPI					
CDC7_Scer/1-272	LLYIMT-----				-GSSRVAPLCDAKVR-----	
YCK1_Scer/1-261	TYKIL-----				-NGTPNIPYAYYFGQEG-----	
YCK2_Scer/1-261	TYKIL-----				-AGTPGIPQEYYFGQEG-----	
YCK3_Scer/1-296	AYRIL-----				-NGCVGIPHAYYFGQEG-----	
HRR25_Scer/1-259	VYRYL-----				-SGGVGIPFIRWFGREG-----	
PRR1_Scer/1-317	VLKSLN-----				-HPCIVKLLGINNPIF-----	
MPS1_Scer/1-277	LLEKLK-----				-DQKRVIQLLDYEMGDG-----	
GCN2_Scer/1-268	LLASLN-----				-HQYVVRYAAWLEED-----	
KIN3_Scer/1-319	ILSQL-----				-KHENVFYNWDFDEQ-----	
PRK1_Scer/1-277	AMKLLRNN-----				-KHSVSYIDSHAARS-----	
ARK1_Scer/1-277	AMRLLKNN-----				-RYVVSYIDSHAAKA-----	
AKL1_Scer/1-285	VMKKLKGA-----				-PNIVQYFDSNASRR-----	
VHS1_Scer/1-321	LHLRVHH-----				-HKNIVTIHEVLQS-----	
SKM1_Scer/1-280	VMNDS-----				-RQENIVNFLEAYIIDD-----	
CLA4_Scer/1-280	VMKDS-----				-RHKNIVNFLEAYLRTD-----	
STE20_Scer/1-252	VMKGS-----				-KHPNIVNFIDSYVLKG-----	
CDC15_Scer/1-248	LLKNL-----				-NHN-NIVKYHGFIRKS-----	
FMP48_Scer/1-285	VMKILSSCN-----				-PHPNICSMLDFYTDDS-----	
KSP1_Scer/1-260	IQTKIG-----				-RHQNIAALLDFFDS-----	
CAK1_Scer/1-304	IILNKLGN-----				-KCKHILPLLESKATDN-----	

Table S4, continued

	181	190	200	210	220	230	
TOS3_Scer/1-295	S-----		TKVYLVLEYCSRGPVKWCPENK-----			MEI	
TPK3_Scer/1-255	-----		DSQQVFMVMDYIEGGELFSLLRK-----				
TPK1_Scer/1-255	-----		DAQQIFMIMDYIEGGELFSLLRK-----				
TPK2_Scer/1-255	-----		DARNIFMVMMDYIEGGELFSLLRK-----				
YPK1_Scer/1-256	-----		SPEKLYFVLAFINGGELFYHLQK-----				
YPK2_Scer/1-256	-----		SPEKLYLVLAFINGGELEFYHLQH-----				
SCH9_Scer/1-260	-----		TPTDLYLVTDYMSGGELEFWHLQK-----				
YNR047W_Scer/1-282	-----		SEDYLYLCMEYCMGGEFFRALQTR-----			K	
RIM15_Scer/1-269	-----		NKDNLFVLMEMYLPGGDLATLKM-----				
IPL1_Scer/1-252	-----		DEKRVYLLMEYLVNGEMYKLLRL-----				
PKH2_Scer/1-265	-----		DESSLYFLLEYAPNGDFLSLMKK-----				
KCC4_Scer/1-265	-----		TNNNLYLILEYAEKGELFNLLVD-----				
GIN4_Scer/1-271	-----		TNTDLYLVLEYAEKGELFNLLVE-----				
HSL1_Scer/1-289	-----		NKSELYLVLEYVDGGELFDYLV-----				
SNF1_Scer/1-252	-----		SKDEIIMVIEYA-GNELEFDYIVQ-----				
KIN1_Scer/1-279	-----		LSNHFYMLFEYVSGGQLLDYIIQ-----				
KIN4_Scer/1-268	-----		NSKYIGIVLEFVSGGEFYKYIQR-----				
YPL141C_Scer/1-273	-----		NSRYIGIVLEYACGGEFYKYIQR-----				
CMK1_Scer/1-263	-----		SKDKFYIITQLAKGGLFDRILK-----				
CMK2_Scer/1-263	-----		SKDKFYIWTQLATGGELFDRILS-----				
RAD53_Scer/1-269	-----		DTESYYVMEFFVSGGDLMDFVAA-----				
RCK2_Scer/1-316	-----		TDSYYYIIQELLTGGEIFGEIVR-----				
APG1_Scer/1-302	-----		TSTDFYLIMEYCALGDLTFLLKR-----				
MEK1_Scer/1-284	-----		RNNHLYIFQDLIPGGDLFSYLAKG-----DC				
YKL171W_Scer/1-283	-----		YAMYCLTERINDGTLYDLVISW-----				
CDC5_Scer/1-256	-----		DDSNVYILLEICPNGLSMELLKR-----				
PSK2_Scer/1-259	YYY-----		IETPVHGETGSIDLFDVIEF-----				
YDL025C_Scer/1-274	--D-----		TYLLVMEMEYAPY-DFFNLVMS-----				
SKY1_Scer/1-312	P-----		NGVHVVMVFEVLGENLLALIKKY-----				
YAK1_Scer/1-257	HKN-----		HLCLVFELLSNNLYELLKO-----				
PTK2_Scer/1-305	TTR-----		GWGFIMEL-GVKDLEFQLMER-----				
PHO85_Scer/1-291	-----		KLTLVFEF-MDNDLKYYMDS-----				
CDC28_Scer/1-288	H-----		KLYLVFEF-LDLDLKRYMEG-----				
SLT2_Scer/1-296	PDG-----		SINGLYLYEEL-MECDMHQIIKSGQ-----				
KSS1_Scer/1-301	ID-----		KLNavyLVEEL-METDLQKVINNONS-----				
FUS3_Scer/1-297	FE-----		NFNEVYIIQEL-MQTDLHRVISTQ-----				
HOG1_Scer/1-280	-----		PLEDIYFVTEL-QGTDLHRLLQTR-----				
RIM11_Scer/1-284	ERDSQ-----		DEIYLNLILEY-MPQSLYQRLRH-VHQRT-----				
MRK1_Scer/1-284	EKDEE-----		DEVYLNVLVDY-MPQSLYQRLRH-VNLKM-----				
MCK1_Scer/1-293	SPQDN-----		KVYQHЛАМЕС-LPETLQIEINRY-VTNKL-----				
CKA1_Scer/1-324	S-----		KTPALVFEYVDNVD-FRILY-----				
CDC7_Scer/1-272	-----		DQVIAVLPYYPPHEE-FRTFYR-----				
YCK1_Scer/1-261	L-----		HNILVIDLLGP-SLEDFD-----				
YCK2_Scer/1-261	L-----		HNILVIDLLGP-SLEDFD-----				
YCK3_Scer/1-296	M-----		HNILIIDLLGP-SLEDFEW-----				
HRR25_Scer/1-259	E-----		YNAMVIDLLGP-SLEDFNY-----				
PRR1_Scer/1-317	VTSKKPLCDLIICKTPRALPPCDMIMSYPAGDLIAAVMAR-----		LLYLIMECGDHDLSQILNO-----				
MPS1_Scer/1-277	S-----		MLFIIQMEYCENRTLYDLIHS-----				
GCN2_Scer/1-268	K-----		EVLYLYMЕYCSRГDLSQMIKH-----YKQEHKY				
KIN3_Scer/1-319	VNG-----		IAYEVFVLMEFCERGGLIDFMNT-----RLQNR				
PRK1_Scer/1-277	MLHN-----		GSYEVFVLMEYCERGGLIDFMNT-----RLQNR				
ARK1_Scer/1-277	RDGV-----		QGFEVLLLMELCPNKSLLDMNQ-----RLSTK				
AKL1_Scer/1-285	-----		AVCTFIVMDYPTDLFTSIVDN-----				
VHS1_Scer/1-321	E-----		ELWVIMEYMEGGCLTDILDA-----VARSNTG				
SKM1_Scer/1-280	D-----		DLWVVMEMEFGGSLTDIIEN-----SPTNDN				
CLA4_Scer/1-280	-----		DLWVIMEYMEGGSLTDVVTH-----				
STE20_Scer/1-252	-----		ELYILLEYCANGSLRRLISR-----				
CDC15_Scer/1-248	Y-----		YYIMVLEYCECGDLYDFLDIAK-----SQGS				
FMP48_Scer/1-285	-----		YIIMEYCSGGDLYEAIKAD-----				
KSP1_Scer/1-260	N-----		DLLLFPFEEMNLYEFMQM-----HYKRDRRKKNPYYD				
CAK1_Scer/1-304	-----						

Table S4, continued

	241	250	260	270	280	290
TOS3_Scer/1-295	KA--VGP-----			SILTFQQSRKVVLVDVSGLEYLHSQG-----		
TPK3_Scer/1-255	SQ--RFP-----			-NPVAKFYAAEVCLALEYLHSKD-----		
TPK1_Scer/1-255	SQ--RFP-----			-NPVAKFYAAEVCLALEYLHSKD-----		
TPK2_Scer/1-255	SQ--RFP-----			-NPVAKFYAAEVILALEYLHAWN-----		
YPK1_Scer/1-256	EG--RFD-----			LSRARFYTAELLCALDNLHKLD-----		
YPK2_Scer/1-256	EG--RFS-----			LARSRFYIAELLCALDSLHKLD-----		
SCH9_Scer/1-260	EG--RFS-----			EDRAKFYIAELVLALEHLHDND-----		
YNR047W_Scer/1-282	TK--CIC-----			EDDARFYASEVTAALEYLHLLG-----		
RIM15_Scer/1-269	MG--YLP-----			DQWAKQYLTEIVGVNDMHQNG-----		
IPL1_Scer/1-252	HG--PFN-----			DILASDYIYQIANALDYMHKKN-----		
PKH2_Scer/1-265	YG--SLD-----			ETCARYYYAAQIIDAIIDYLHSNG-----		
KCC4_Scer/1-265	HG--PLP-----			EREAINCFRQIIIGISYCHALG-----		
GIN4_Scer/1-271	RG--PLP-----			EHEAIRFFRQIIIGVSYCHALG-----		
HSL1_Scer/1-289	KG--KLP-----			EREAIHYFKQIIVEGVSYCHSFN-----		
SNF1_Scer/1-252	RD--KMS-----			EQEARRFFQQIISAVEYCHRHK-----		
KIN1_Scer/1-279	HG--SIR-----			EHQARKFARGIASALIYLNHANN-----		
KIN4_Scer/1-268	KR--RLK-----			ESSACRLFAQLISGVNYMHKG-----		
YPL141C_Scer/1-273	KR--RLK-----			EMNACRLFSQLISGVHYIHSGK-----		
CMK1_Scer/1-263	KG--KFT-----			EEDAVRILVEILSAVKYMHQSQN-----		
CMK2_Scer/1-263	RG--KFT-----			EVDAVEIIVQILGAVEYMHSKN-----		
RAD53_Scer/1-269	HG--AVG-----			EDAGREISRQILTAIKYIHSGM-----		
RCK2_Scer/1-316	LT--YFS-----			EDILSRHVIKQLALAVKHMHSLG-----		
APG1_Scer/1-302	RK--ELMENHPLLRTVFEKYPPPSENHNGLHRAFVLSYLOOLASALKFLRSKN-----					
MEK1_Scer/1-284	LT--SMS-----			ETESLLIVFQILQALNYLHDQD-----		
YKL171W_Scer/1-283	DEFKRKSKIPFAER-----			CRLTIFLSLQLLSALKYMHSKT-----		
CDC5_Scer/1-256	RK--VLT-----			EPEVRRFFTTCQICGAIKYMHSSR-----		
PSK2_Scer/1-259	KK--DMV-----			EHEAKLVFKQVVVASIKHLHDQG-----		
YDL025C_Scer/1-274	N--LMT-----			QDEVNCYFKQLCHGKVNYLHSMG-----		
SKY1_Scer/1-312	EH--RG-----			IPLIYVKQISKQLLLGLDYMHRRCG-----		
YAK1_Scer/1-257	NKF--HG-----			LSIQQLIRFTTQTQILDSDLCVLKESK-----		
PTK2_Scer/1-305	TG--WKN-----			VPFNEKYCLFKQVAQGIKFCHDNG-----		
PHO85_Scer/1-291	----RTVGN-----			TPRGLELNLVKYFQWQOLLOGLAFCHENK-----		
CDC28_Scer/1-288	----IPKDO-----			PLGADIVKKFMMQLCKGIAYCHSHR-----		
SLT2_Scer/1-296	-----			PLTDAHYQSFTYQILCGLKYIHSAD-----		
KSS1_Scer/1-301	-----			GFSTLSDDHVQYFTYQILRALKSIHSAQ-----		
FUS3_Scer/1-297	-----			MLSDDHIQYFIYTQTLRAVKVLHGSN-----		
HOG1_Scer/1-280	-----			PLEKQFVQYFLYQILRGLKYVHSAG-----		
RIM11_Scer/1-284	-----			PMSRLEIKYYMFQLFKSLNYLHHFAN-----		
MRK1_Scer/1-284	-----			QMPRVEIKFYAYQLFKALNYLHNVPR-----		
MCK1_Scer/1-293	-----			EMPLKHIRLYTYQIARGMLYLHGLG-----		
CKA1_Scer/1-324	-----			KLTDLERIFRYMFELLKALDYCHSMG-----		
CDC7_Scer/1-272	-----			DLPIKGICKYIWELLRALKFVHSKG-----		
YCK1_Scer/1-261	CG--R-----			KFSVKTVVQVAVQMITLIEDLHAHD-----		
YCK2_Scer/1-261	CG--R-----			RFSVKTVVQVAVQMITLIEDLHAHD-----		
YCK3_Scer/1-296	CG--R-----			KFSVKTTCMVAQQMIDRVRAIHDHD-----		
HRR25_Scer/1-259	CH--R-----			RFSFKTVIMLALQMFCRIQYIHGRS-----		
PRR1_Scer/1-317	NG--RLE-----			AWLIQRIFTEVVLAVKYLHENS-----		
MPS1_Scer/1-277	RSGMPLD-----			FNFVRFTYKEMLLCIKVVHDAG-----		
GCN2_Scer/1-268	ENLNQQ-----			RDEYWRLFRQILEALSYIHSQG-----		
KIN3_Scer/1-319	----IP-----			EKIVWGILAQLLTALYKCHYGVELPTLTT		
PRK1_Scer/1-277	----LQ-----			ESEILEIMSQTQVGITAMHALQP-----		
ARK1_Scer/1-277	----LH-----			EFEILQIMSQTQGVAAHALQP-----		
AKL1_Scer/1-285	----LT-----			EAEIVKIMYDVALSISQMHYLPV-----		
VHS1_Scer/1-321	-R--HFVTN-----			GLLVKKVFLQICSALNYCHEHG-----		
SKM1_Scer/1-280	EHSSPLN-----			ENQMAYIVKETCQGLKFLHNK-----		
CLA4_Scer/1-280	SH--SPLT-----			EPOQIAYIVRETCQGLKFLHDK-----		
STE20_Scer/1-252	CI--LT-----			EGOIGAVCRETLSGLEFLHSK-----		
CDC15_Scer/1-248	-SSTGLS-----			ENESKTYVTQTLGLKYLHGE-----		
FMP48_Scer/1-285	PSSPSL-----			QIDMQKIIKQLCSAISFAHSLG-----		
KSP1_Scer/1-260	----AVPKK-----			TKSITHIITQIMDAIEYVHNKG-----		
CAK1_Scer/1-304	LL--NPS--IPIV-----			ADPPVQKYTNQLDVNRYSLSFFRQMVEGIAFLHENK-----		

Table S4, continued

	301	310	320	330	340	350
TOS3_Scer/1-295		-----I	THRDIKPSNLLI		SSNG	-----
TPK3_Scer/1-255		-----I	IYRDLKPENILL		DKNG	-----
TPK1_Scer/1-255		-----I	IYRDLKPENILL		DKNG	-----
TPK2_Scer/1-255		-----I	IYRDLKPENILL		DRNG	-----
YPK1_Scer/1-256		-----V	VYRDLKPENILL		DYQG	-----
YPK2_Scer/1-256		-----V	IYRDLKPENILL		DYQG	-----
SCH9_Scer/1-260		-----I	IVYRDLKPENILL		DANG	-----
YNR047W_Scer/1-282		-----F	IYRDLKPENILL		HQSG	-----
RIM15_Scer/1-269		-----I	IHHDLKPENILL		DNAG	-----
IPL1_Scer/1-252		-----I	IHRDIKPENILI		GFNN	-----
PKH2_Scer/1-265		-----I	IHRDIKPENILL		DGEM	-----
KCC4_Scer/1-265		-----I	IVHDLKPENILL		DSFY	-----
GIN4_Scer/1-271		-----I	IVHDLKPENILL		DHKY	-----
HSL1_Scer/1-289		-----I	ICRDLKPENILL		DKKN	-----
SNF1_Scer/1-252		-----I	IVHDLKPENILL		DEHL	-----
KIN1_Scer/1-279		-----I	IVHDLKIENIMI		SDSS	-----
KIN4_Scer/1-268		-----L	VHDLKLENLLL		DKHE	-----
YPL141C_Scer/1-273		-----L	VHDLKLENLLL		DKNE	-----
CMK1_Scer/1-263		-----I	IVHDLKPENLLY		IDKSDES	-----
CMK2_Scer/1-263		-----V	VHDLKPENVLY		VDKSENS	-----
RAD53_Scer/1-269		-----I	SHRDLKPDNILI		EQDDP	-----
RCK2_Scer/1-316		-----V	VHDLIKPENLLFEPIEFTRSIKPKLRSDD			
APG1_Scer/1-302		-----L	VHDLIKPQNLLI		STPLIGY	-----
MEK1_Scer/1-284		-----I	VHDLKLKDNLILL		CTPEP	-----
YKL171W_Scer/1-283		-----I	VHDLIKLENCLL		QKEGKK	-----
CDC5_Scer/1-256		-----V	IHRDLKLGNIFF		DSNY	-----
PSK2_Scer/1-259		-----I	IHRDLKDENVIV		DSHG	-----
YDL025C_Scer/1-274		-----L	AHDLKLDNCVV		TKDG	-----
SKY1_Scer/1-312		-----I	IHTDIKPENVLM		EENLI	-----
YAK1_Scer/1-257		-----L	IHCDLKPENILL		CAP	-----
PTK2_Scer/1-305		-----I	AHDLKPENVLI		SKEG	-----
PHO85_Scer/1-291		-----I	IHRDLKPQNLLI		NKR	-----
CDC28_Scer/1-288		-----I	IHRDLKPQNLLI		NKD	-----
SLT2_Scer/1-296		-----V	VHDLKPGNLLV		NAD	-----
KSS1_Scer/1-301		-----V	IHRDIKPNSNLL		NSN	-----
FUS3_Scer/1-297		-----V	IHRDLKPSNLLI		NSN	-----
HOG1_Scer/1-280		-----V	IHRDLKPSNILI		NEN	-----
RIM11_Scer/1-284		-----V	CHRDIKPQNLLV		DPET	-----
MRK1_Scer/1-284		-----I	CHRDIKPQNLLV		DPTT	-----
MCK1_Scer/1-293		-----V	CHRDIKPNSNVLV		DPET	-----
CKA1_Scer/1-324		-----I	IMHRDVKPHNVMI		DHKN	-----
CDC7_Scer/1-272		-----I	IHRDIKPTNFLF		NLEL	-----
YCK1_Scer/1-261		-----L	IYRDIKPDNFLI		GRPG	-----
YCK2_Scer/1-261		-----L	IYRDIKPDNFLI		GRPG	-----
YCK3_Scer/1-296		-----L	IYRDIKPDNFLI		SQYQRISPEGKVIKSCA	-----
HRR25_Scer/1-259		-----F	IHRDIKPDNFLM		GVGR	-----
PRR1_Scer/1-317		-----I	IHRDLKLENILL		KYSFDDIN	-----
MPS1_Scer/1-277		-----I	IHSVLDLK PANFVL		VKG	-----
GCN2_Scer/1-268		-----I	IHRDLKPMNIFI		DESR	-----
KIN3_Scer/1-319		-----I	IYDRMKPPVKGNIVIHRDLKPGNIFLSYDDS-D		YNINEQVDGHEE	VNSNY
PRK1_Scer/1-277		-----P	LIHRDIKIENVI		SHDG	-----
ARK1_Scer/1-277		-----P	LIHRDIKIENVI		SANN	-----
AKL1_Scer/1-285		-----S	LIHRDIKIENVL		DAKN	-----
VHS1_Scer/1-321		-----I	IYHCDIKPENLL		DTED	-----
SKM1_Scer/1-280		-----K	IHRDIKSDNILL		NSQG	-----
CLA4_Scer/1-280		-----H	IHRDIKSDNILL		DTRA	-----
STE20_Scer/1-252		-----G	VLHRDIKSDNILL		SMEG	-----
CDC15_Scer/1-248		-----G	VIHRDIKAANILL		SADN	-----
FMP48_Scer/1-285		-----I	AHDLKPENILL		TING	-----
KSP1_Scer/1-260		-----I	IHRDIKPENILI		SGIDW	-----
CAK1_Scer/1-304		-----I	IHRDIKPQNIML		TNNNTSTVS	-----

Table S4, continued

	361	370	380	390	400	410
TOS3_Scer/1-295				TVKISDFGVAMST-ATGSTNIQ		SSHE
TPK3_Scer/1-255				HIKITDFGFAKYVP		
TPK1_Scer/1-255				HIKITDFGFAKYVP		
TPK2_Scer/1-255				HIKITDFGFAKEVQ		
YPK1_Scer/1-256				HIALCDFGLCKLNM		KD
YPK2_Scer/1-256				HIALCDFGLCKLNM		KD
SCH9_Scer/1-260				NIALCDFGLSKADL		KD
YNR047W_Scer/1-282				HIMLSDFDLSIQAKDSKVPPVKGSAQSTLVDTKICSD		
RIM15_Scer/1-269				HVKLTDFGLSRAGLI		RRH
IPL1_Scer/1-252				VIKLTDFFGWSIINP		P
PKH2_Scer/1-265				KIKLTDFGTAKLLN-PTNNNSVSK		PEYDL
KCC4_Scer/1-265				NIKIADFGMAALQT		D
GIN4_Scer/1-271				NIKIADFGMAALET		E
HSL1_Scer/1-289				RRIKIADFGMAALEL		P
SNF1_Scer/1-252				NVKIADFGLSNIMT		D
KIN1_Scer/1-279				EIKIIDFGLSNIYD		S
KIN4_Scer/1-268				NLVITDFGFVNNEFF		ED
YPL141C_Scer/1-273				NLVITDFGFVNNEFC		SR
CMK1_Scer/1-263				PLVVADFGIAKRLKS		D
CMK2_Scer/1-263				PLVIADFGIAKQLKG		E
RAD53_Scer/1-269				VLVKITDFGLAKVQG		N
RCK2_Scer/1-316				PQTKADEGIFTPGVGFFFFGIVKLADFGGLSKQIFS		
APG1_Scer/1-302				HDSKSFHELGFVGIIYNLPILKIADFGFARFLP		N
MEK1_Scer/1-284				CTRIVLADFGIAKDLN		SN
YKL171W_Scer/1-283				DWKVFLCDFGMSCHFD		EKH
CDC5_Scer/1-256				NLKIGDFGLAAVLA		NE
PSK2_Scer/1-259				FVKLIDFGSAAYIKSG		
YDL025C_Scer/1-274				ILKLIDFGSAAVVFQYP		YEDT
SKY1_Scer/1-312				QIKIADLGNAWCYD		
YAK1_Scer/1-257				DKPELKIIDFGSSCEEAA		
PTK2_Scer/1-305				ICKLTDFFGISDWYHVIPH		DYTSP
PHO85_Scer/1-291				GQLKLGDFGLARAFIG		IP
CDC28_Scer/1-288				GNLKLGDGFGLARAFIG		VP
SLT2_Scer/1-296				CQLKICDFGLARGYSENPVENSO		
KSS1_Scer/1-301				CDLKVCDFGLARCLA-SSSDSRE		TL
FUS3_Scer/1-297				CDLKVCDFGLARIIDESAADNSE		PTGQ
HOG1_Scer/1-280				CDLKICDFGLARIQD		
RIM11_Scer/1-284				WSLKLCDFGSAKQLKP		
MRK1_Scer/1-284				FSFKICDFGSAKCLKP		
MCK1_Scer/1-293				GVLKICDFGSAKKLEH		
CKA1_Scer/1-324				KKLRLIDWGGLAEFYHV		
CDC7_Scer/1-272				GRGVLVDFGLAEAQMDY		KSTR
YCK1_Scer/1-261				QPDANNHLIDFGMAKQYRDPK		TKQHIP
YCK2_Scer/1-261				QPDANKVHLIDFGMAKQYRDPK		TKQHIP
YCK3_Scer/1-296				SSNNNDPNLIYMVDFGMAKQYRDPK		TKQHIP
HRR25_Scer/1-259				RGSTVHVIDFGGLSKKYRDFN		THRHIP
PRR1_Scer/1-317				SFRDSPIYCKQNFIELADFGLCKKIE		N
MPS1_Scer/1-277				ILKIIDFGIANAVP		EHTV
GCN2_Scer/1-268				NVKIGDFGLAKNVHRSLDILK		LDSQNLPGS
KIN3_Scer/1-319				RDHRVNSGKRGPMDYSQV	VVKLGDFGLAKSLE	TS
PRK1_Scer/1-277				LYKVDFFGSVSGVIRPPR		NTQEFNY
ARK1_Scer/1-277				EYKLCDFGSVCGIIRPPR		NSQELSY
AKL1_Scer/1-285				NFKLADFGSTSTCFPIVT		THQDIAL
VHS1_Scer/1-321				NVFLCDFGLSTTST		
SKM1_Scer/1-280				LVKITDFGFCAVLT		EK
CLA4_Scer/1-280				RVKITDFGFCAVLT		DK
STE20_Scer/1-252				DIKLTDFGFCAQIN		EL
CDC15_Scer/1-248				TVKLADFGVSTIVN		
FMP48_Scer/1-285				DIKLADWGHAIQSP		
KSP1_Scer/1-260				TIKLTDWGLATTDK		
CAK1_Scer/1-304				PKLYIIDFGISYDMANN		SQTSQEP

Table S4, continued

	421	430	440	450	460	470
TOS3_Scer/1-295	QLKSRALGTPAFFAPELCSTEKEYS-----				CSSAIDIWLSGVTIYCLLF-	
TPK3_Scer/1-255	-DVTYTLCGTPDYIAPEVST-----				KPYNKSVDWWSGVLIYEMLA-	
TPK1_Scer/1-255	-DVTYTLCGTPDYIAPEVST-----				KPYNKSIDWWSGILYEMLA-	
TPK2_Scer/1-255	-TVTWTLCGTPDYIAPEVITT-----				KPYNKSVDWWSLGVLIYEMLA-	
YPK1_Scer/1-256	DDKTDTFCGTPEYLAPELLL-----				LGYTKAVDWWTLGVLLYEMLT-	
YPK2_Scer/1-256	NDKTDTFCGTPEYLAPEILLG-----				OQYTKTVDWWTLGILLYEMMT-	
SCH9_Scer/1-260	--RTNTFCGTTEYLAPELLLDE-----				TGYTKMVDFWSLGVLIFEMCC-	
YNR047W_Scer/1-282	GFRNTNSFVGTEEYIAPEVIRG-----				NGHTAAVDWWTLGILLYEMLF-	
RIM15_Scer/1-269	KKQNKKFFGTPDYLAPETIEK-----				GEDNKQCDWWSGCIFFEPLL-	
IPL1_Scer/1-252	ENRRKTVCGTIDYLSPEMVES-----				REYDHTIDAALGVLAELLT-	
PKH2_Scer/1-265	STRSKSFVGTAEYVSPELLND-----				SFTDYRCDIWAFGCILFQMIA-	
KCC4_Scer/1-265	ADLLETSCGSYPHYAAPEIVSG-----				LPYEGFASDVWSCGVILFALLT-	
GIN4_Scer/1-271	GKLLLETSCGSYPHYAAPEIVSG-----				IPYQGFASDVWSCGVILFALLT-	
HSL1_Scer/1-289	NKLLKTSCGSYPHYASPEIVMG-----				RPYHGGPSDVWSCGIVLFALLT-	
SNF1_Scer/1-252	GNFLKTSCGSYPHYAAPEVISG-----				KLYAGPEVDVWSCGVILYVMLC-	
KIN1_Scer/1-279	RKQLHTFCGSLYFAPELLKA-----				NPYTGPEVDVWSFGVVLFLVLC-	
KIN4_Scer/1-268	NELMKTSCGSPCYAAPELVVST-----				KAYEARAKDVWSCGVILYAMLA-	
YPL141C_Scer/1-273	NELMKTSCGSPCYAAPELVISA-----				EPYEARKADIWSCGVILYAILA-	
CMK1_Scer/1-263	EELLYKPAGSLGYVAPEVLT-----				QDGHGKPCDIWSIGVITYTLLC-	
CMK2_Scer/1-263	EDLIYKAAGSLGYVAPEVLT-----				QDGHGKPCDIWSIGVITYTLLC-	
RAD53_Scer/1-269	GSFMKTFCGTLAYVAPEVIRGKDTSVSPDEYEE-----				RNEYSSLVDMWSMGCLVYVILT-	
RCK2_Scer/1-316	-KNTKTPCGTGVGYTAPEVVKD-----				EHYSMKVDMWGIGCVLYTMLC-	
APG1_Scer/1-302	TSLAETLCGSPLYMAPEILN-----				YOKYNAKADLWSVGTVVFEMCC-	
MEK1_Scer/1-284	KERMHTVVGTPYCAPEVGFRANRKAYQSFSRAATLEQRGYDSKCDLWSLGVITHIMLT-					
YKL171W_Scer/1-283	EPEPSKYIGSLPYASPELDFP-----				CIVSPLGPASDIWALGVMLYTMLV-	
CDC5_Scer/1-256	SERKYTICGTPNYIAPEVLMGK-----				HSGHSFEVDIWSLGVMLYALLI-	
PSK2_Scer/1-259	--PFDVFVGMTMDYIAPEVLLGGSS-----				YKGKPQDIWALGVLLYTIY-	
YDL025C_Scer/1-274	IVKSHGIVGSDPYLAPELLKQT-----				SYDPRVADVWSIAIIFYCMLV-	
SKY1_Scer/1-312	-EHYTNISIQTREYRSPEVLLG-----				APWCGCADIWSTAELFELIT-	
YAK1_Scer/1-257	-RTVYTYIQSRFYRAPEIILG-----				IPYSTSIDMWWSLGIVAEFL-	
PTK2_Scer/1-305	VKTCQGMIGSPPYTPPEVMYFDAKKHYPEK-----				FQKPYNPLAMDSYALGIMLITMIN-	
PHO85_Scer/1-291	VNTFSSEEVTLWYRAPDVLMGS-----				RTYSTSIDIWSCGCILAEMIT-	
CDC28_Scer/1-288	LRAYTHEIVTLWYRAPEVLLGG-----				KQYSTGVDTWSIGCIFAEMCN-	
SLT2_Scer/1-296	--FLTEYVATRWYRAPEIMLSY-----				QGYTKAIDVWSAGCILAEFLG-	
KSS1_Scer/1-301	VGFMTTEYVATRWYRAPEIMLTF-----				QEYTTAMDIWSCGCILAEMVS-	
FUS3_Scer/1-297	QSGMTEYVATRWYRAPEVMLTS-----				AKYSRAMDVWSCGCILAELFL-	
HOG1_Scer/1-280	-PQMTGYVSTRYYRAPEIMLTW-----				QKYDVEVDIWSAGCIFAEMIE-	
RIM11_Scer/1-284	TEPNVSYICSRYYRAPELIFGA-----				TNYTNQIDIWSSGCVMAELL-	
MRK1_Scer/1-284	DQPNVSYICSRYYRAPELMFGA-----				TNYSNQVDVWSSACVIAELL-	
MCK1_Scer/1-293	NQPSISYICSRFYRAPELIIGC-----				TQYTTQIDIWGLGCVGMGEMLI-	
CKA1_Scer/1-324	NMEYNVRVASRFFKGPELLVDY-----				RMDYSDLWLSFGTMLASMIF-	
CDC7_Scer/1-272	RIKRANRAGTRGFRAPEVLMKC-----				GAQSTKIDIWSVGVILLSLLG-	
YCK1_Scer/1-261	YREKKSLSGTARYMSINTHLG-----				REQSRRDDMEALGHVFFYFLR-	
YCK2_Scer/1-261	YREKKSLSGTARYMSINTHLG-----				REQSRRDDMEAMGHVFFYFLR-	
YCK3_Scer/1-296	YRERKSLSGTARYMSINTHFG-----				REQSRRDDLESIGHVFFYFLR-	
HRR25_Scer/1-259	YRENKSLTGTARYASVNTHLG-----				YRENKSLTGTARYASVNTHLG-----	
PRR1_Scer/1-317	NEMC TARCGSEDYVSPEILMGP-----				IEQSRRDDLESIGVLIYFCK-	
MPS1_Scer/1-277	NIYRETOQIGTPNYMAPEALVAMNYTQNSENQHEG-----				YDGHLSDTWALGVILYSLFE-	
GCN2_Scer/1-268	NWKVGRPSDMWSGCIIYQMIY-----				NRGASTLSDIWSLGATVVEMLT-	
KIN3_Scer/1-319	SDNLNTSAIGTAMYVATEVLDGTG-----				KEPYDCAKVDLWAMGIVFLNIVF-	
PRK1_Scer/1-277	IQFATTYVGTPYYMSPEVLDQP-----				MDSKVTDISTGIYKAPEVLFQ-----	
ARK1_Scer/1-277	VQHDILNTTAQYRSPEMIDLYR-----				VKCYDGGDVDWSLIIISQWFQR	
AKL1_Scer/1-285	VQQDILKNTTAQYRSPEMIDTFR-----					
VHS1_Scer/1-321	LTQNIYVHTTPQYRSPEMIDLYR-----					
SKM1_Scer/1-280	YIKPNVCIGSSYYMPPERISFDGRVSSSKSGG-----					
CLA4_Scer/1-280	RSKRATMVGTPYWMAPPEIVN-----					
STE20_Scer/1-252	RSKRATMVGTPYWMAPPEVVK-----					
CDC15_Scer/1-248	NLKRTTMVGTPYWMAPPEVVS-----					
FMP48_Scer/1-285	-SSALTLAGTLNWMAPEILG-----					
KSP1_Scer/1-260	-KSNDFOQIGTDNYRAPETFSGRVSNSCFKKNFDRSSAPLYNTQADYWSLGATIFYLMF-					
CAK1_Scer/1-304	-TSMDRNVGSERYMSPELFDNSLDIKER-----					

Table S4, continued

	481	490	500	510	520	530
TOS3_Scer/1-295	-----GKLPFNANS-----	-----GLELFDSIINKPLEFPSY-----				
TPK3_Scer/1-255	-----GYTPFYNSN-----	-----TMKTYENILNAELKFP-----				
TPK1_Scer/1-255	-----GYTPFYDSN-----	-----TMKTYEKILNAELRFP-----				
TPK2_Scer/1-255	-----GYTPFYDTT-----	-----PMKTYEKILQGKVVYP-----				
YPK1_Scer/1-256	-----GLPPYYDED-----	-----VPKMYKKILQEQPLVFP-----				
YPK2_Scer/1-256	-----GLPPYYDEN-----	-----VPVMYKKILQQPLLFP-----				
SCH9_Scer/1-260	-----GWSPFFAEN-----	-----NQKMYQKIAFGKVKFPR-----				
YNR047W_Scer/1-282	-----GFTPFKGDN-----	-----TNETFTNLKNEVSFPNN-----				
RIM15_Scer/1-269	-----GYPPFHAET-----	-----PDAVFKKILSGVIQWPEFK-----				
IPL1_Scer/1-252	-----GAPPFEEEM-----	-----KDTTYKRIAALDIKMP-----				
PKH2_Scer/1-265	-----GKPPFKATN-----	-----EYLTTFQKVMVKVQYAFTPG-----				
KCC4_Scer/1-265	-----GRLPFDEEN-----	-----GNVRDILLKVQKGQF-----				
GIN4_Scer/1-271	-----GRLPFDEED-----	-----GNIRTLLLKVQKGEF-----				
HSL1_Scer/1-289	-----GHLPFNDD-----	-----NIKKLLLKVQSGKY-----				
SNF1_Scer/1-252	-----RRLPFDDE-----	-----SIPVLFKNISNGVY-----				
KIN1_Scer/1-279	-----GKVPFDDE-----	-----NSSVLHEKIKQGKV-----				
KIN4_Scer/1-268	-----GYLPWDDDHEN-----	-----PTGDDIARLYKYITQTPL-----				
YPL141C_Scer/1-273	-----GYLPWDDDPNN-----	-----PEGSDIGRLNYINSTPL-----				
CMK1_Scer/1-263	-----GYSAFRAER-----	-----VQDFLDECCTTGEPV-----				
CMK2_Scer/1-263	-----GYSPIFAES-----	-----VEGFMEECTASRYPV-----				
RAD53_Scer/1-269	-----GHLPFSGST-----	-----QDQLYKQIGRGSY-----				
RCK2_Scer/1-316	-----GFPPFYDEK-----	-----IDTLTEKISRGEY-----				
APG1_Scer/1-302	-----GTPPFRASN-----	-----HLELFKKIKRAN-----				
MEK1_Scer/1-284	-----GISPFYGDG-----	-----SERSIIQNAKIGKLN-----				
YKL171W_Scer/1-283	-----GKLPFNHEF-----	-----EPRLRSLIKVGEGDRFS-----				
CDC5_Scer/1-256	-----GKPPFQARD-----	-----VNTIYERIKCR-----				
PSK2_Scer/1-259	-----KENPYYNI-----	-----DEILEGELRFDKS-----				
YDL025C_Scer/1-274	-----KRFPWKAP-----	-----KKSFNSFRLF-----	-----EEPEDED-----			
SKY1_Scer/1-312	-----GDFLFEPDEGHS-----	-----YTKDDDHIQAQIIIELLGELPSYLLRNGKYTR-----				
YAK1_Scer/1-257	-----GIPIFPGAS-----	-----EYNOLTRIIDTLGYP-----				
PTK2_Scer/1-305	-----NIIPFIDS-----	-----CNTDARFREFEVSYDNFINHQNP-----	-----HFRDKGCHK-----			
PHO85_Scer/1-291	-----GKPLFPGTN-----	-----DEEQQLKLIFDIMGTP-----	-----NESLWPSVTK-----			
CDC28_Scer/1-288	-----RKPIFSGDS-----	-----EIDQIFKIFRVLGTP-----	-----NEAIWPDIVY-----			
SLT2_Scer/1-296	-----GKPIFKGKD-----	-----YVNQLNQILQVLGTP-----	-----PDETLLRIGSK-----			
KSS1_Scer/1-301	-----GKPLFPGRD-----	-----YHHQLWLILEVLGTP-----	-----SFEDFNQIKSK-----			
FUS3_Scer/1-297	-----RRPIFPGRD-----	-----YRHQLLLIFGIIGTPHSNDLRCIESP-----				
HOG1_Scer/1-280	-----GKPLFPGKD-----	-----HVHQFSIITDLLGSP-----	-----PKDVINTICSE-----			
RIM11_Scer/1-284	-----GQPMFPGES-----	-----GIDQLVEIIKILGTP-----	-----SKQEICSMNPN-----			
MRK1_Scer/1-284	-----GKPLFSGES-----	-----GIDQLVEIIKIMGIP-----	-----TKDEISGMNPN-----			
MCK1_Scer/1-293	-----GKAIFQGQE-----	-----PLLQLREIAKLLGPP-----	-----DKRFFFSNPA-----			
CKA1_Scer/1-324	-----KREPFFHGT-----	-----NTDQLVKIVKVLGTSDFEKYLKYEIT-----				
CDC7_Scer/1-272	-----RRFPMFQSLD-----	-----DADSLELCTIFGWKELRKCAALHGLFEASGLID-----				
YCK1_Scer/1-261	-----GHLWPQGLKAP-----	-----NNKQKYEKIG-----				
YCK2_Scer/1-261	-----GQLWPQGLKAP-----	-----NNKQKYEKIG-----				
YCK3_Scer/1-296	-----GSLWPQGLKAP-----	-----NNKQKYEKIG-----				
HRR25_Scer/1-259	-----GSLWPQGLKAT-----	-----TKKQKYDRIM-----				
PRR1_Scer/1-317	-----DRLPFDPNNAS-----	-----ARQRSRATSHRIARFDWR-----				
MPS1_Scer/1-277	-----GKPPYGSFQ-----	-----GQNRLLAIMNPDVKIPFPEHTS-----				
GCN2_Scer/1-268	-----PFST-----	-----GMERVNILKKLRSVSIE-----				
KIN3_Scer/1-319	-----LHPPFQAKN-----	-----YLEIQTKEKNG-----				
PRK1_Scer/1-277	-----YTTPFEKS-----	-----GEAGILHARYQYPSFP-----				
ARK1_Scer/1-277	-----YTTPFEKG-----	-----GDLAILSGKFEFPFLYP-----				
AKL1_Scer/1-285	-----FTTPFEMT-----	-----GQFAILHSKYEFPVN-----				
VHS1_Scer/1-321	-----IRNPWLKA-----	-----DKTEDNTYYYFTKDPN-----				
SKM1_Scer/1-280	-----GEPPYLN-----	-----EDPLKALYLIANN-----				
CLA4_Scer/1-280	-----GEPPYLN-----	-----EDPLKALYLIATN-----				
STE20_Scer/1-252	-----GEPPYLN-----	-----ETPLRALYLIATN-----				
CDC15_Scer/1-248	-----KNPPYHN-----	-----LTDANIYYAVEN-----				
FMP48_Scer/1-285	-----GDCLFRVSKSK-----	-----KVQHLKNFDEFEKD-----				
KSP1_Scer/1-260	-----HKNPFSIAN-----	-----QSDKSFCYFAANREALFD-----				
CAK1_Scer/1-304	-----ETSRMGHVPAMIDDGS-----	-----DDMNSDGSDFRLICS-----				

Table S4, continued

	541	550	560	570	580	590
TOS3_Scer/1-295	-----	-----	EEMLNGATSGIT	-MEEYTDAKDLLKKLLQKDPDKR		
TPK3_Scer/1-255	-----	-----		-PFFHPDAQDLLKLITRDLSER		
TPK1_Scer/1-255	-----	-----		-PFFNEDVKDLLSRLITRDLQR		
TPK2_Scer/1-255	-----	-----		-PYFHPDVVDLLSKLITADLTRR		
YPK1_Scer/1-256	-----	-----		-DGFDRAKDLLIGLLSRDPTRR		
YPK2_Scer/1-256	-----	-----		-DGFDPAAKDLLIGLLSRDPSSR		
SCH9_Scer/1-260	-----	-----		-DVLSQEGRSFVKGLLNRNPKHR		
YNR047W_Scer/1-282	-----	-----		-NEISRTCKDLIKLLTKNESKR		
RIM15_Scer/1-269	-----	-----		-NEEEEERFLTEAKDLIEKLLVVDPAKR		
IPL1_Scer/1-252	-----	-----		-SNISQDAQDLILKLLKYDPKDR		
PKH2_Scer/1-265	-----	-----		-FPLIIRDLVKKILVKNLDRR		
KCC4_Scer/1-265	-----	-----		-EMPNDTEISRDAQDLIGKILVVDPRQR		
GIN4_Scer/1-271	-----	-----		-EMPSDDEISREAQDLIRKILTVDPERR		
HSL1_Scer/1-289	-----	-----		-QMPSN--LSSEARDLISKILVIDPEKR		
SNF1_Scer/1-252	-----	-----		-TLPKF--LSPGAAGLIKRMILIVNPLNR		
KIN1_Scer/1-279	-----	-----		-EYPQH--LSIEVISLLSKMLVVDPKRR		
KIN4_Scer/1-268	-----	-----		-KFPEY--ITPIPRDLLRRILVPNPRRR		
YPL141C_Scer/1-273	-----	-----		-KFPDY--ILPIPRDLLRRMLVSDPKKR		
CMK1_Scer/1-263	-----	-----		-KFHRPYWDSVSNKAKQFILKALNLDPSKR		
CMK2_Scer/1-263	-----	-----		-TFHMPYWDNISIDVKRFLIKALRLNPADR		
RAD53_Scer/1-269	-----	-----		-HEGPLKDFRISEEARDFIDSLLQVDPNNR		
RCK2_Scer/1-316	-----	-----		-TFLKPWWDEISAGAKNAVAKLLELEPSKR		
APG1_Scer/1-302	-----	-----		-VITFPSYCNIPELKELICSLTFDPAOR		
MEK1_Scer/1-284	-----	-----		-FKLKQWDIVSDNAKSFKV DLLQTDVVKR		
YKL171W_Scer/1-283	-----	-----		-LAQVCKFDRKKNEGFIGQGLYDTVIGCLTIDDK		
CDC5_Scer/1-256	-----	-----		-DFSFPRDKPISDEGKILIRDILSLDPIER		
PSK2_Scer/1-259	-----	-----		-EHVSEECISLIKRILTREVDKR		
YDL025C_Scer/1-274	-----	DIVRG		-PNKILRLLPRHSRTIIGRMLALEPKQR		
SKY1_Scer/1-312	-----	TFFNSRGLLRNISKLKFWPLEDVLT		-KYKFSKDEAKEISDFLSPMLQLDPRKR		
YAK1_Scer/1-257	-----			-PSWMIR--ECLIHFLGGVLNLNPILER		
PTK2_Scer/1-305	-----	PGPGSE		-YSLARNFKNTDATRIAWRLADPNPATR		
PHO85_Scer/1-291	-----	LPKYNPNI-QQR	--PPRDLRQVLQPH	--KEPLDGNLMDFLHGLLQLNPDMR		
CDC28_Scer/1-288	-----	LPDFKPSF-PQW	--RRKDLSQVVP	--SLDPRGIDLLDKLLAYDPINR		
SLT2_Scer/1-296	-----	NVQDY	--IHOLGFIPKVPFVNLYPN	--ANSQALDLLEQMLAFDPQKR		
KSS1_Scer/1-301	-----	RAKEY	--IANLPMRPLPWEtvwskt	--DLNPDMIDLDDKMLQFNPDKR		
FUS3_Scer/1-297	-----	RAREY	--IKSLPMYPAAPPLEKMFPR	--VNPKGIDLLQRMVLVFDPAKR		
HOG1_Scer/1-280	-----	NTLKF	--VTSPLPHRDPIPF SERFKT	--VEPDADV DLL EKMLVFDPKKR		
RIM11_Scer/1-284	-----	YMEH	--KFPQIKPIPLSRVFKK	--EDDQTVEFLADVLKYDPLER		
MRK1_Scer/1-284	-----	YEDH	--VFPNIKIPITLAEIFKA	--EDPDTDLTTKTLKYHPCER		
MCK1_Scer/1-293	-----	YDGPLFSKPLFSGSSQQRFEKYFGH	--SGPDGIDLLMKILVYEPQQR			
CKA1_Scer/1-324	-----	LPREFYD	--MDQYIRKFWHRFIN-DGNKHLSGNDEIIDLIDNLLRYDHQER			
CDC7_Scer/1-272	-----	KPNGYSNG	--LKEFVYDLLNKECTIGTFPE	--HYWCFCVQLEQCFEMDPQKR		
YCK1_Scer/1-261	-----	EKKRSTNVY	--EKKRSTNVY	--DLAQG--LPVQFGRYLEIV		
YCK2_Scer/1-261	-----	EKKRLTNVY	--EKKRLTNVY	--DLAQG--LPIQFGRYLEIV		
YCK3_Scer/1-296	-----	MTKQLNPD	--MTKQLNPD	--DLLNNNAIPYQFATYLKYA		
HRR25_Scer/1-259	-----	EKKLNVSV	--EKKLNVSV	--TLC SG--LPLEFQEYMAYC		
PRR1_Scer/1-317	-----			--WYRLSDYKTNVGKQIVENTLTRK		
MPS1_Scer/1-277	-----			--NNEKIPKS AIELMKACLYRNPDKR		
GCN2_Scer/1-268	-----			--FPPDFDDN--KMKEVKKIIRLLIDHDPNKR		
KIN3_Scer/1-319	-----			--KCDTVPEY--YSRGLNAIIHSMIDVNLTR		
PRK1_Scer/1-277	-----			--QYSDRLKNLIRLMLMEAPSQR		
ARK1_Scer/1-277	-----			--NYSEQLKGLIRDILVQDPRHR		
AKL1_Scer/1-285	-----			--KYSSKLINLIIIMLAENPNLR		
VHS1_Scer/1-321	-----			--ILKQILPLSDDFYSLLSKILQVNPKNR		
SKM1_Scer/1-280	-----			--GSPKLRHPE--SVSKQTKQFLDACLQVNVESR		
CLA4_Scer/1-280	-----			--GTPKLIKHP--SLSLEIKRFLSVCLCVDRYR		
STE20_Scer/1-252	-----			--GTPKLKEPE--NLSSSLKKFLDWCLCVEPEDR		
CDC15_Scer/1-248	-----			--DTYYPPS--SFSEPLKDFLSKCFVKNMYKR		
FMP48_Scer/1-285	-----			--PFAFIYRKYVVPRLSCGYNDEEDLHVSLQHTR		
KSP1_Scer/1-260	-----			--VFSTMAYDFFQVRLRYSLTIDPANRD		
CAK1_Scer/1-304	-----			--IFEKVKQKFINCILGMVSFSPNER		

Table S4, continued

	601	610	620
TOS3_Scer/1-295	IK-----	LADIKVHPFM-----	
TPK3_Scer/1-255	LGNLQNGSEDVKNHPWF-----		
TPK1_Scer/1-255	LGNLQNGTEDVKNHPWF-----		
TPK2_Scer/1-255	IGNLQSGSRDIKAHPWF-----		
YPK1_Scer/1-256	LG---YNGADEIRNHPFF-----		
YPK2_Scer/1-256	LG---VNGBTDEIRNHPFF-----		
SCH9_Scer/1-260	LGA-IDDGRELRAHPFF-----		
YNR047W_Scer/1-282	LGC-KMGAADVKKHPFF-----		
RIM15_Scer/1-269	LG--AKGIQEIKDHPYF-----		
IPL1_Scer/1-252	MR-----	LGDVKMHPWI-----	
PKH2_Scer/1-265	LT-----	ISQIKEHHFF-----	
KCC4_Scer/1-265	IK-----	IRDILSHPLL-----	
GIN4_Scer/1-271	IK-----	TRDILKHPLL-----	
HSL1_Scer/1-289	IT-----	TQEILKHPLI-----	
SNF1_Scer/1-252	IS-----	IHEIMQDDWF-----	
KIN1_Scer/1-279	AT-----	LKVVEHHWM-----	
KIN4_Scer/1-268	IN-----	LOTIKRHVWL-----	
YPL141C_Scer/1-273	IN-----	LKQIKKHEWL-----	
CMK1_Scer/1-263	PT-----	AAELLEDPWI-----	
CMK2_Scer/1-263	PT-----	ATELLDDPWI-----	
RAD53_Scer/1-269	ST-----	AAKALNHPWI-----	
RCK2_Scer/1-316	YD-----	IDQFLDDPWL-----	
APG1_Scer/1-302	IG-----	FEEFFANKVV-----	
MEK1_Scer/1-284	LN-----	SKQGLKHIWI-----	
YKL171W_Scer/1-283	RWK-----	LKRRIEVL-----	
CDC5_Scer/1-256	PS-----	LTEIMDYVWF-----	
PSK2_Scer/1-259	PT-----	IDEIYEDKWL-----	
YDL025C_Scer/1-274	VL-----	MNDVVKDDWL-----	
SKY1_Scer/1-312	AD-----	AGGLVNHPWL-----	
YAK1_Scer/1-257	WT-----	PQQAMLHPFI-----	
PTK2_Scer/1-305	YT-----	MDDLFNDPFF-----	
PHO85_Scer/1-291	LS-----	AKQALHHPWF-----	
CDC28_Scer/1-288	IS-----	ARRAAIH PYF-----	
SLT2_Scer/1-296	IT-----	VDEALEHPYL-----	
KSS1_Scer/1-301	IS-----	AAEALRHPYL-----	
FUS3_Scer/1-297	IT-----	AKEALEHPYL-----	
HOG1_Scer/1-280	IT-----	AADALAH PYS-----	
RIM11_Scer/1-284	FN-----	ALQCLCSPYF-----	
MRK1_Scer/1-284	LV-----	PLQCLLSSYF-----	
MCK1_Scer/1-293	LS-----	PRRILAHQFF-----	
CKA1_Scer/1-324	LT-----	AKEAMGHPWF-----	
CDC7_Scer/1-272	SS-----	AEDLLKTPFF-----	
YCK1_Scer/1-261	RS-----	LSFEECPDYEGYRKLLL	
YCK2_Scer/1-261	RN-----	LSFEETPDYEGYRMILL	
YCK3_Scer/1-296	RS-----	LKFDEDPDYD----YLI	
HRR25_Scer/1-259	KN-----	LKFDEKPDYLFLARLFK	
PRR1_Scer/1-317	NQR--	WSINEIYESPV-----	
MPS1_Scer/1-277	WT-----	VDKVLSSSTFL-----	
GCN2_Scer/1-268	PG-----	ARTLLNSGWL-----	
KIN3_Scer/1-319	PS-----	TFELLQDIQI-----	
PRK1_Scer/1-277	PN-----	ICQVLEEVSR-----	
ARK1_Scer/1-277	PN-----	VYQLLKRI-----	
AKL1_Scer/1-285	PN-----	IYQVLYHLCE-----	
VHS1_Scer/1-321	MS-----	LQELMKEVSS-----	
SKM1_Scer/1-280	AS-----	VRKLLTFEFL-----	
CLA4_Scer/1-280	AS-----	TEELLHHGFF-----	
STE20_Scer/1-252	AS-----	ATELLHDEYI-----	
CDC15_Scer/1-248	PT-----	ADQLLKHVWI-----	
FMP48_Scer/1-285	QY-----	IWQDLPDIYD-----	
KSP1_Scer/1-260	LK--	MMRTELQNLSEYT-----	
CAK1_Scer/1-304	WS-----	CQRILQELE-----	

## Supplemental References

1. D. M. Gelperin, M. A. White, M. L. Wilkinson, Y. Kon, L. A. Kung, K. J. Wise, N. Lopez-Hoyo, L. Jiang, S. Piccirillo, H. Yu, M. Gerstein, M. E. Dumont, E. M. Phizicky, M. Snyder, E. J. Grayhack, Biochemical and genetic analysis of the yeast proteome with a movable ORF collection. *Genes Dev.* **19**, 2816-2826 (2005).
2. R. D. Gietz, R. H. Schiestl, A. R. Willems, R. A. Woods, Studies on the transformation of intact yeast cells by the LiAc/SS-DNA/PEG procedure. *Yeast* **11**, 355-360 (1995).
3. S. Ghaemmaghami, W. K. Huh, K. Bower, R. W. Howson, A. Belle, N. Dephoure, E. K. O'Shea, J. S. Weissman, Global analysis of protein expression in yeast. *Nature* **425**, 737-741 (2003).
4. E. A. Winzeler, D. D. Shoemaker, A. Astromoff, H. Liang, K. Anderson, B. Andre, R. Bangham, R. Benito, J. D. Boeke, H. Bussey, A. M. Chu, C. Connelly, K. Davis, F. Dietrich, S. W. Dow, M. El Bakkoury, F. Foury, S. H. Friend, E. Gentalen, G. Giaever, J. H. Hegemann, T. Jones, M. Laub, H. Liao, N. Liebundguth, D. J. Lockhart, A. Lucau-Danila, M. Lussier, N. M'Rabet, P. Menard, M. Mittmann, C. Pai, C. Rebischung, J. L. Revuelta, L. Riles, C. J. Roberts, P. Ross-MacDonald, B. Scherens, M. Snyder, S. Sookhai-Mahadeo, R. K. Storms, S. Veronneau, M. Voet, G. Volckaert, T. R. Ward, R. Wysocki, G. S. Yen, K. Yu, K. Zimmermann, P. Philippsen, M. Johnston, R. W. Davis, Functional characterization of the *S. cerevisiae* genome by gene deletion and parallel analysis. *Science* **285**, 901-906 (1999).
5. W. P. Voth, Y. W. Jiang, D. J. Stillman, New 'marker swap' plasmids for converting selectable markers on budding yeast gene disruptions and plasmids. *Yeast* **20**, 985-993 (2003).
6. H. Zhu, M. Bilgin, R. Bangham, D. Hall, A. Casamayor, P. Bertone, N. Lan, R. Jansen, S. Bidlingmaier, T. Houfek, T. Mitchell, P. Miller, R. A. Dean, M. Gerstein, M. Snyder, Global analysis of protein activities using proteome chips. *Science* **293**, 2101-2105 (2001).

7. M. Costanzo, O. Schub, B. Andrews, G1 transcription factors are differentially regulated in *Saccharomyces cerevisiae* by the Swi6-binding protein Stb1. *Mol. Cell Biol.* **23**, 5064-5077 (2003).
8. Y. J. Sheu, B. Stillman, Cdc7-Dbf4 phosphorylates MCM proteins via a docking site-mediated mechanism to promote S phase progression. *Mol. Cell* **24**, 101-113 (2006).
9. R. P. Bhattacharyya, A. Remenyi, M. C. Good, C. J. Bashor, A. M. Falick, W. A. Lim, The Ste5 scaffold allosterically modulates signaling output of the yeast mating pathway. *Science* **311**, 822-826 (2006).
10. E. Bilsland-Marchesan, J. Arino, H. Saito, P. Sunnerhagen, F. Posas, Rck2 kinase is a substrate for the osmotic stress-activated mitogen-activated protein kinase Hog1. *Mol. Cell. Biol.* **20**, 3887-3895 (2000).
11. I. M. Cheeseman, S. Anderson, M. Jwa, E. M. Green, J. Kang, J. R. Yates, 3rd, C. S. Chan, D. G. Drubin, G. Barnes, Phospho-regulation of kinetochore-microtubule attachments by the Aurora kinase Ipl1p. *Cell* **111**, 163-172 (2002).
12. L. Wan, T. de los Santos, C. Zhang, K. Shokat, N. M. Hollingsworth, Mek1 kinase activity functions downstream of RED1 in the regulation of meiotic double strand break repair in budding yeast. *Mol. Biol. Cell* **15**, 11-23 (2004).
13. R. Sopko, D. Huang, J. C. Smith, D. Figeys, B. J. Andrews, Activation of the Cdc42p GTPase by cyclin-dependent protein kinases in budding yeast. *EMBO J.* **26**, 4487-4500 (2007).
14. J. L. Ma, S. J. Lee, J. K. Duong, D. F. Stern, Activation of the checkpoint kinase Rad53 by the phosphatidyl inositol kinase-like kinase Mec1. *J. Biol. Chem.* **281**, 3954-3963 (2006).
15. A. Reinders, N. Burckert, T. Boller, A. Wiemken, C. De Virgilio, *Saccharomyces cerevisiae* cAMP-dependent protein kinase controls entry into stationary phase through the Rim15p protein kinase. *Genes Dev.* **12**, 2943-2955 (1998).
16. D. J. Robbins, E. Zhen, H. Owaki, C. A. Vanderbilt, D. Ebert, T. D. Geppert, M. H. Cobb, Regulation and properties of extracellular signal-regulated protein kinases 1 and 2 in vitro. *J. Biol. Chem.* **268**, 5097-5106 (1993).

17. A. Devault, E. Gueydon, E. Schwob, Interplay between S-cyclin-dependent kinase and Dbf4-dependent kinase in controlling DNA replication through phosphorylation of yeast Mcm4 N-terminal domain. *Mol. Biol. Cell* **19**, 2267-2277 (2008).
18. J. Ptacek, G. Devgan, G. Michaud, H. Zhu, X. Zhu, J. Fasolo, H. Guo, G. Jona, A. Breitkreutz, R. Sopko, R. R. McCartney, M. C. Schmidt, N. Rachidi, S. J. Lee, A. S. Mah, L. Meng, M. J. Stark, D. F. Stern, C. De Virgilio, M. Tyers, B. Andrews, M. Gerstein, B. Schweitzer, P. F. Predki, M. Snyder, Global analysis of protein phosphorylation in yeast. *Nature* **438**, 679-684 (2005).
19. M. Rodriguez, S. S. Li, J. W. Harper, Z. Songyang, An oriented peptide array library (OPAL) strategy to study protein-protein interactions. *J. Biol. Chem.* **279**, 8802-8807 (2004).
20. Z. Songyang, S. Blechner, N. Hoagland, M. F. Hoekstra, H. Piwnica-Worms, L. C. Cantley, Use of an oriented peptide library to determine the optimal substrates of protein kinases. *Curr. Biol.* **4**, 973-982 (1994).
21. E. F. Johnson, K. D. Stewart, K. W. Woods, V. L. Giranda, Y. Luo, Pharmacological and functional comparison of the polo-like kinase family: insight into inhibitor and substrate specificity. *Biochemistry* **46**, 9551-9563 (2007).
22. Z. Songyang, K. P. Lu, Y. T. Kwon, L. H. Tsai, O. Filhol, C. Cochet, D. A. Brickey, T. R. Soderling, C. Bartleson, D. J. Graves, A. J. DeMaggio, M. F. Hoekstra, J. Blenis, T. Hunter, L. C. Cantley, A structural basis for substrate specificities of protein Ser/Thr kinases: primary sequence preference of casein kinases I and II, NIMA, phosphorylase kinase, calmodulin-dependent kinase II, CDK5, and Erk1. *Mol. Cell. Biol.* **16**, 6486-6493 (1996).
23. U. E. Rennefahrt, S. W. Deacon, S. A. Parker, K. Devarajan, A. Beeser, J. Chernoff, S. Knapp, B. E. Turk, J. R. Peterson, Specificity profiling of Pak kinases allows identification of novel phosphorylation sites. *J. Biol. Chem.* **282**, 15667-15678 (2007).
24. R. R. White, Y. G. Kwon, M. Taing, D. S. Lawrence, A. M. Edelman, Definition of optimal substrate recognition motifs of Ca<sup>2+</sup>-calmodulin-dependent protein kinases IV and II reveals shared and distinctive features. *J. Biol. Chem.* **273**, 3166-3172 (1998).

- 25.I. A. Manke, A. Nguyen, D. Lim, M. Q. Stewart, A. E. Elia, M. B. Yaffe, MAPKAP kinase-2 is a cell cycle checkpoint kinase that regulates the G2/M transition and S phase progression in response to UV irradiation. *Mol. Cell* **17**, 37-48 (2005).
- 26.J. A. Gonzalez-Vera, E. Lukovic, B. Imperiali, A rapid method for generation of selective Sox-based chemosensors of Ser/Thr kinases using combinatorial peptide libraries. *Bioorg. Med. Chem. Lett.* **19**, 1258-1260 (2009).
- 27.J. M. Lizcano, M. Deak, N. Morrice, A. Kieloch, C. J. Hastie, L. Dong, M. Schutkowski, U. Reimer, D. R. Alessi, Molecular basis for the substrate specificity of NIMA-related kinase-6 (NEK6). Evidence that NEK6 does not phosphorylate the hydrophobic motif of ribosomal S6 protein kinase and serum- and glucocorticoid-induced protein kinase in vivo. *J. Biol. Chem.* **277**, 27839-27849 (2002).
- 28.C. J. Fiol, J. H. Haseman, Y. H. Wang, P. J. Roach, R. W. Roeske, M. Kowalcuk, A. A. DePaoli-Roach, Phosphoserine as a recognition determinant for glycogen synthase kinase-3: phosphorylation of a synthetic peptide based on the G-component of protein phosphatase-1. *Arch. Biochem. Biophys.* **267**, 797-802 (1988).
- 29.T. Obata, M. B. Yaffe, G. G. Leparc, E. T. Piro, H. Maegawa, A. Kashiwagi, R. Kikkawa, L. C. Cantley, Peptide and protein library screening defines optimal substrate motifs for AKT/PKB. *J. Biol. Chem.* **275**, 36108-36115 (2000).
- 30.D. M. Gwinn, D. B. Shackelford, D. F. Egan, M. M. Mihaylova, A. Mery, D. S. Vasquez, B. E. Turk, R. J. Shaw, AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol. Cell* **30**, 214-226 (2008).
- 31.M. L. Miller, L. J. Jensen, F. Diella, C. Jorgensen, M. Tinti, L. Li, M. Hsiung, S. A. Parker, J. Bordeaux, T. Sicheritz-Ponten, M. Olhovsky, A. Pascalescu, J. Alexander, S. Knapp, N. Blom, P. Bork, S. Li, G. Cesareni, T. Pawson, B. E. Turk, M. B. Yaffe, S. Brunak, R. Linding, Linear motif atlas for phosphorylation-dependent signaling. *Sci. Signal.* **1**, ra2 (2008).
- 32.S. Himpel, W. Tegge, R. Frank, S. Leder, H. G. Joost, W. Becker, Specificity determinants of substrate recognition by the protein kinase DYRK1A. *J. Biol. Chem.* **275**, 2431-2438 (2000).